

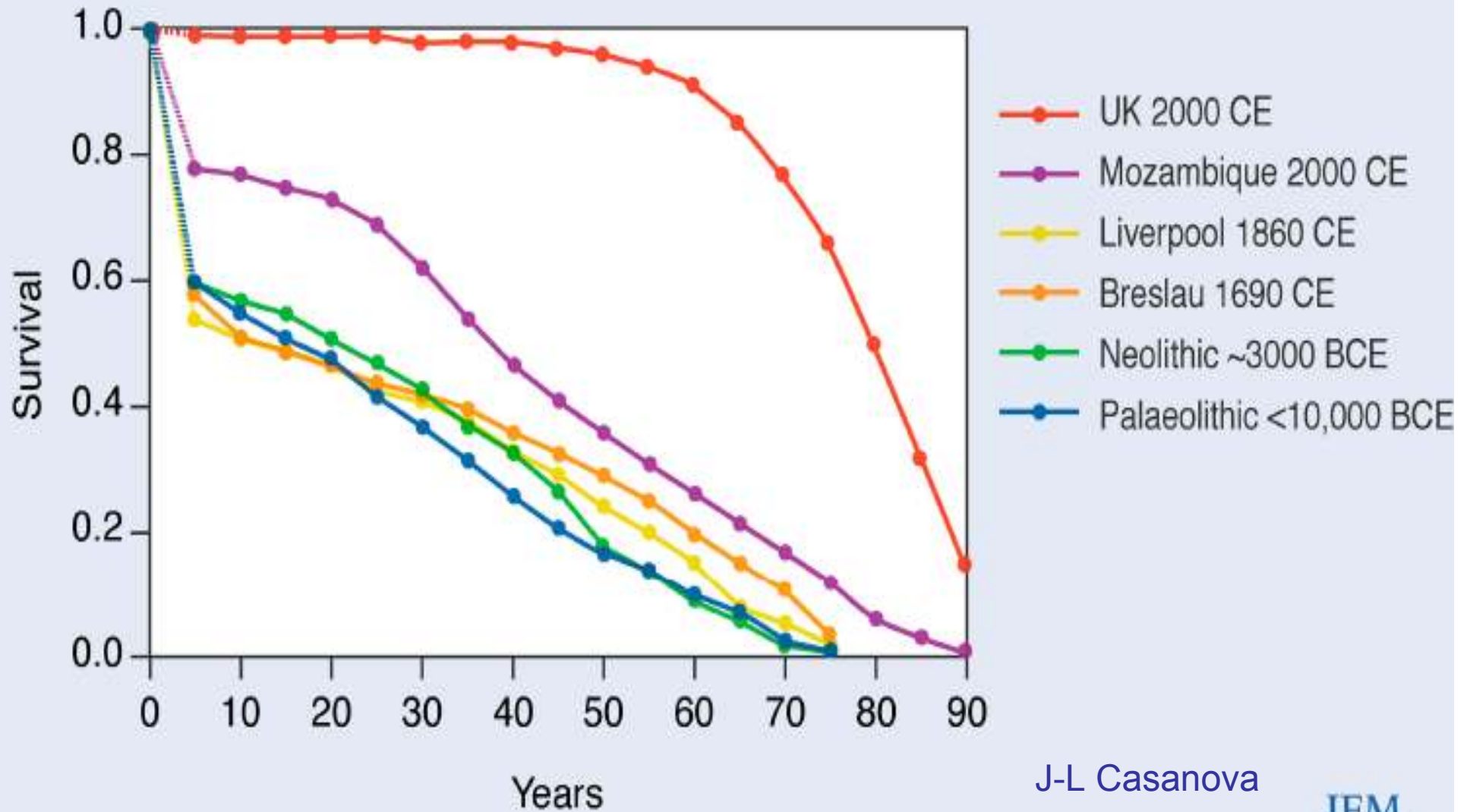
# T cell differentiation, migration and immune regulation

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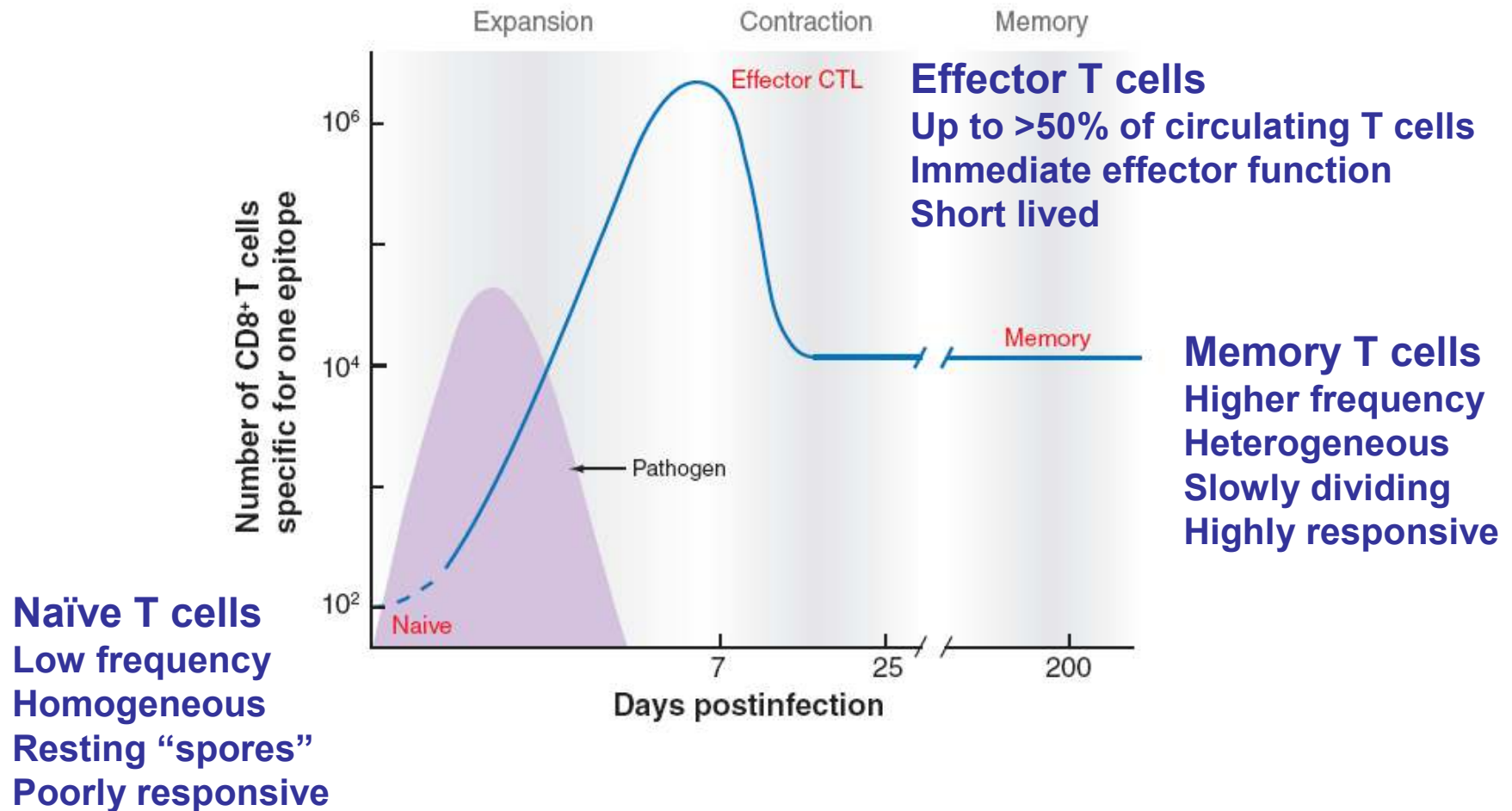
# Increased life expectancy and medical progress



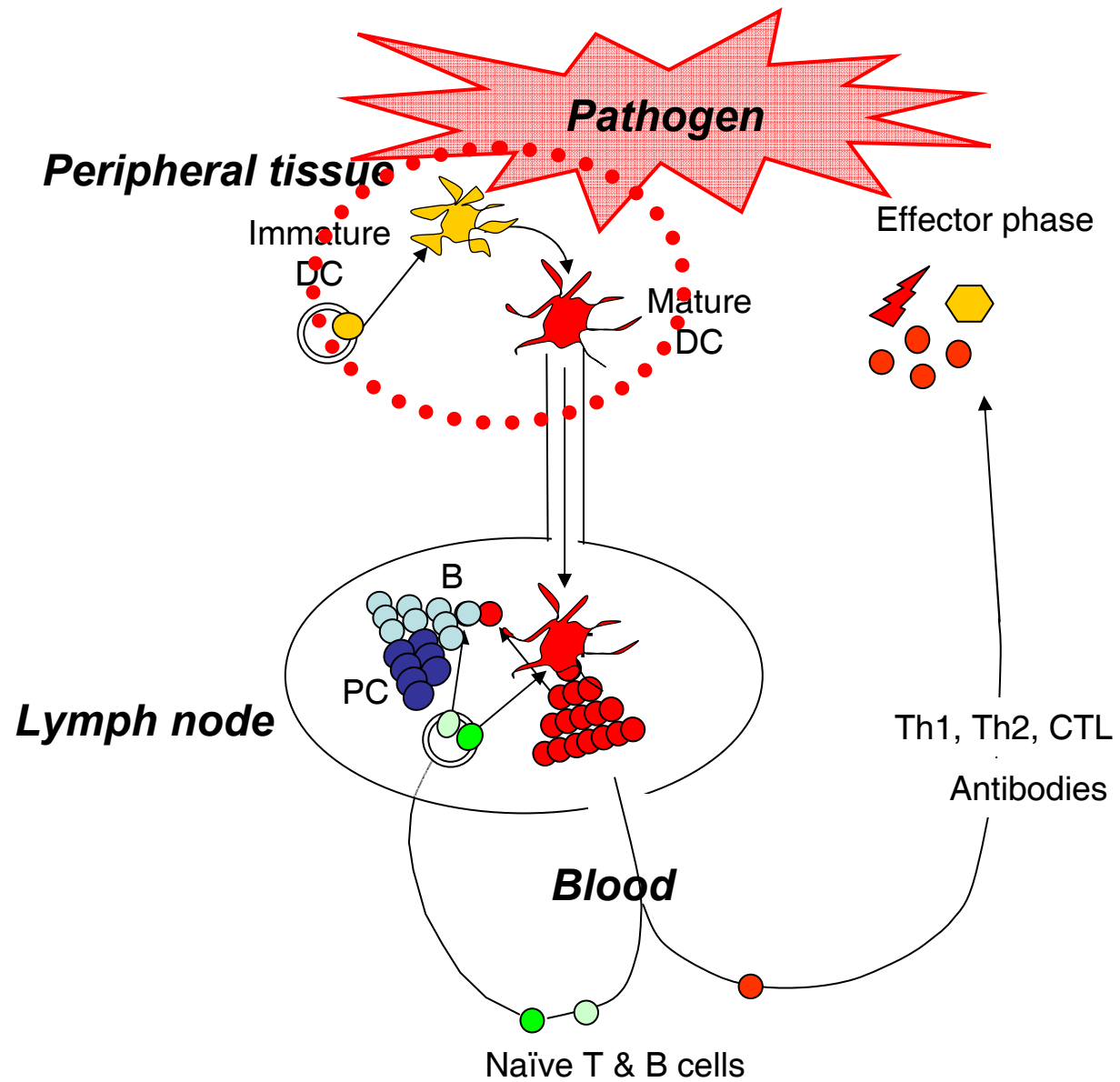
J-L Casanova

JEM

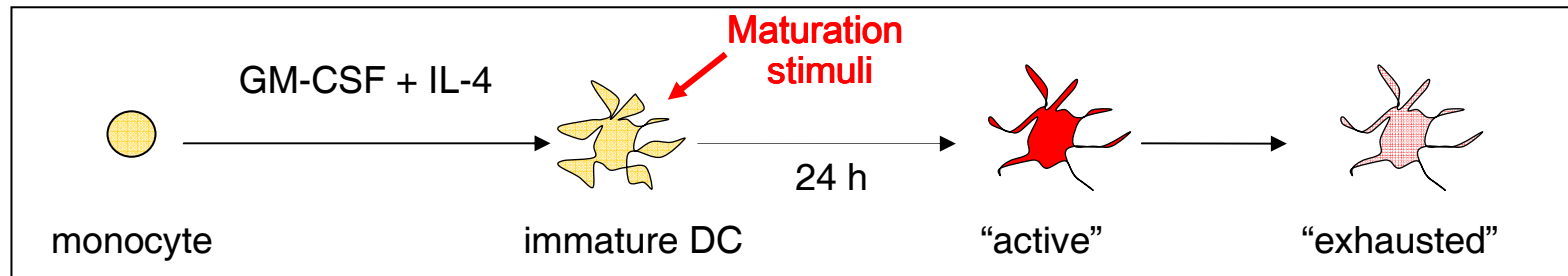
# From naïve to memory T cells



# Cell migration in the immune response



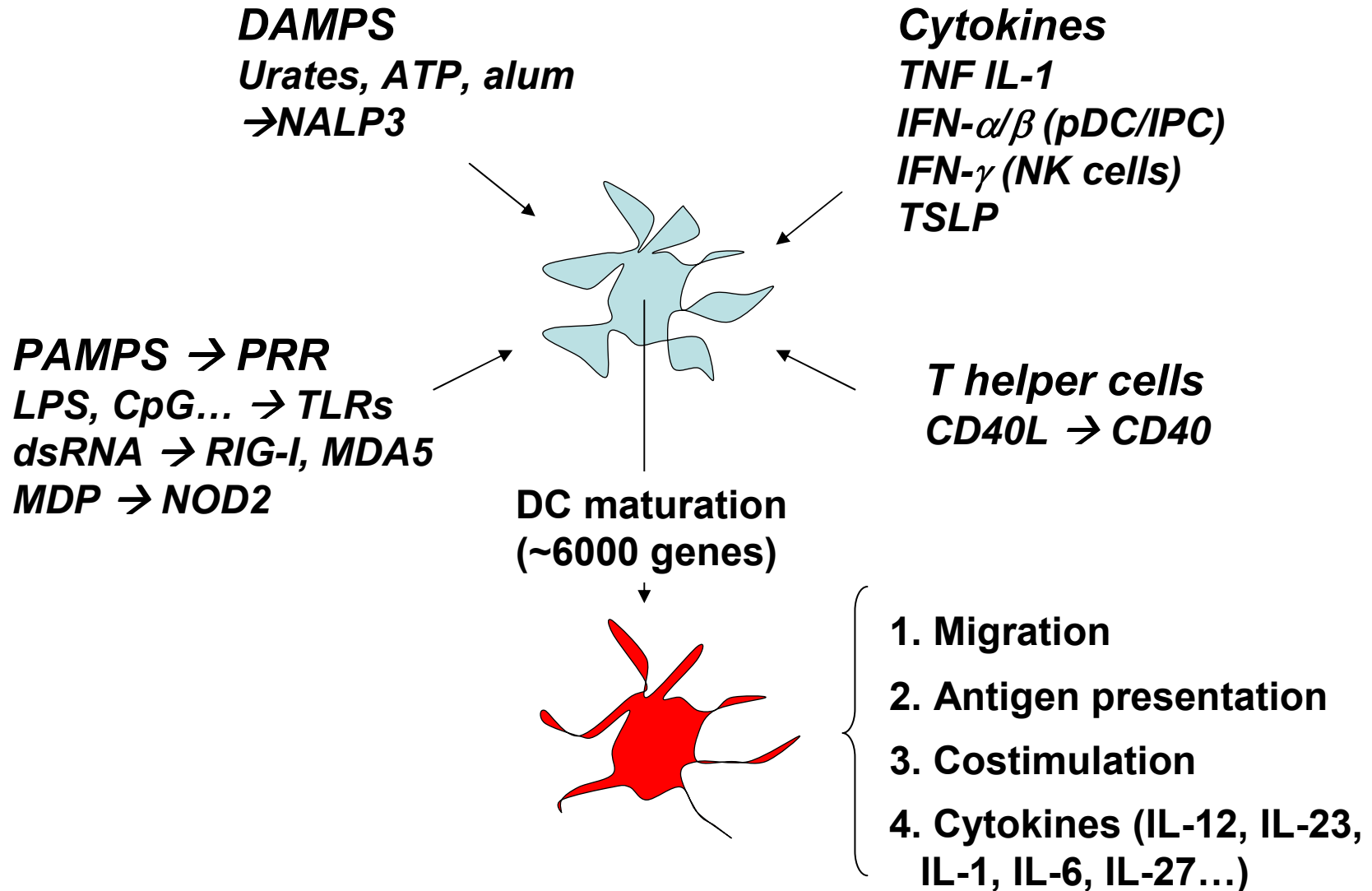
# Dendritic cell maturation in vitro



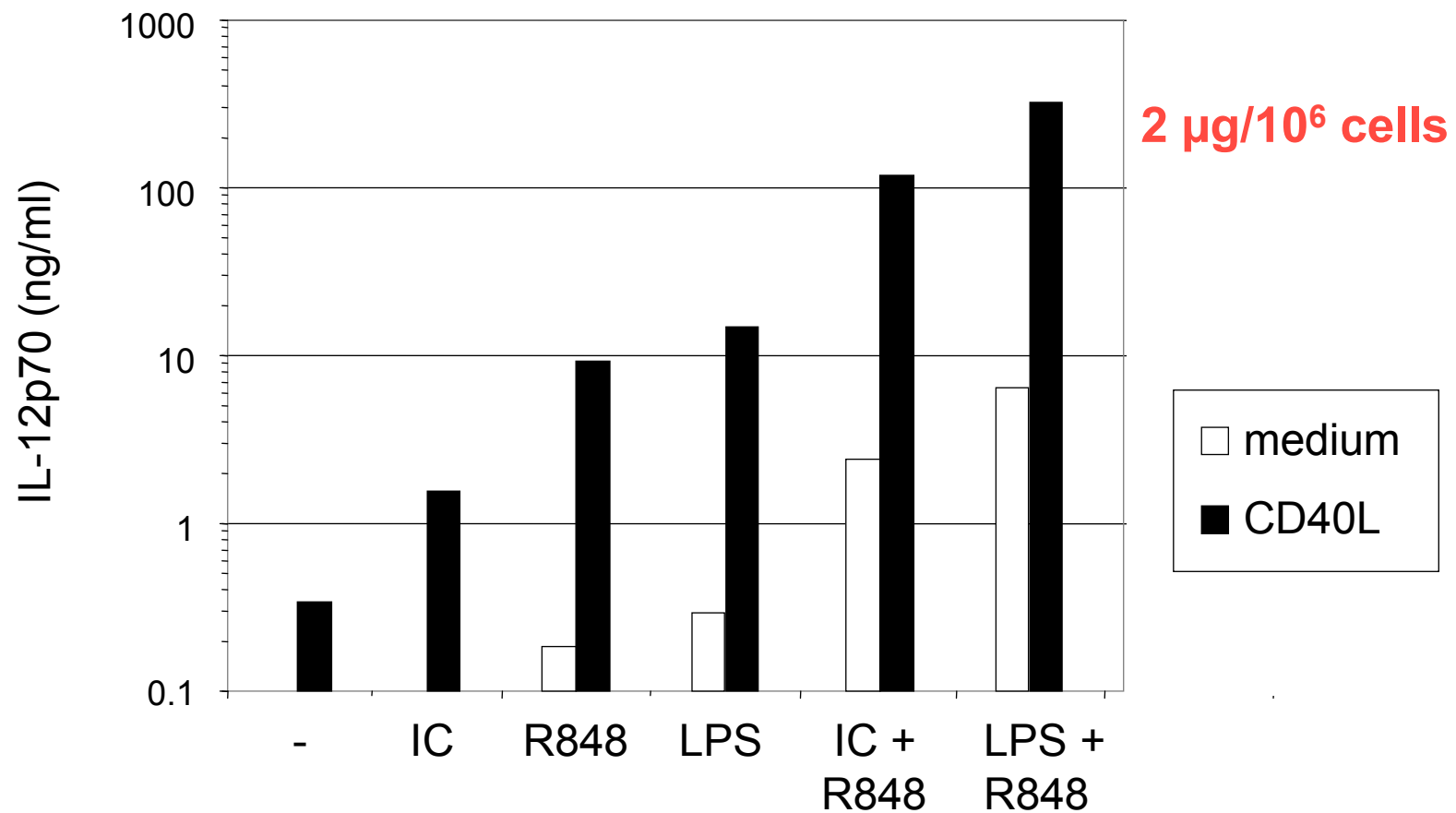
Antigen capture <sup>1</sup>	Macropinocytosis	++	+	-
	Mannose R	++	+	-
Antigen presentation <sup>2</sup>	MHC II synthesis	+	++	-
	MHC II halflife	10 h		>100 h
	MHC I synthesis	+	++	++
Costimulation <sup>1</sup>	B7	-	+	+
Migration <sup>4</sup>	CCR5	+	(+)	-
	CCR7	-	+	++
Cytokines <sup>5</sup>	TNF IL-6 IL-1	-	+	-
	IFN-I	-	+	-
	IL-12	-	+/-	-

1) Sallusto et al *JEM* 1994; *JEM* 1995; 2) Cella et al *Nature* 1997; 3) Cella et al *JEM* 1999; 4) Sallusto et al *EJI* 1998; 5) Langenkamp et al *Nat Immunol* 2000

# DC activation: integration of multiple stimuli

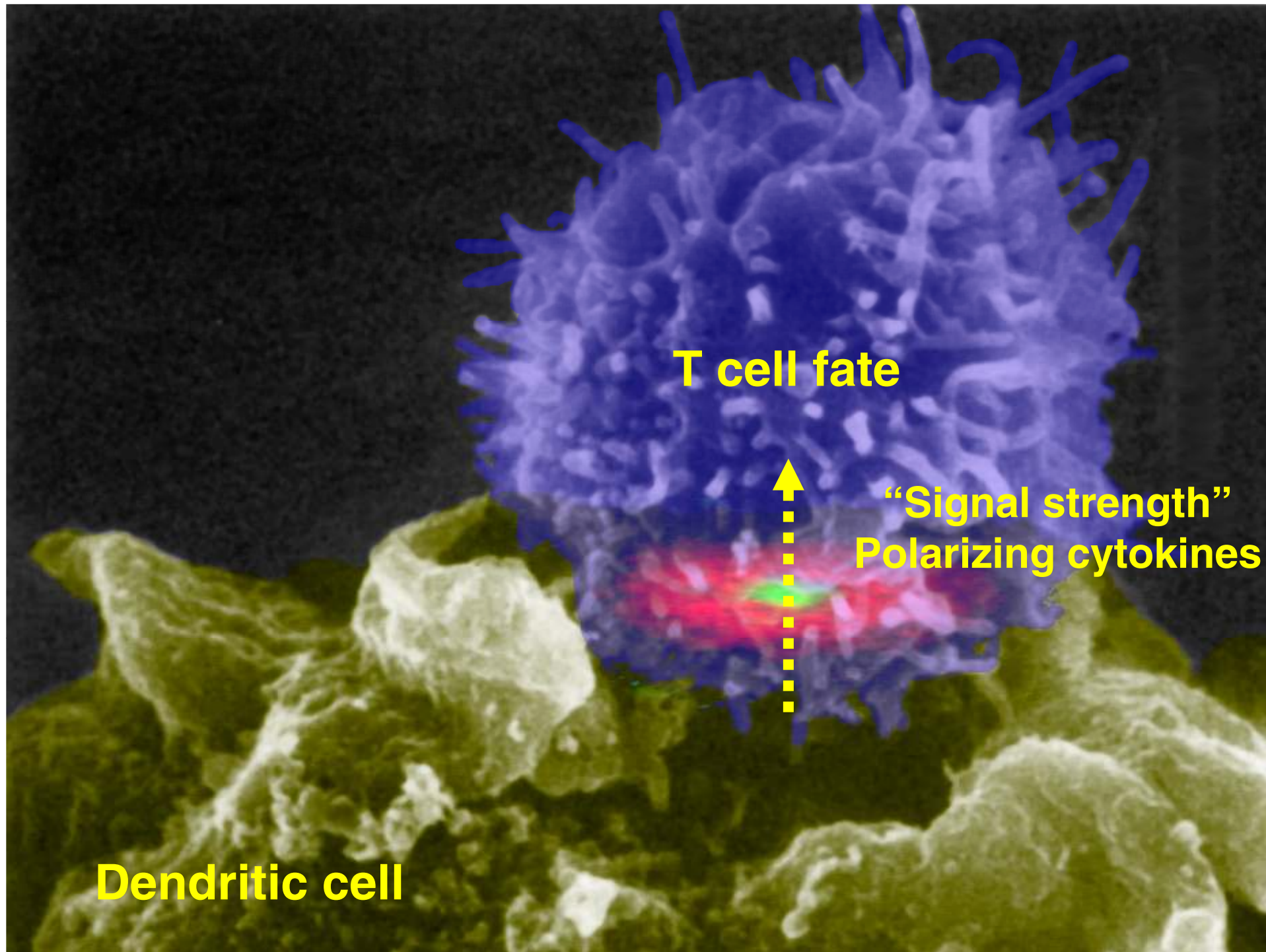


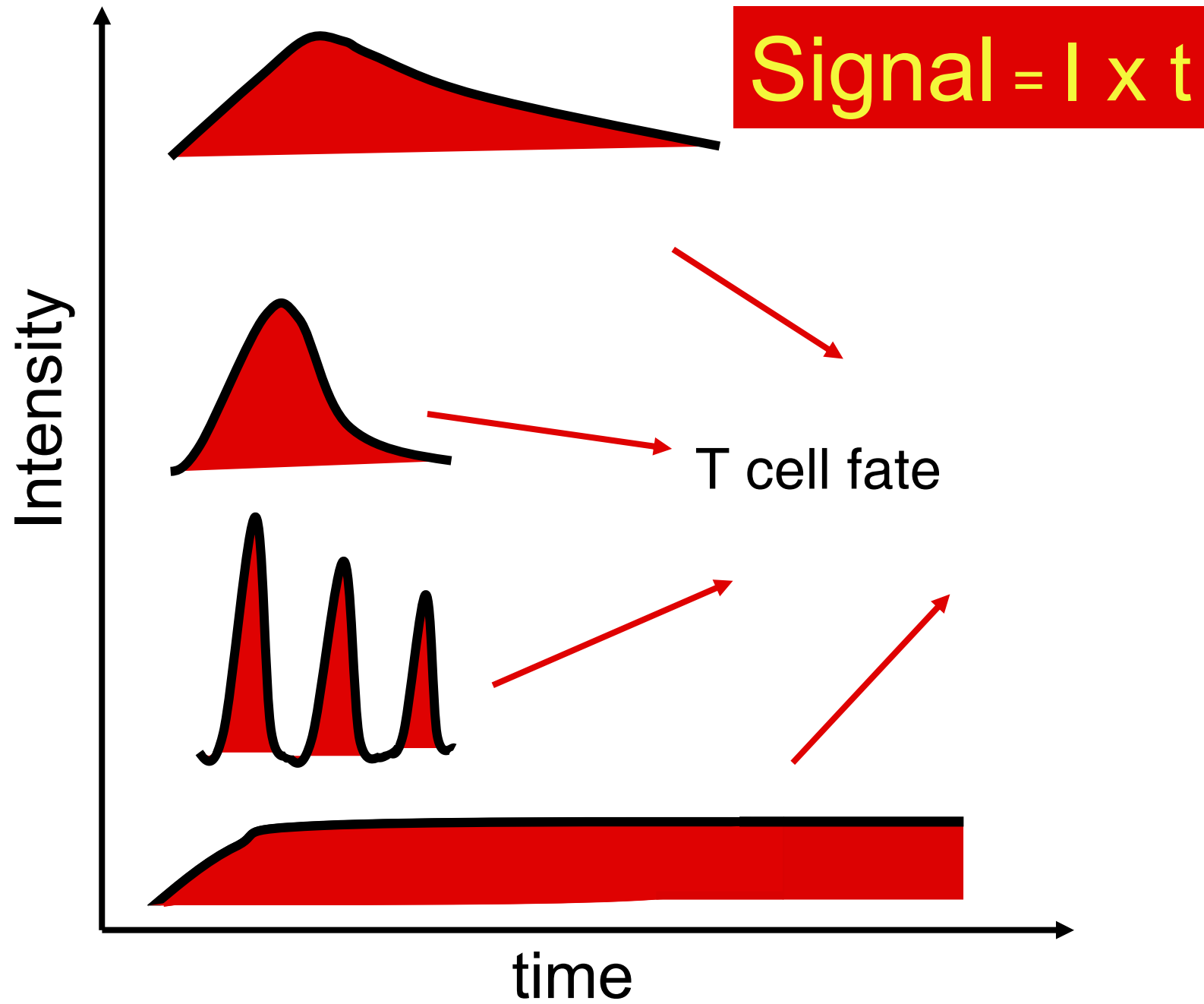
# IL-12p70 production is triggered by synergic TLR stimulation and is further boosted by CD40L



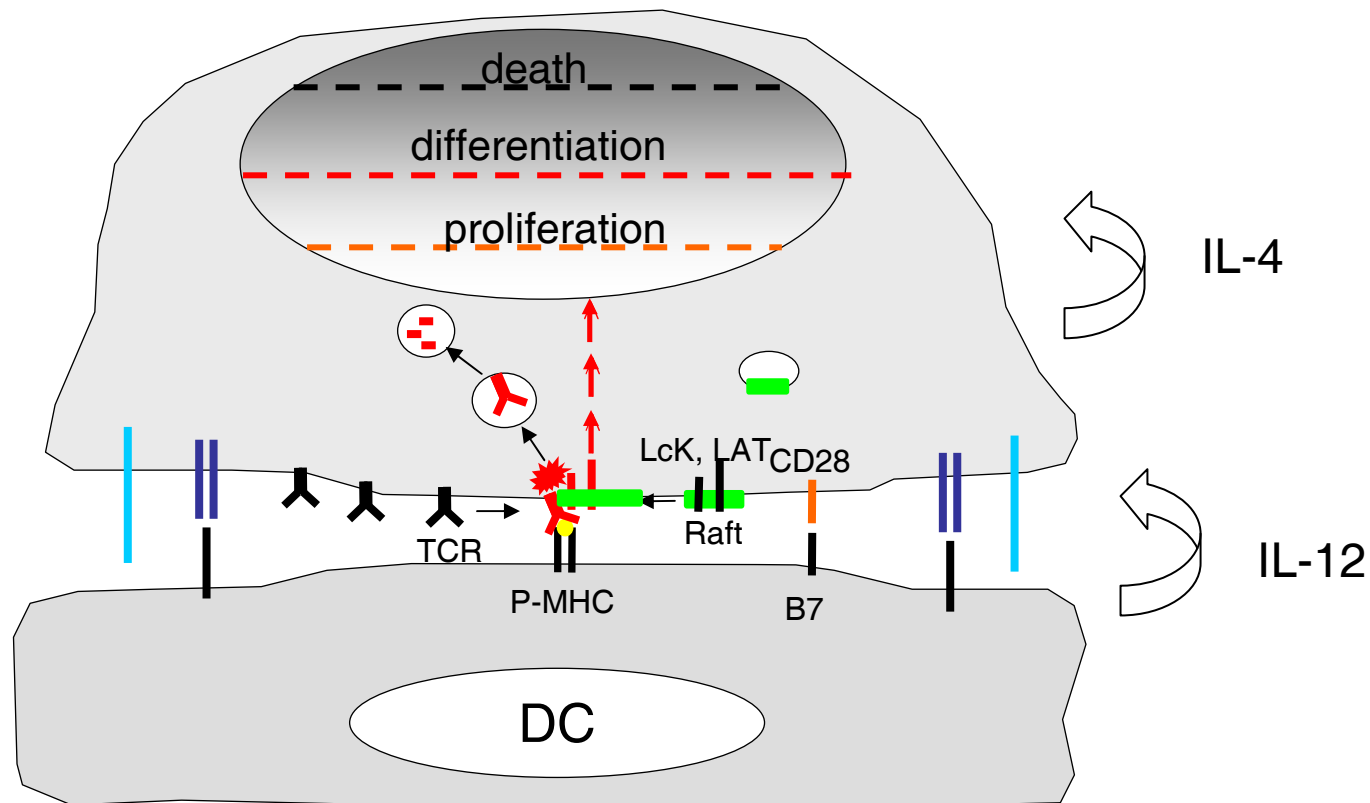
- 1. T cell activation and fate determination**
- 2. Human effector/memory T cell subsets  
( $T_{CM}$ ,  $T_{EM}$ , Th1, Th2, Th17, etc)**
- 3. T cell traffic in steady state and inflammatory conditions (mouse)**





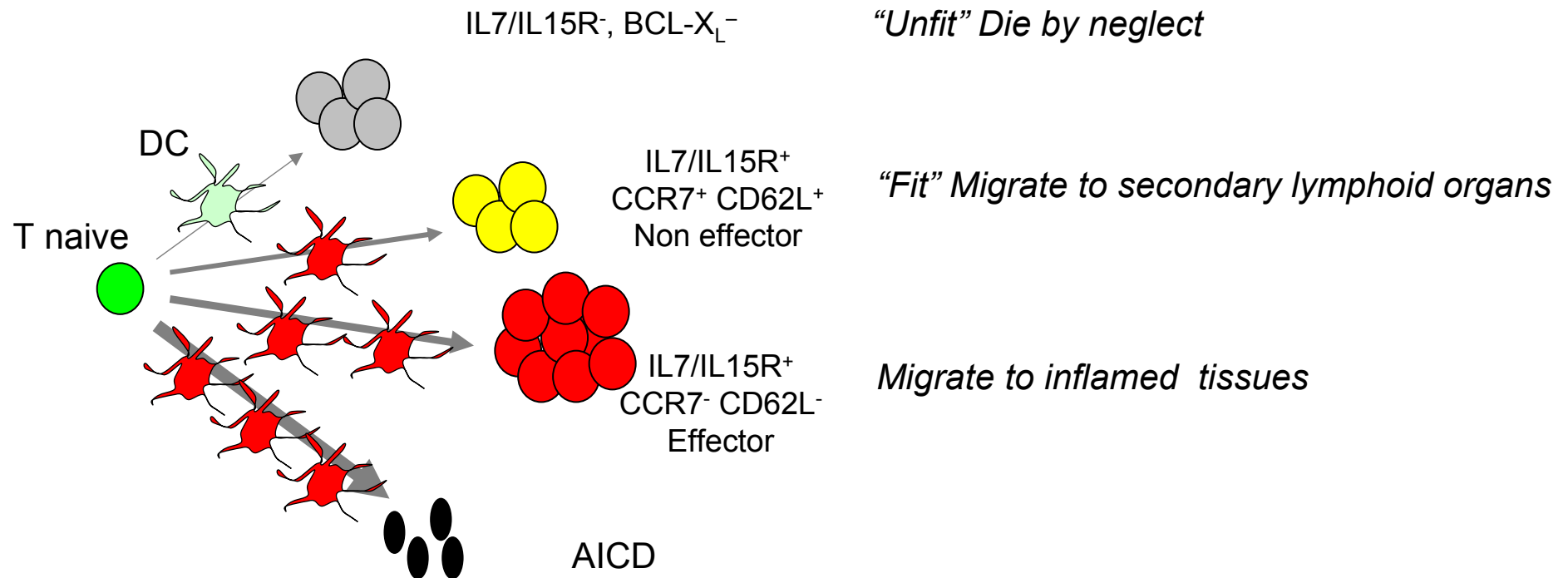


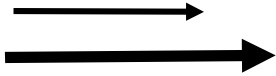
# Signal strength and activation thresholds



1. [peptide-MHC]      Rate of TCR triggering
2. [B7]              Amplification
3. Stability of the synapse      Duration of signalling
4. [Polarizing cytokines]      Differentiation

# The strength of stimulation determines T cell fate

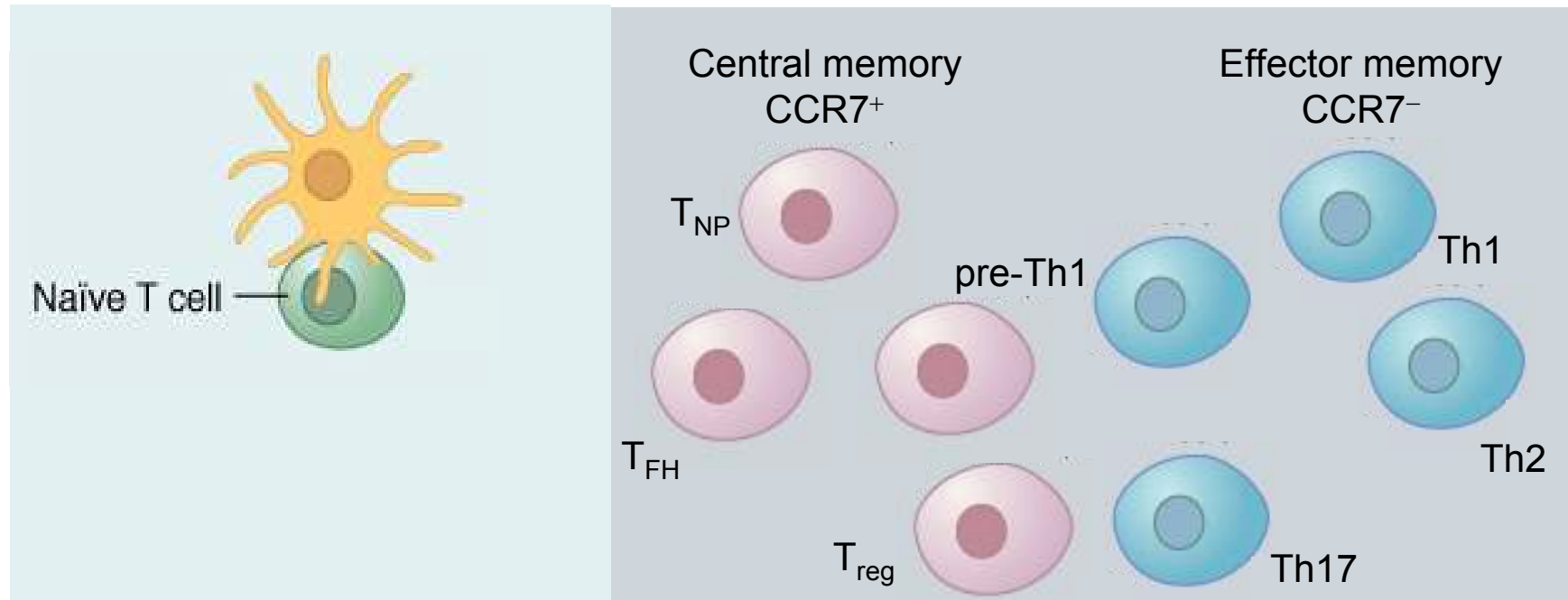


  
 Strength / duration of stimulation  
 DC maturation and density, antigen concentration,  
 stochastic T-DC interactions and T-T competition  
 Polarizing cytokines

lezzi et al, *Immunity* 1998  
 lezzi et al, *JEM* 2000  
 Langenkamp et al, *Nat Immunol* 2000  
 Langenkamp et al, *EJI* 2002  
 Gett et al, *Nat Immunol* 2003  
 Messi et al, *Nat Immunol* 2003

1. T cell activation and fate determination
2. Human effector/memory T cell subsets  
( $T_{CM}$ ,  $T_{EM}$ , Th1, Th2, Th17, etc)
3. T cell traffic in steady state and inflammatory conditions (mouse)

# Dynamics of T lymphocyte responses: intermediates, effectors and memory cells



- *Strength / duration of stimulation*
- *T-DC serial encounters*
- *Polarizing cytokines*
- *Asymmetric division*

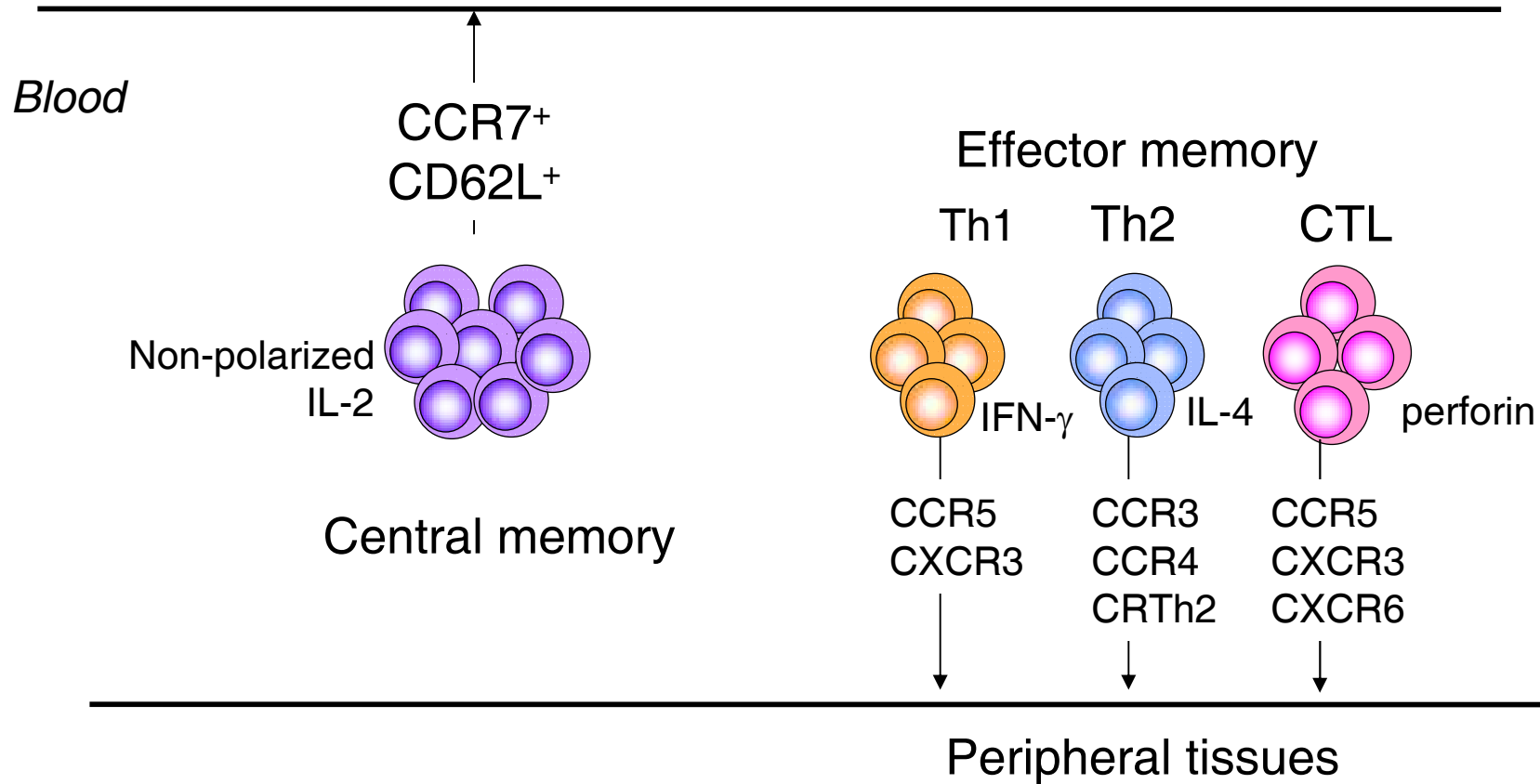
- *Migratory capacity*
- *Effector function*
- *Differentiation potential*
- *Survival*

# Two subsets of memory T cells with distinct migratory capacity and effector function

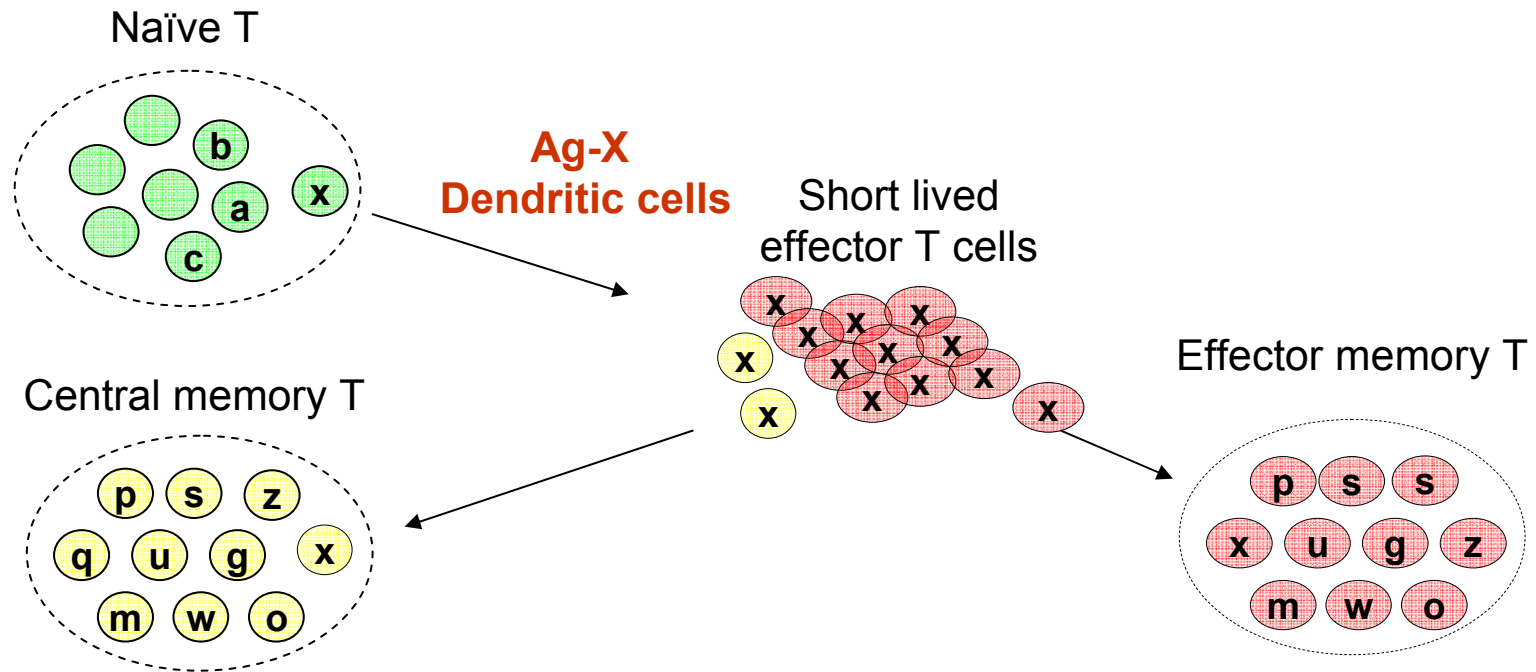
Sallusto et al. *Nature* 401: 708 (1999)

T cell areas of secondary lymphoid organs

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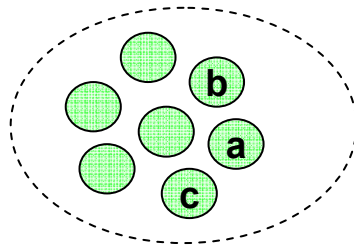
# T cell clonal dynamics: primary responses



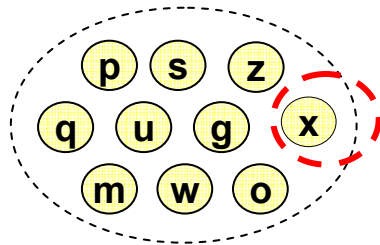


# T cell clonal dynamics: memory phase

Naïve T

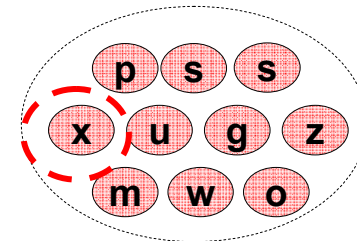


Central memory T



Secondary response

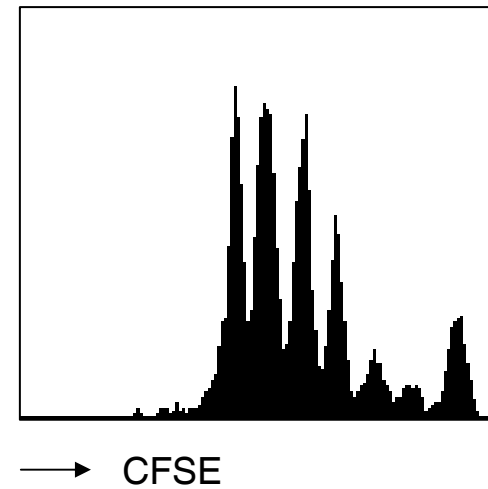
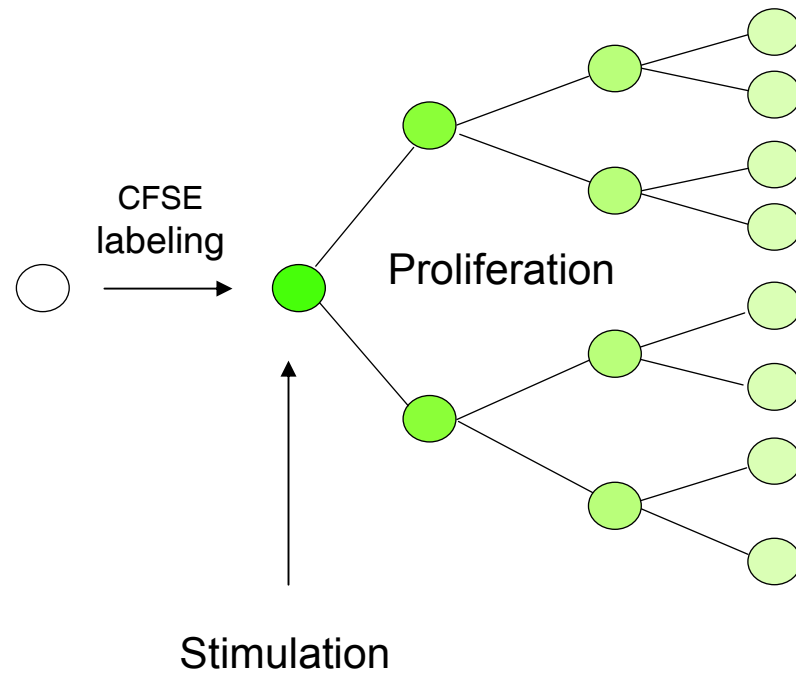
Effector memory T



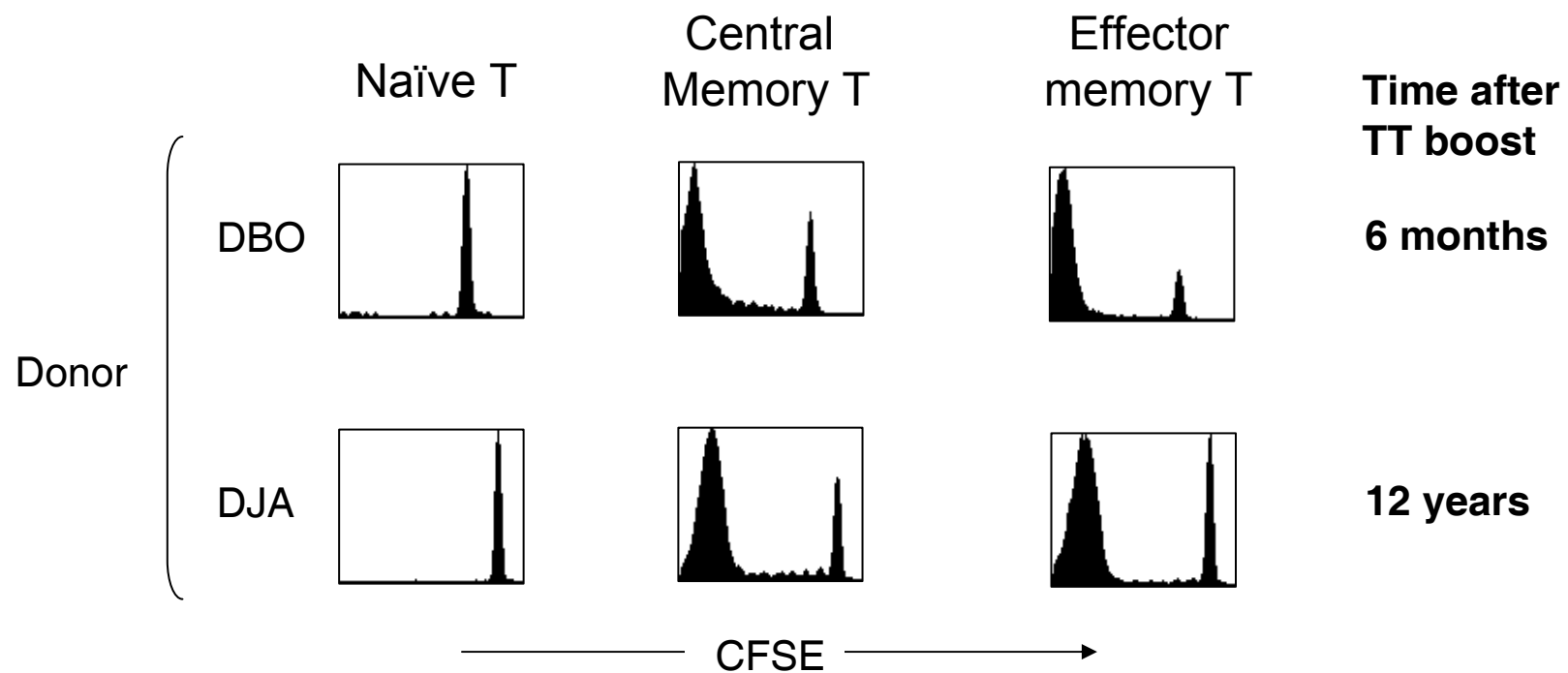
Immediate protection

**Central and effector memory T cells are stable years**

# Tracking T cell division by CFSE dilution (Lyons & Parish)

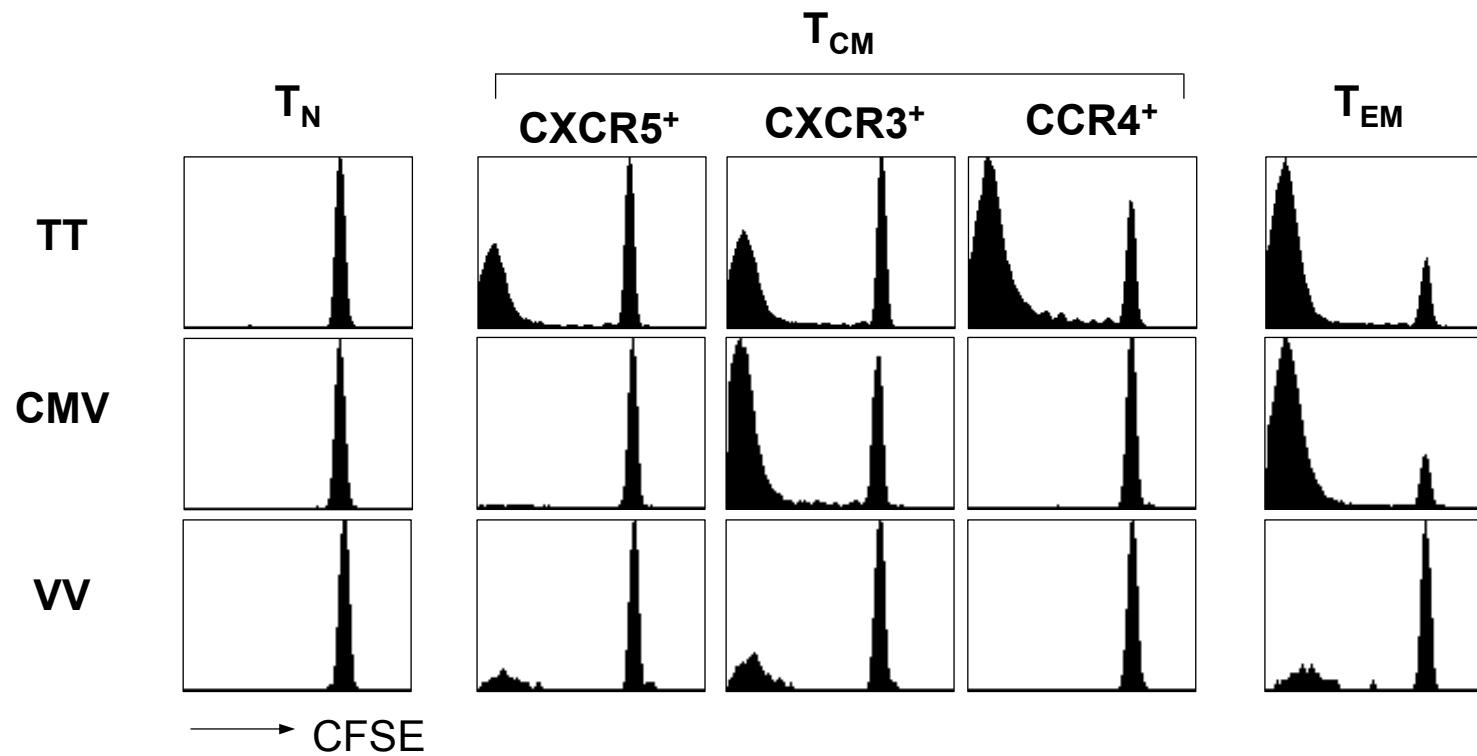


# Stability of central memory and effector memory T cells



CFSE-labeled T cells stimulated by antigen-loaded autologous monocytes

# Antigen-specific T cells are differentially distributed in memory subsets



## Division of labor among memory T cells

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### Central memory T ( $T_{CM}$ )

$T_{CM}$  circulate through secondary lymphoid tissues

Secrete IL-2 but little IFN- $\gamma$  and no perforin

Robust proliferation capacity and transition into effector cells that can migrate to non-lymphoid tissue

Differentiation intermediates

*Secondary responses*

### Effector memory T ( $T_{EM}$ )

$T_{EM}$  circulate through and can reside in non-lymphoid tissue

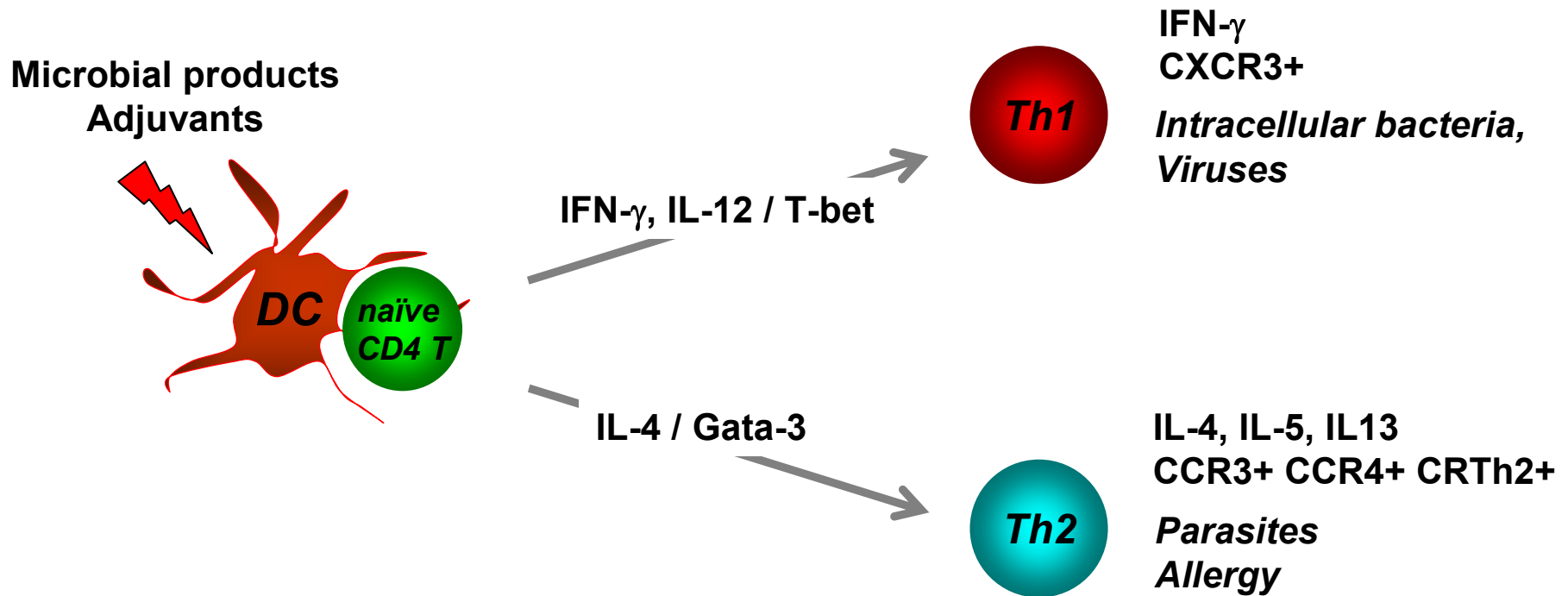
Secrete IFN- $\gamma$  and perforin but little IL-2

Limited proliferation capacity but display of immediate effector function

Terminally differentiated




*Immediate protection*

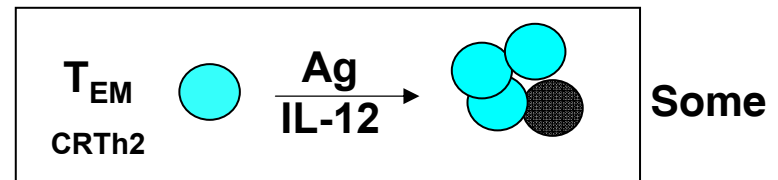
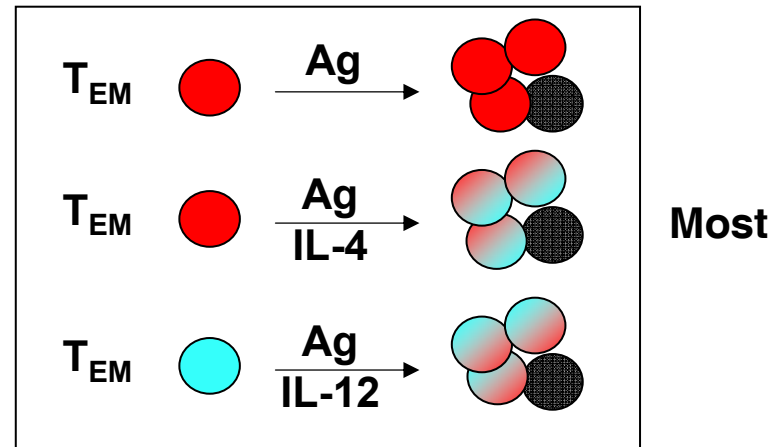
# The Th1 / Th2 paradigm



# Plasticity of cytokine gene expression in T<sub>EM</sub>

Chromatin configuration:

	CCR7	Cytokine production	Open chromatin
TH1		-	IFN- $\gamma$ <i>ifng</i>
TH2		-	IL-4 <i>il4</i>
TH0		-	IL-4+IFN- $\gamma$ <i>ifng and il4</i>

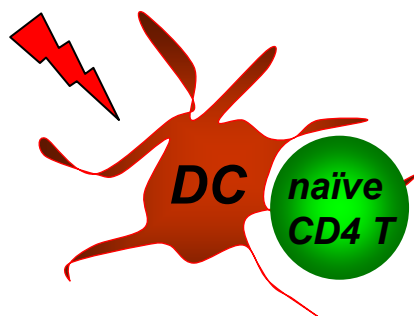


Subset specific, loss of T-bet

# Th17: a novel subset of CD4 effector T cells

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Microbial products  
Adjuvants



IFN- $\gamma$ , IL-12 / T-bet



IFN- $\gamma$   
*Intracellular bacteria,*  
*Viruses*

IL-4 / Gata-3



IL-4, IL-5, IL13  
*Parasites*  
*Allergy*

IL-6, IL-1, TGF $\beta$   
IL-23 / ROR $\gamma$ t



IL-17, IL-22, TNF  
*Autoimmunity*  
*Extracellular bacteria?*



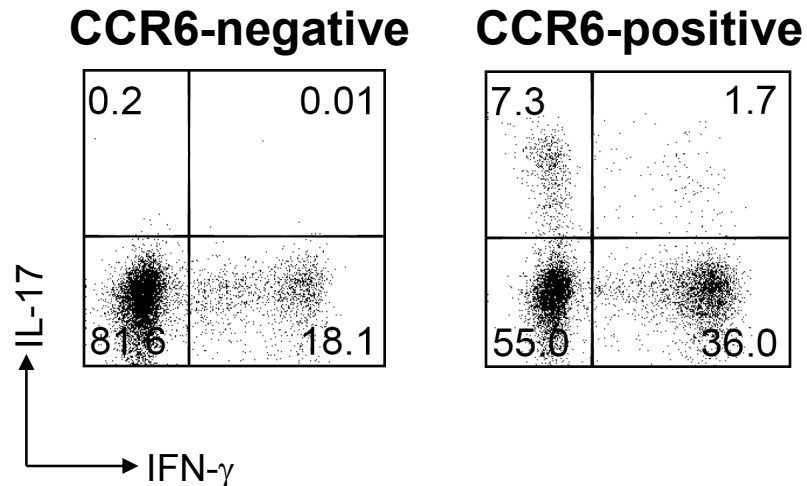
**Can we identify Th17 cells using surface markers?**

**Which are the pathogens that trigger Th17 responses?**

# IL-17 production is characteristic of a CCR6<sup>+</sup> subset

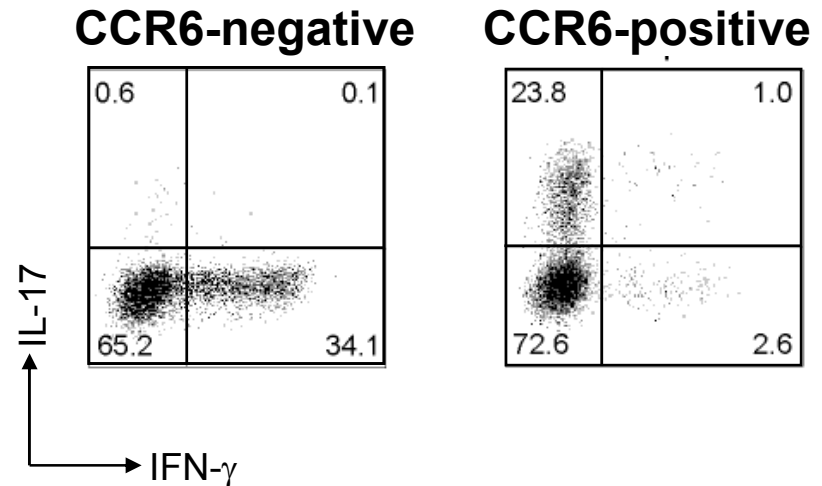
**Human** memory T cells  
(CD4<sup>+</sup> CD45RA<sup>-</sup> CD25<sup>-</sup>)

**Ex-vivo from blood**

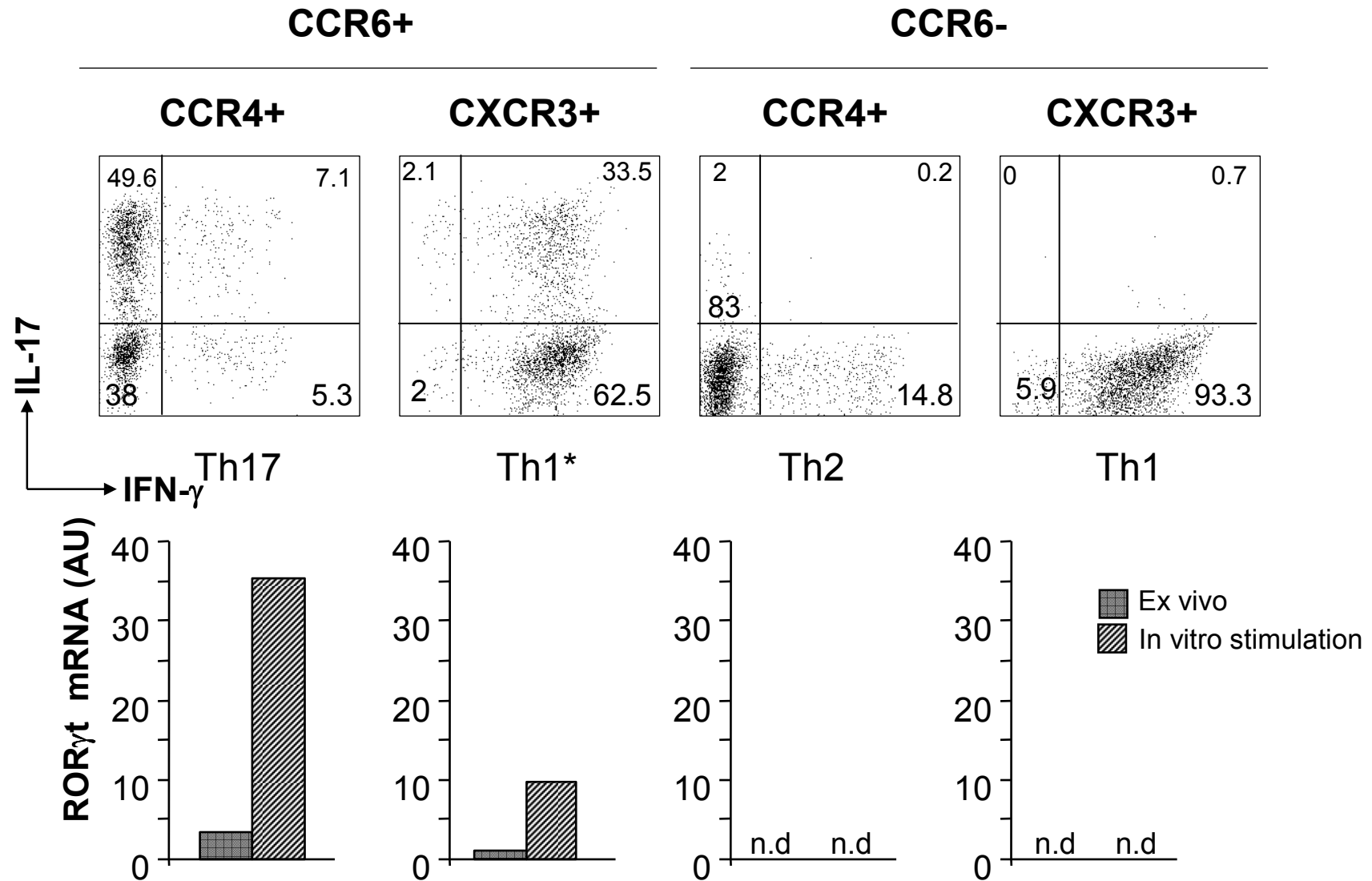


**Mouse** memory T cells  
(CD4<sup>+</sup> CD25<sup>-</sup> CD44<sup>hi</sup>)

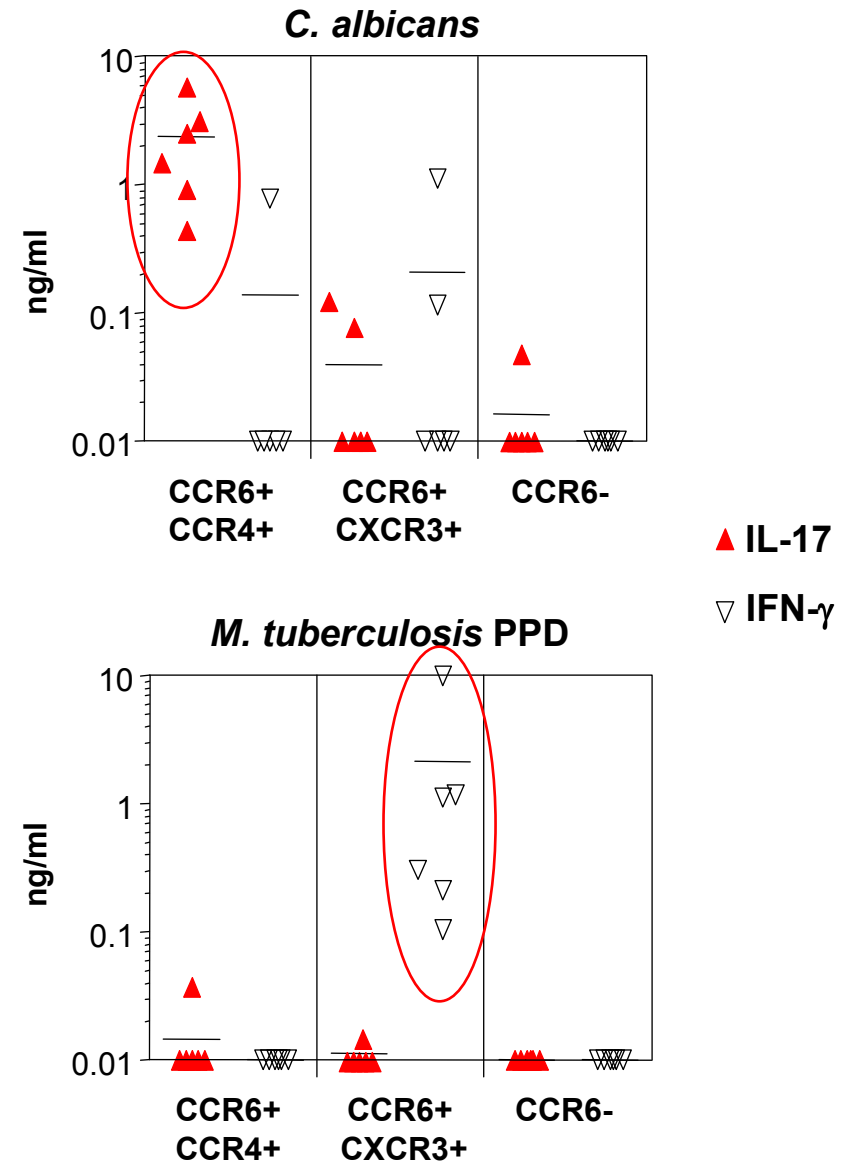
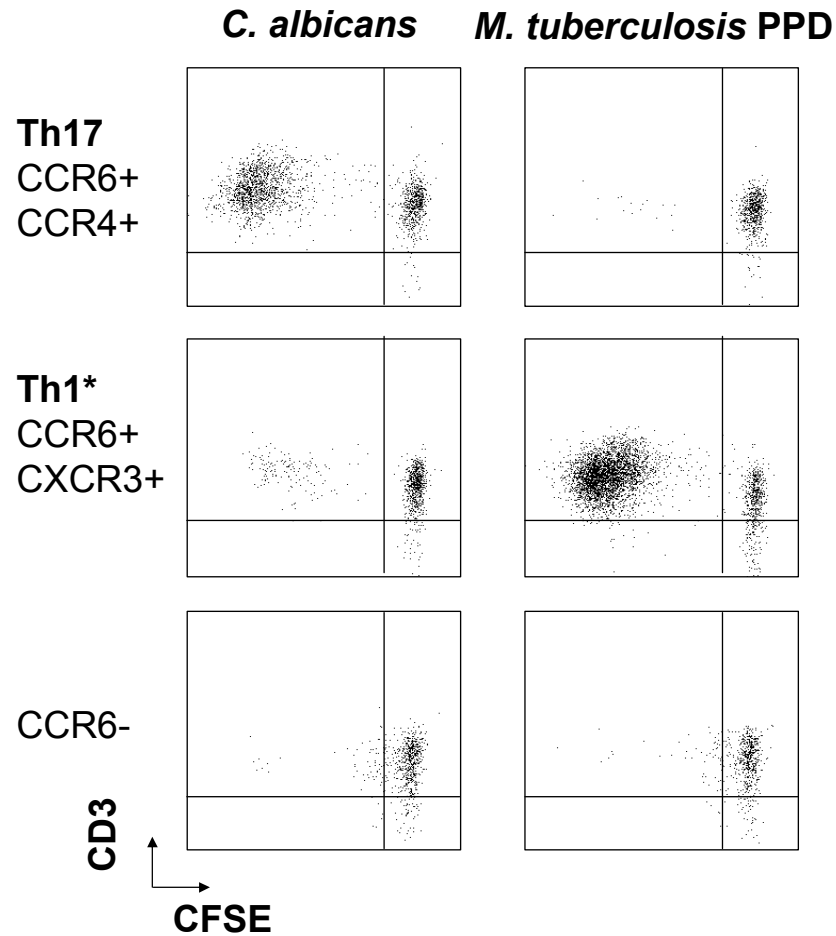
**Ex-vivo from spleen**



# Human Th17 memory cells co-express CCR6 and CCR4 and express ROR $\gamma$ t

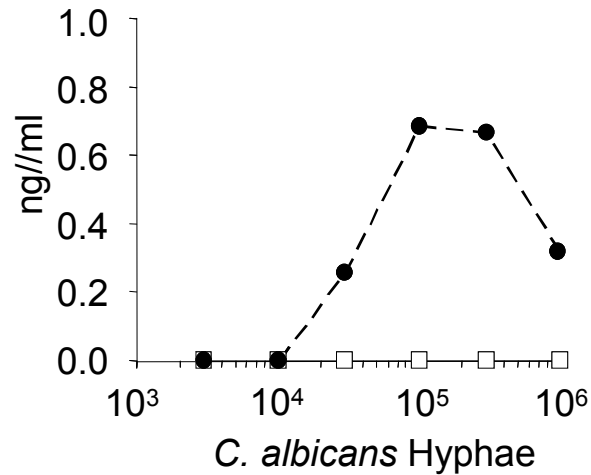
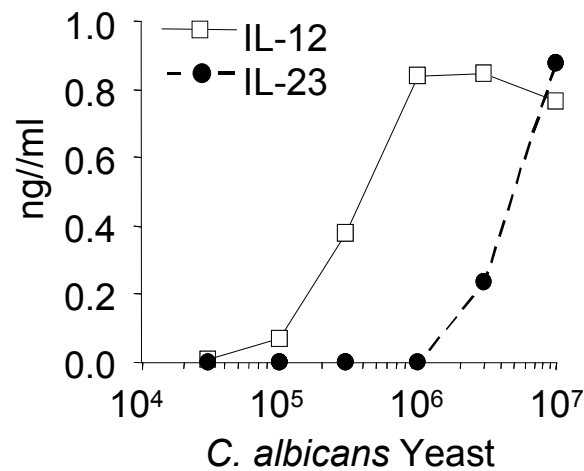


# *C. albicans* specific memory cells are Th17

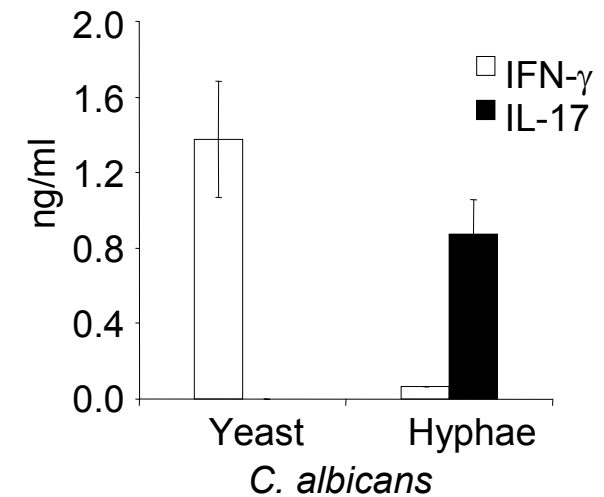


# *C. albicans* hyphae selectively induce IL-23 and prime Th17 cells

IL-12 and IL-23 production by DC



Priming naïve CD4+ T cells



# Th17 and immunity to fungi

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- Th17 memory cells express CCR6 and CCR4
- *C. albicans* specific memory cells are Th17
- The Hyphal form of *C. albicans* triggers IL-23, but not IL-12
- Th17 deficiency and candidiasis in STAT3 deficient patients (Milner et al Nature 2008; Ma et al JEM 2008)

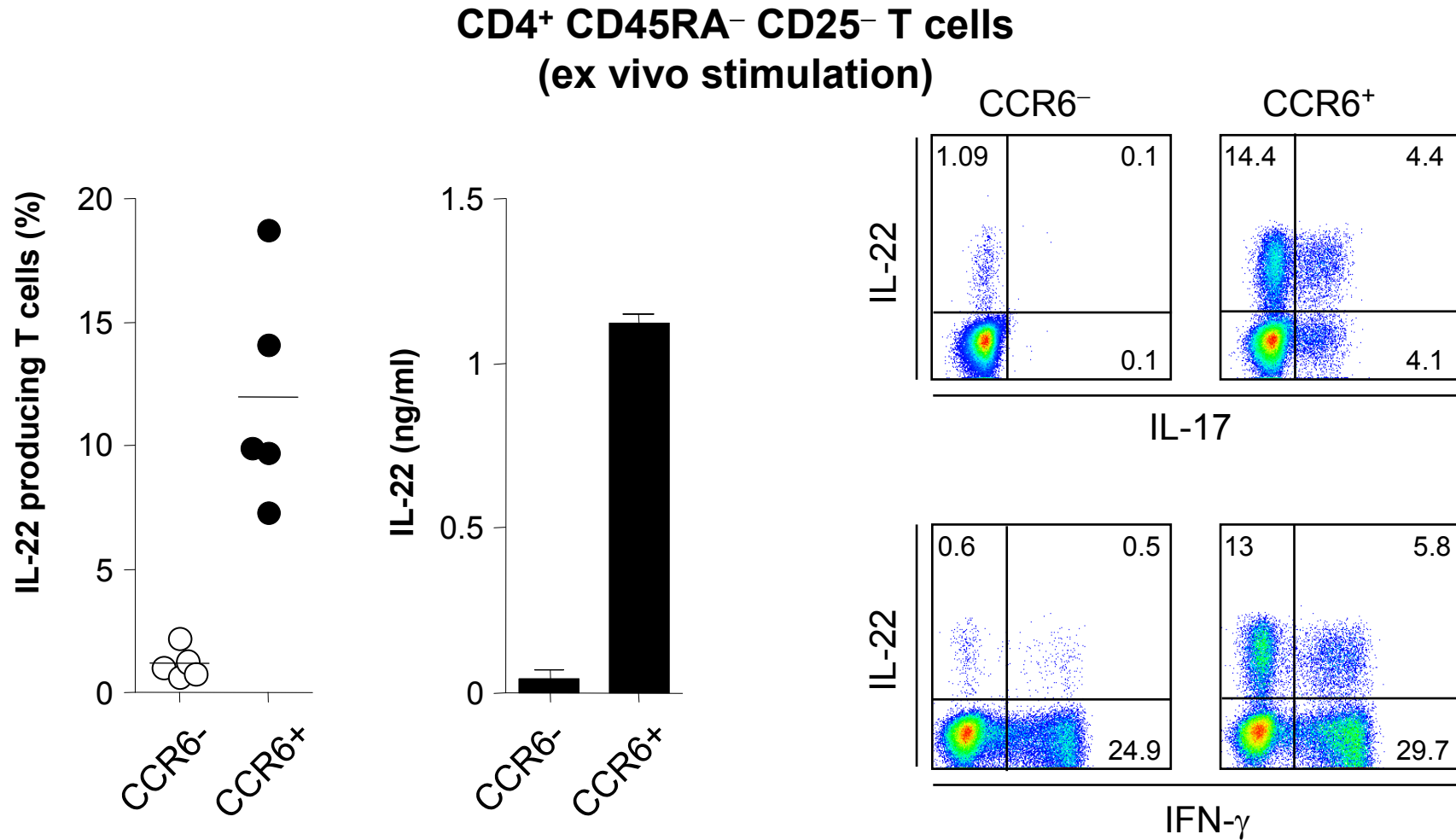
*Th17 cells might recruit neutrophils thus promoting entrapment and killing of hyphae*



## Th22: a new subset of T helper cells?

Thomas Duhén

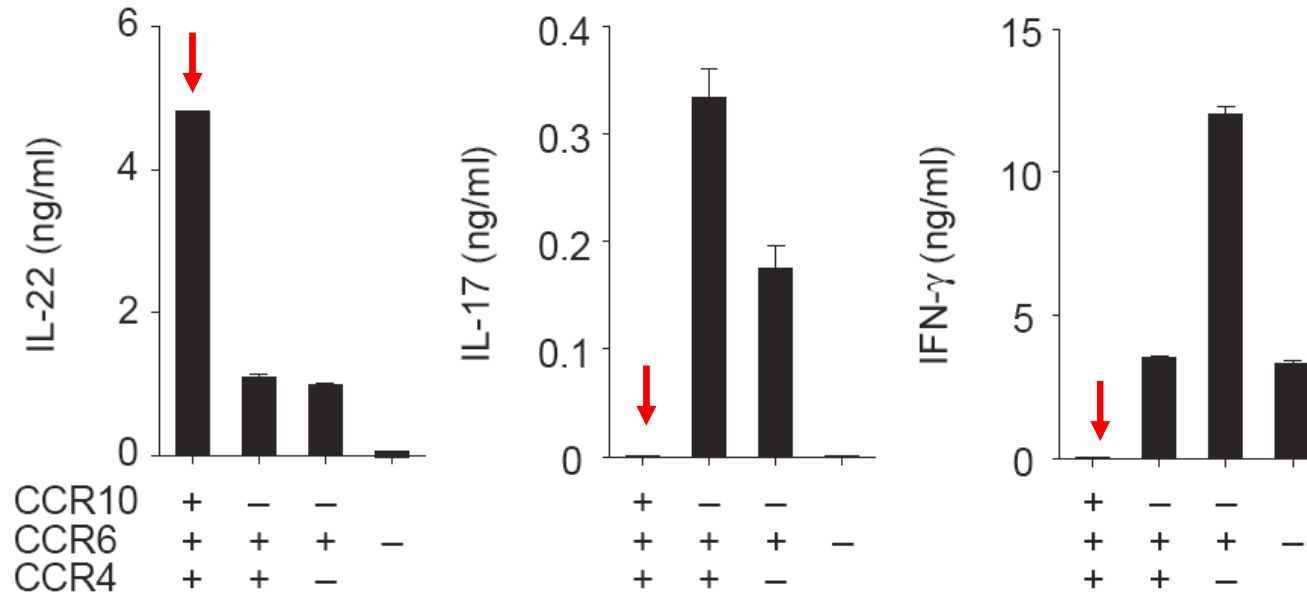
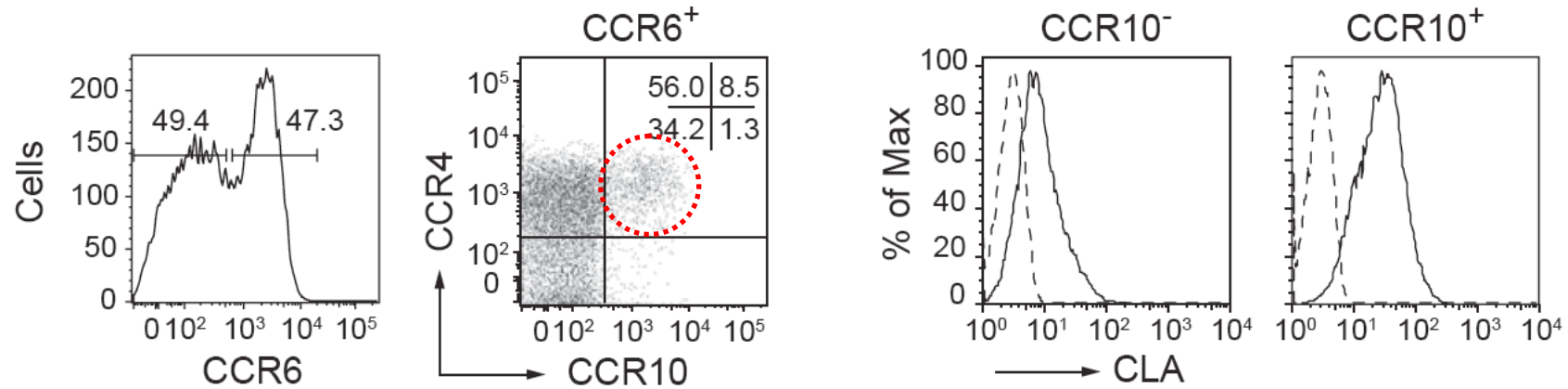
# IL-22-producing T cells are present in the CD4<sup>+</sup> CCR6<sup>+</sup> memory subset



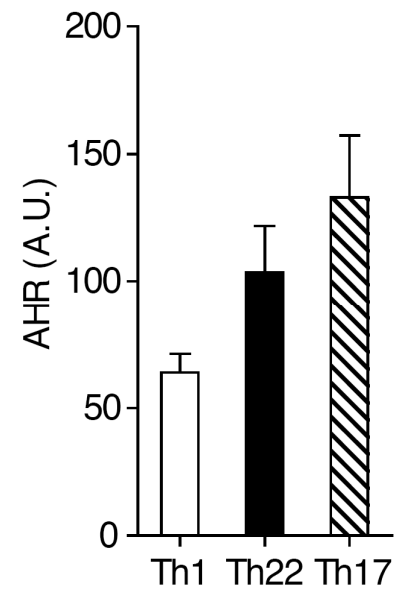
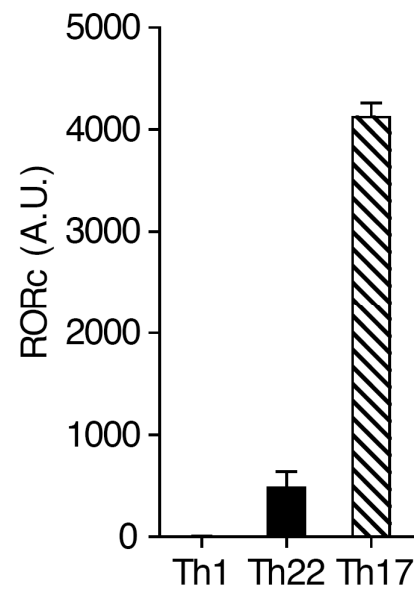
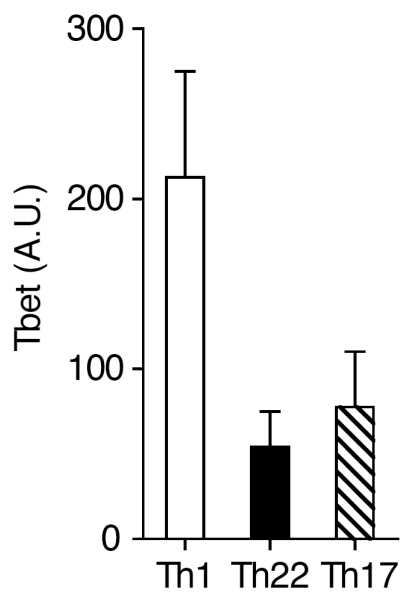
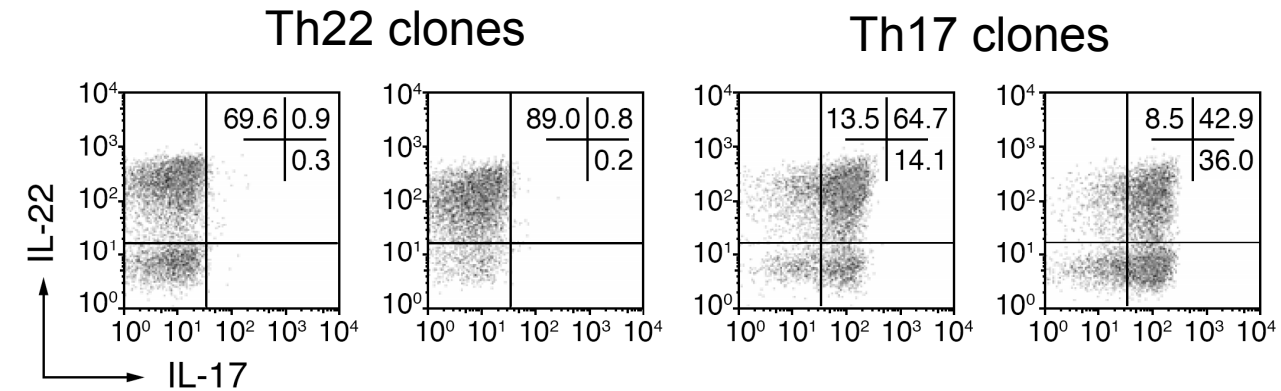
... but most IL-22-producing cells do not produce IL-17



# Skin-homing T cells (CCR4+ CCR10+ CLA+) produce IL-22, but no IL-17



# Th22 clones do not express ROR-c

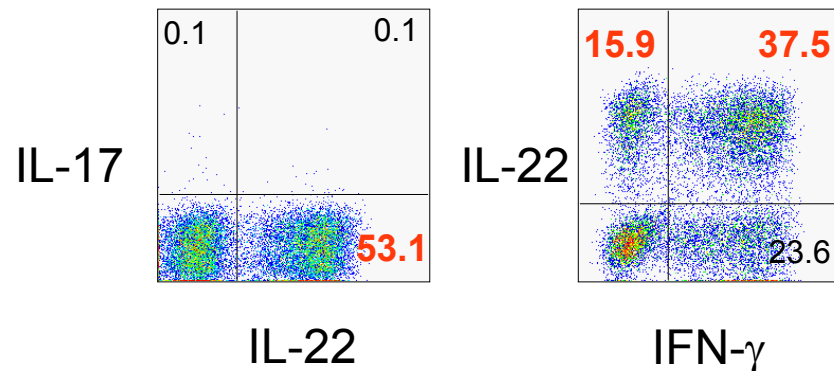
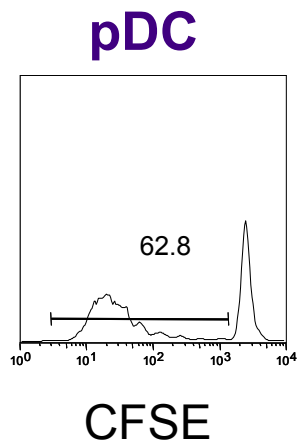
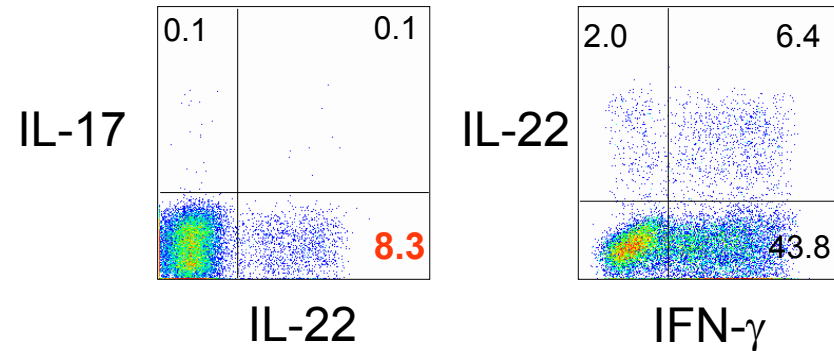
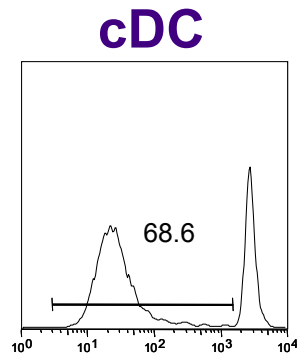


*How are IL-22-only producing CD4<sup>+</sup> T cells generated?*

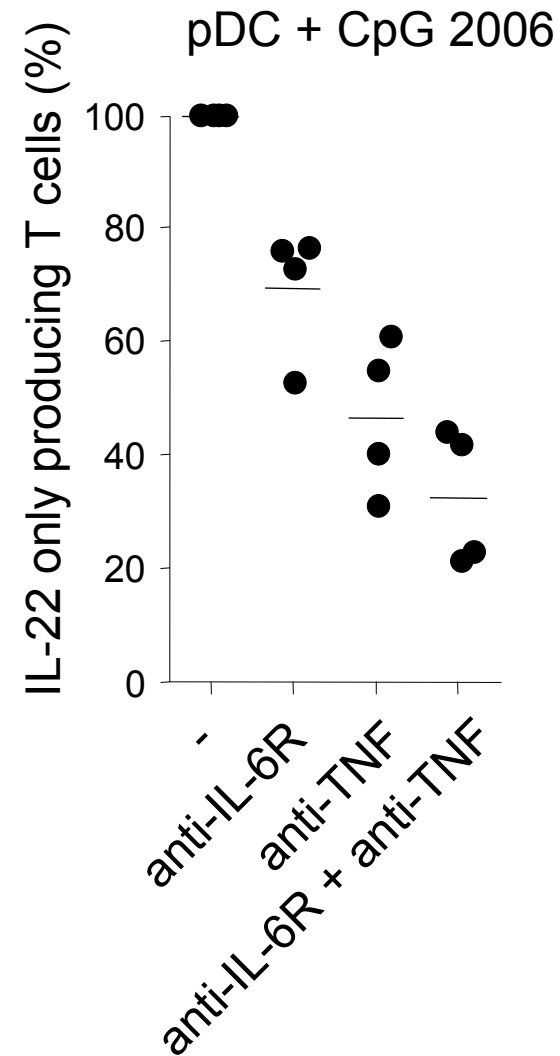
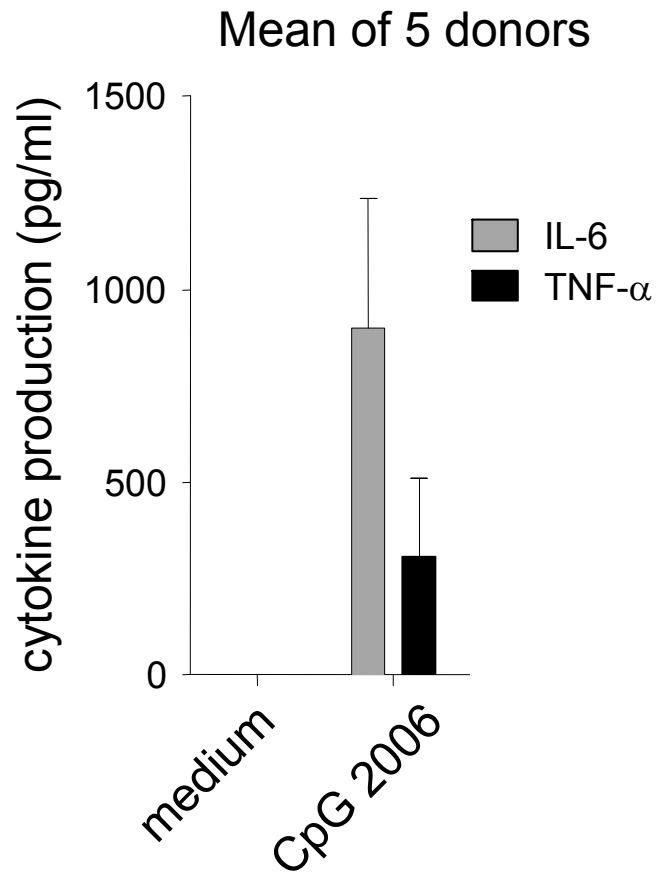
# Plasmacytoid DC prime IL-22-producing T cells

CD4+ naïve T cells  
primed by allogeneic:

Cytokine production  
following PMA + ionomycin

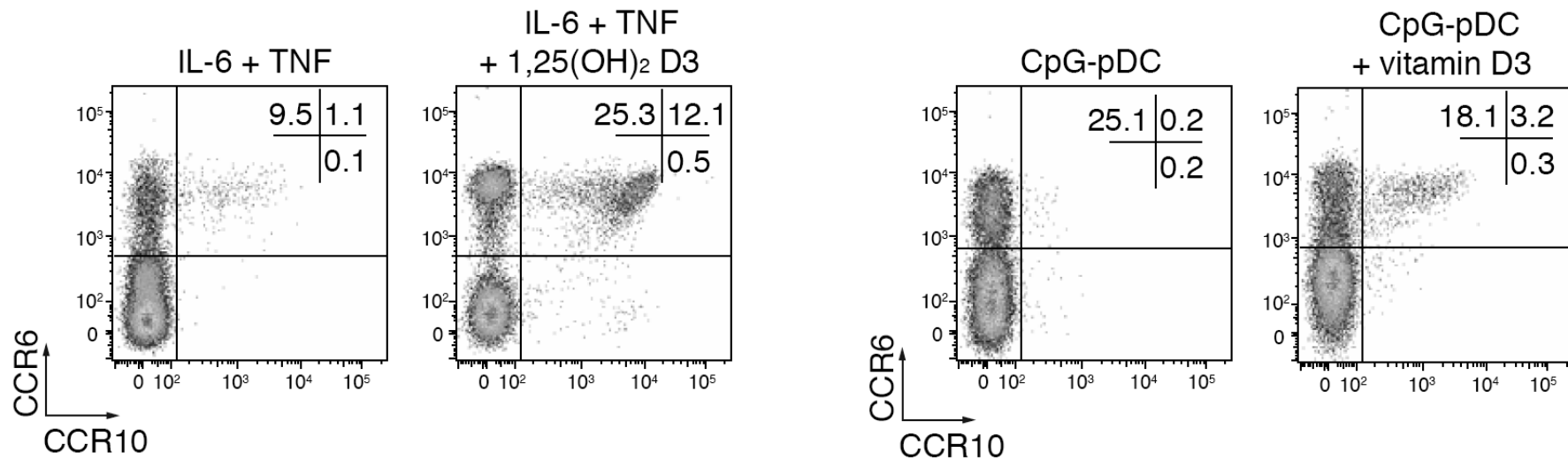


# IL-6 and TNF produced by pDC are required for Th22 polarization



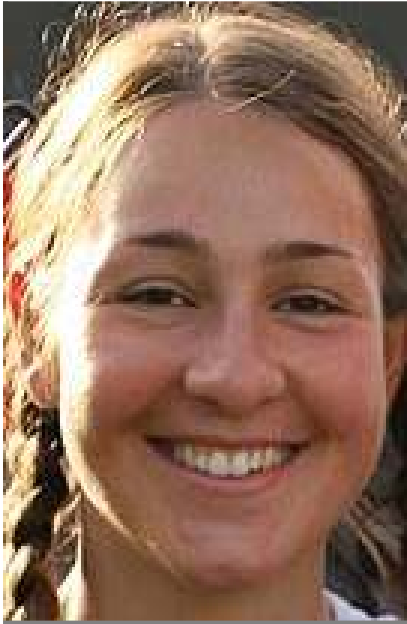
# Vitamin D3 and pDC promote expression of CCR6 and CCR10

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# Th22: a module of adaptive immunity dedicated to epithelial cell physiology?

Module	Polarizing cytokine(s)	Transcription factor	Homing receptor(s)	Effector cytokine(s)	Target cell	Function
<b>T<sub>H</sub>1</b>	IL-12, IFN	T-bet	CXCR3	IFN- $\gamma$	Macrophages	<b>Bacteria</b>
<b>T<sub>H</sub>2</b>	IL-4	GATA-3	CCR4/CRT <sub>H</sub> 2	IL-4, IL-5, IL-13	Eosinophils	<b>Parasites</b>
<b>T<sub>H</sub>17</b>	IL-6, IL-1 $\beta$ , TGF- $\beta$	ROR- $\gamma$ t	CCR6 / CCR4	IL-17, IL-22	Neutrophils	<b>Fungi</b>
<b>Treg</b>	?	FOXP3	CCR7 / CCR6	TGF- $\beta$	DC / T cells	<b>Regulation</b>
<b>T<sub>FH</sub></b>	IL-21	Bcl-6	CXCR5	IL-21	B cells	<b>Antibodies</b>
<b>Tr1</b>	IL-10	?	CCR7 / CCR6	IL-10	T cells	<b>Regulation</b>
<b>T<sub>H</sub>22</b>	IL-6, TNF	?	CCR6 / CCR10	IL-22	Keratinocytes	?



## Human T cell repertoire analysis using amplified T cell libraries

Rebekka Geiger



# Challenges in analyzing the human naïve T cell repertoire

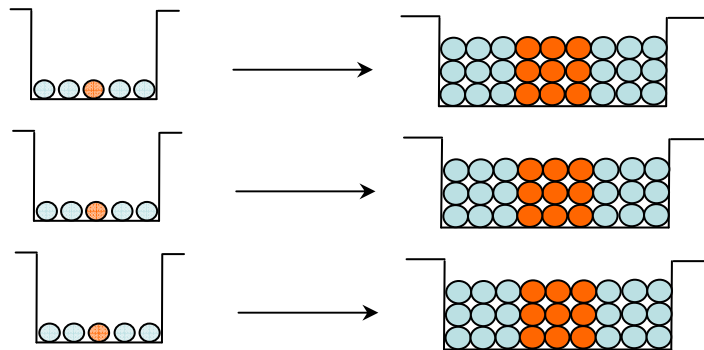
- Low frequency of antigen-specific naïve T cells
- High activation threshold of naïve T cells
- Broad spectrum of avidities
- Limitations of peptide-based and tetramer-based approaches
- Need to measure T cell responses to complex naturally processed antigens

# Analysis of naïve and memory T cell repertoires using amplified T cell libraries

## 1<sup>st</sup> step: Amplification

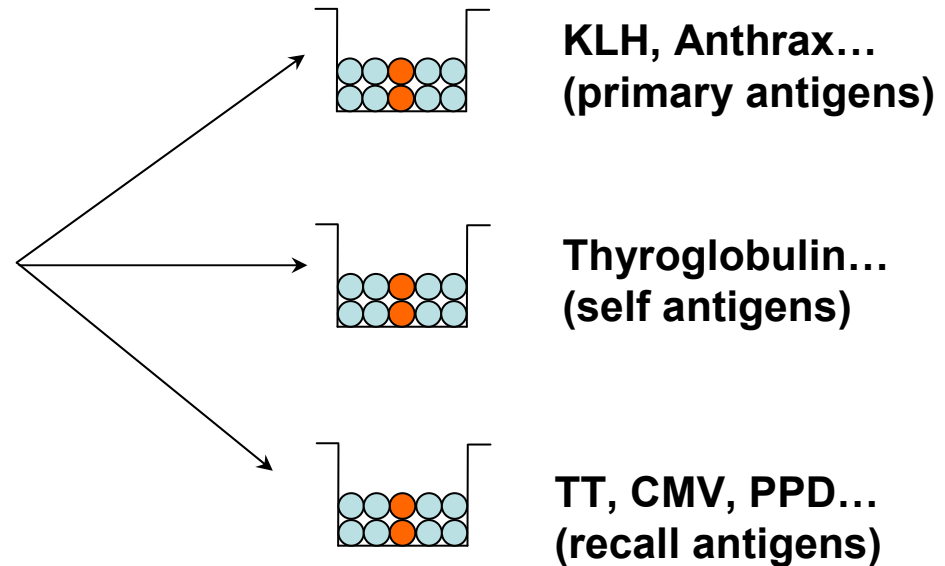
Multiple cultures (96 up to 384) each containing 1000 cells are expanded with PHA and IL-2

1 naïve T cell → 5,000 T cell blasts



## 2<sup>nd</sup> step: Interrogation

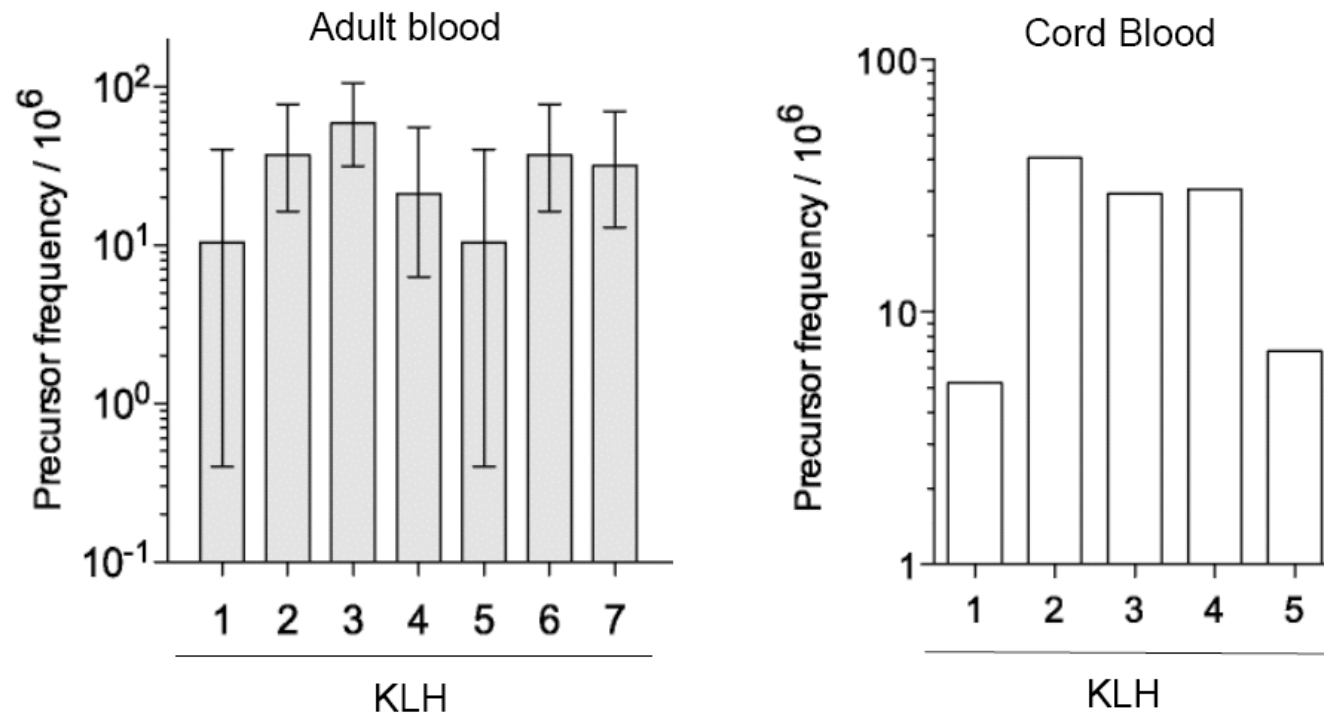
Each culture is tested for proliferation in response to Ag and autologous APC



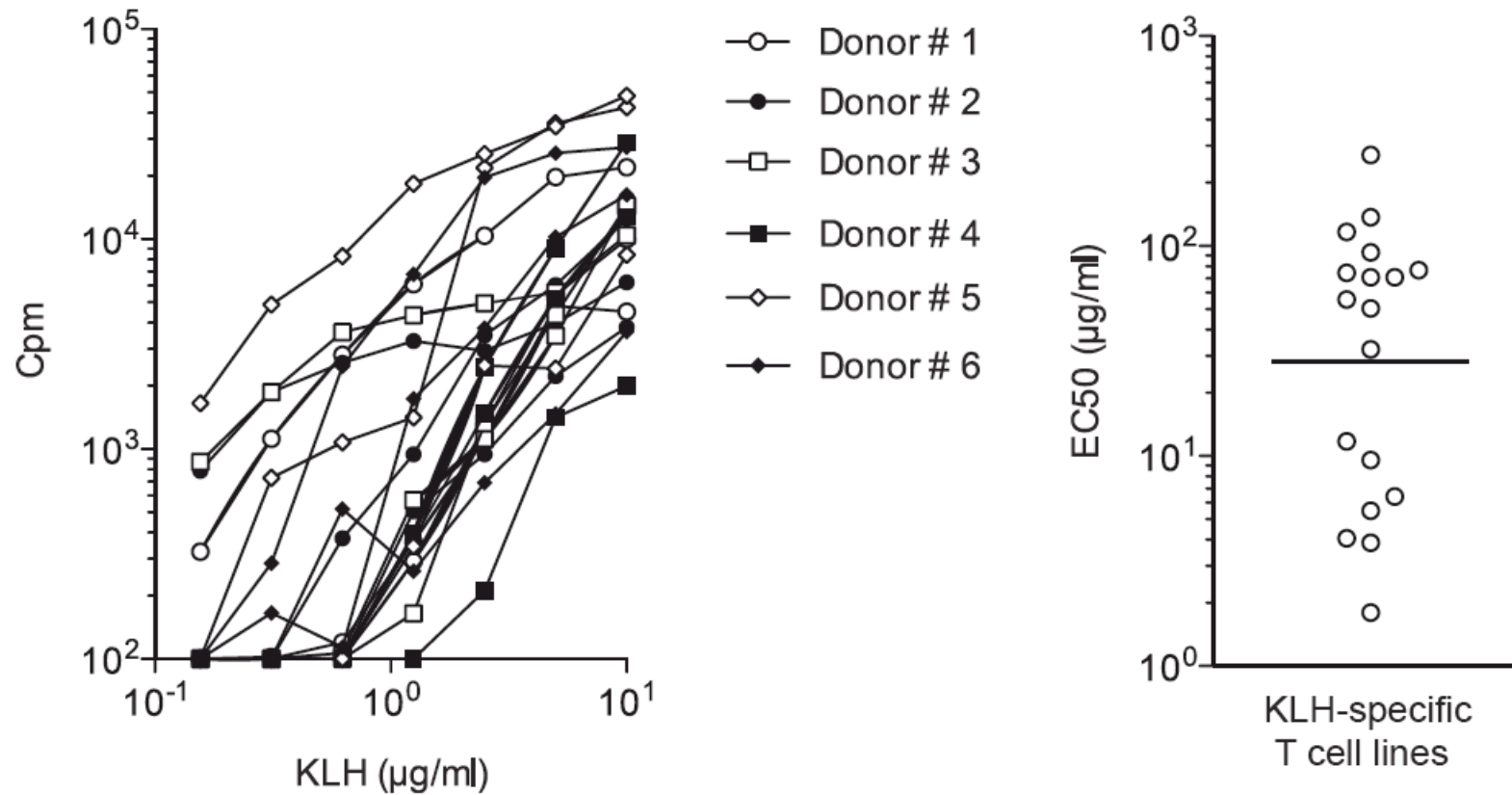


# Frequency of KLH-specific naïve T cells

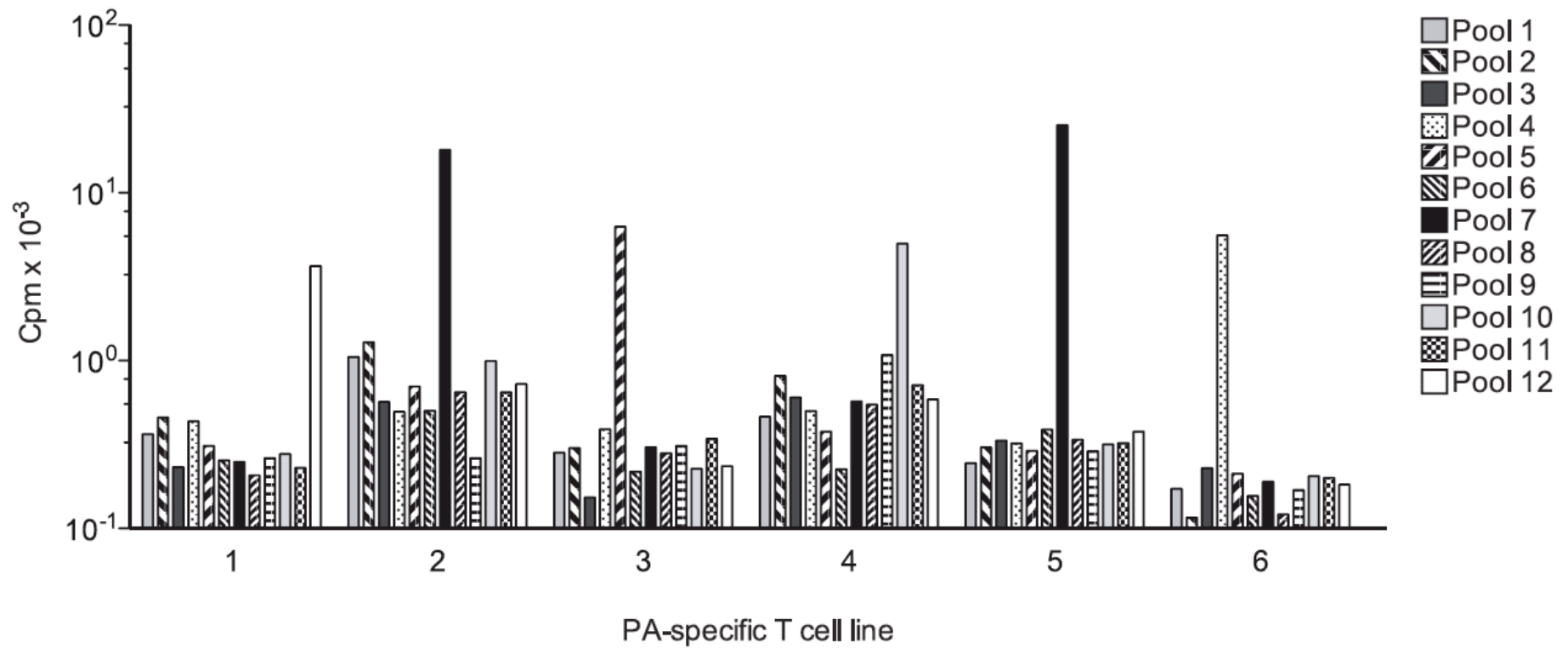
“Amplified T cell libraries” from naïve CD45RA<sup>+</sup> CD45RO<sup>-</sup> CCR7<sup>+</sup> CD4<sup>+</sup> T cells



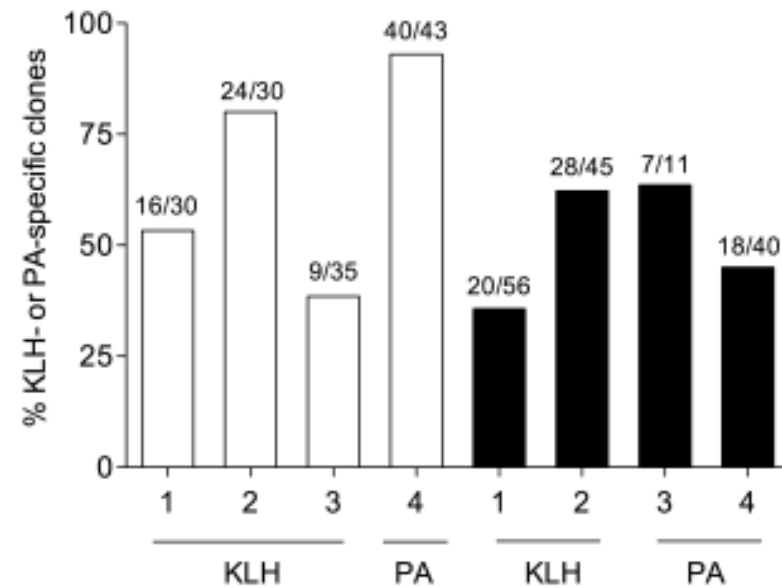
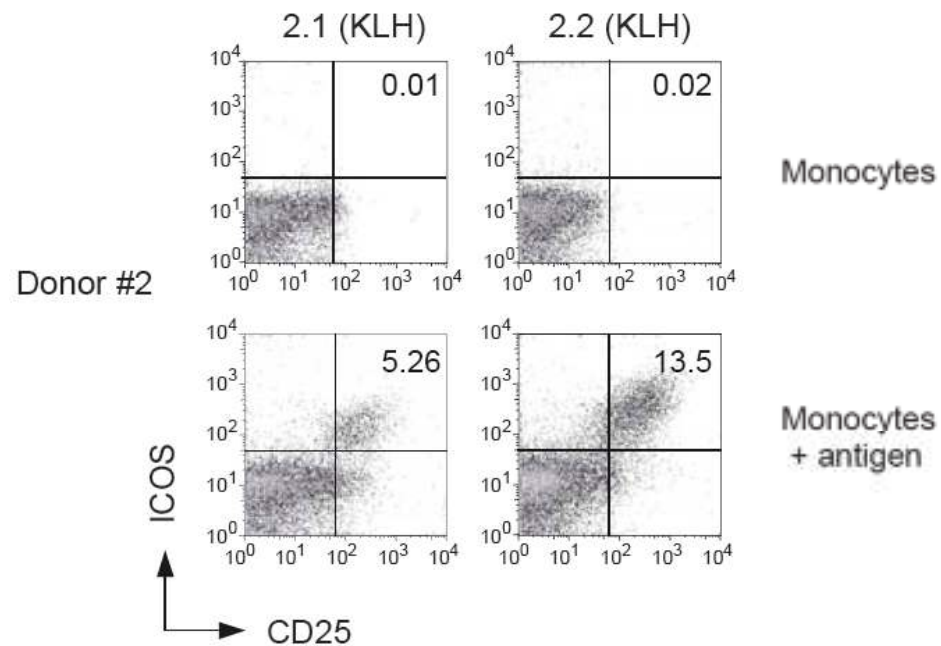
# Broad range of responsiveness



# Broad range of epitope specificities

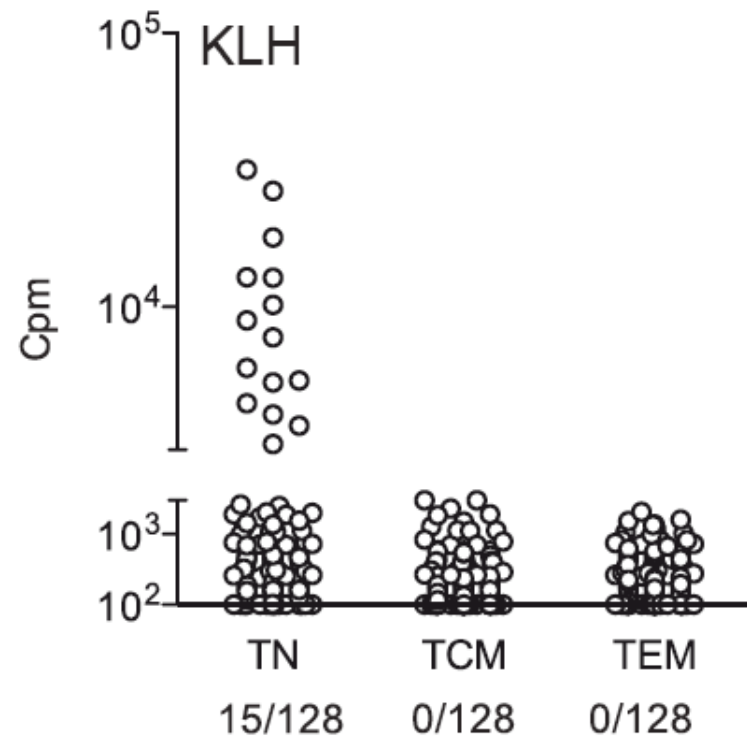


# Isolation of Ag-specific T cell clones from the naïve repertoire

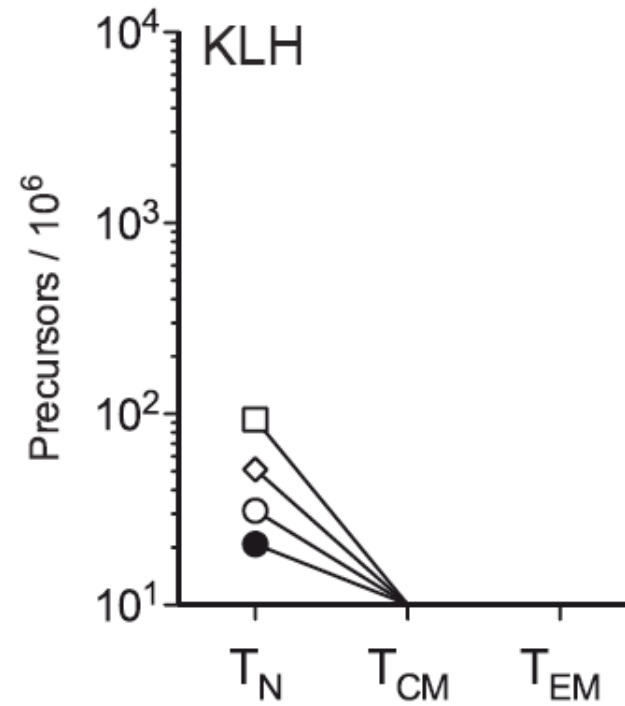


# Analysis of memory repertoires

**C**



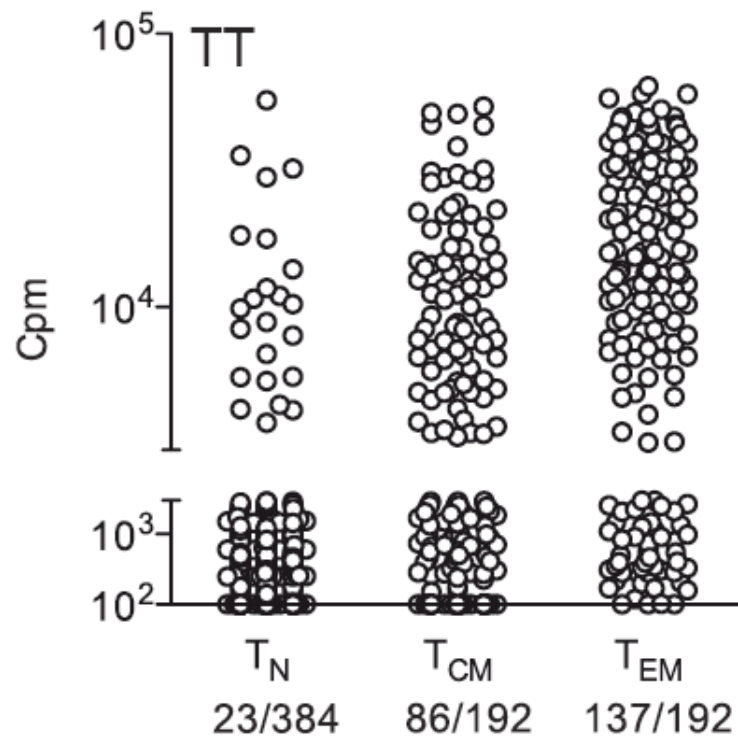
**D**



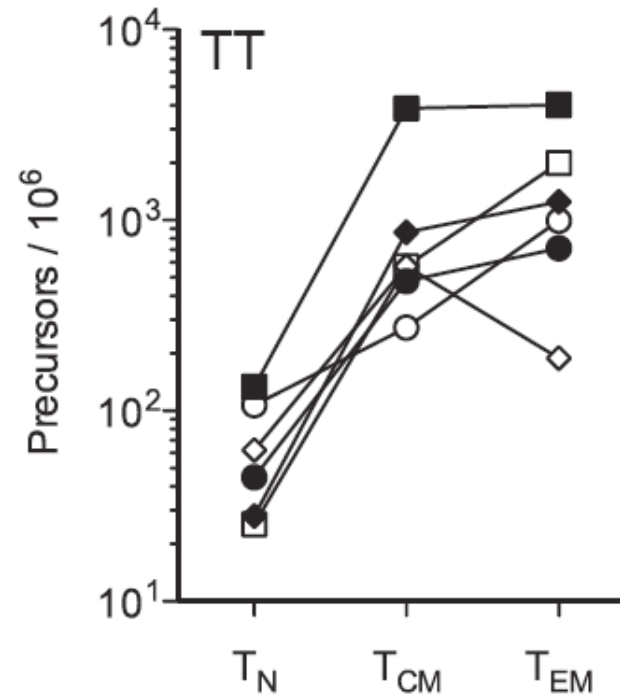


# Increased frequencies in the memory compartment (Tet Tox)

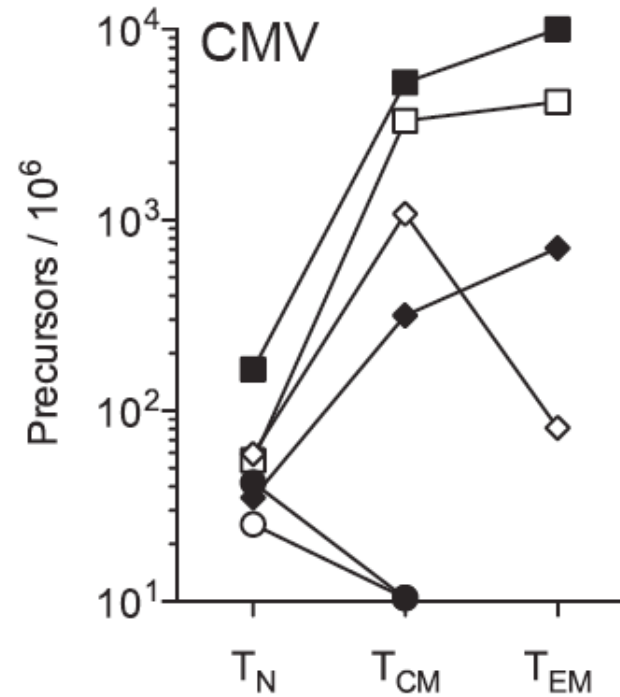
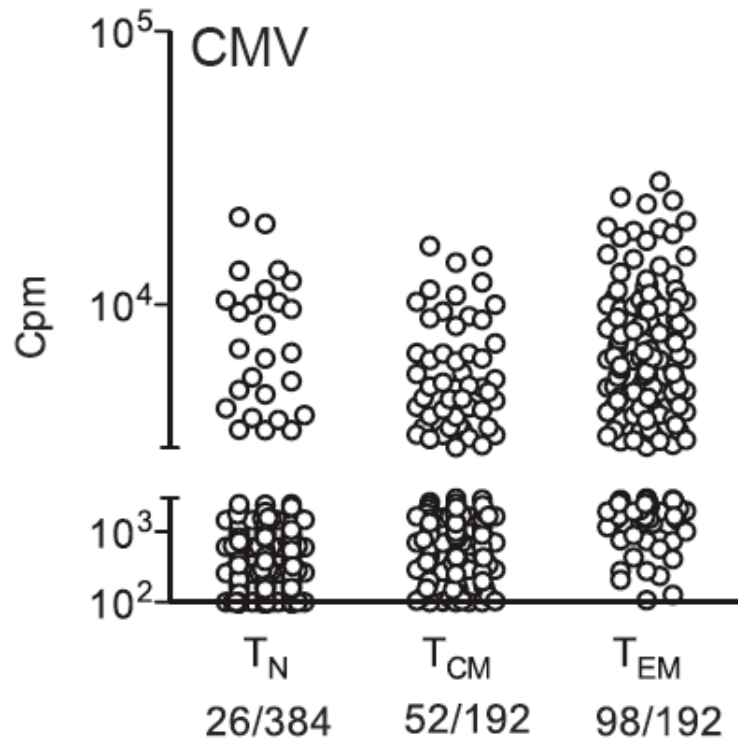
**A**



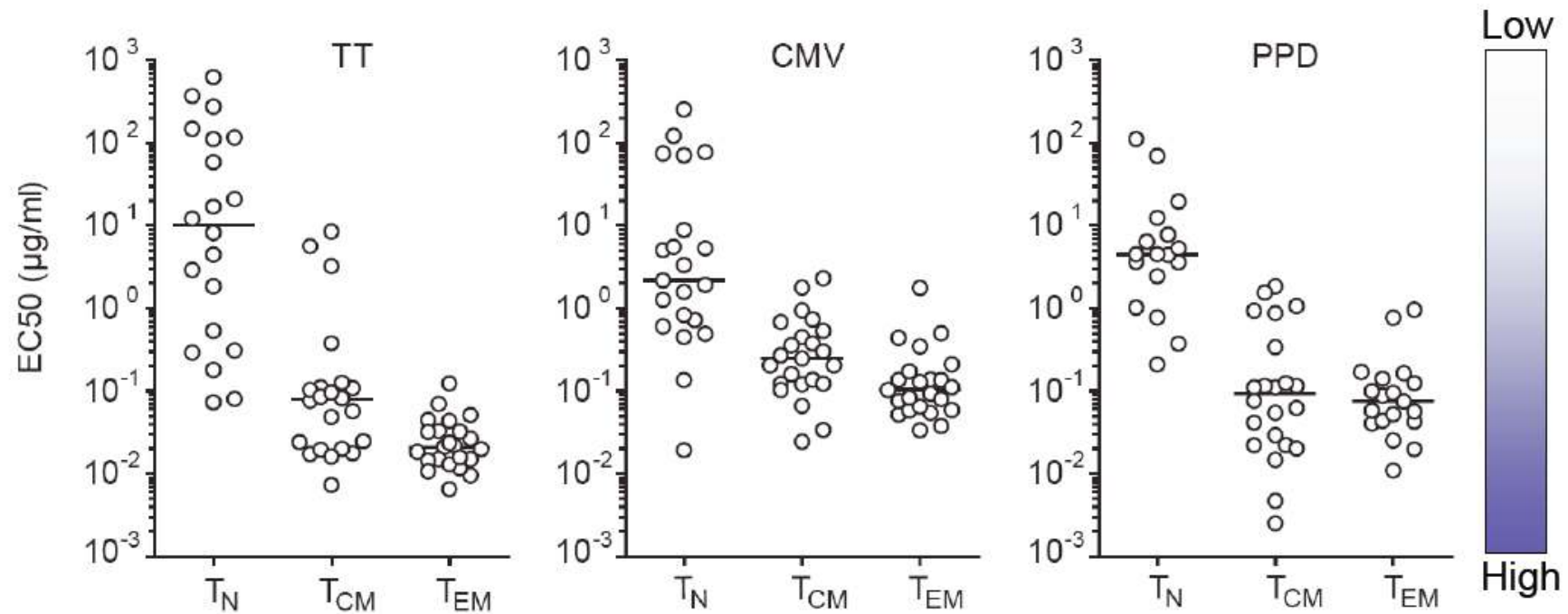
**B**



# Increased frequencies in the memory compartment (CMV)



# Selection of high avidity T cells in the memory pool



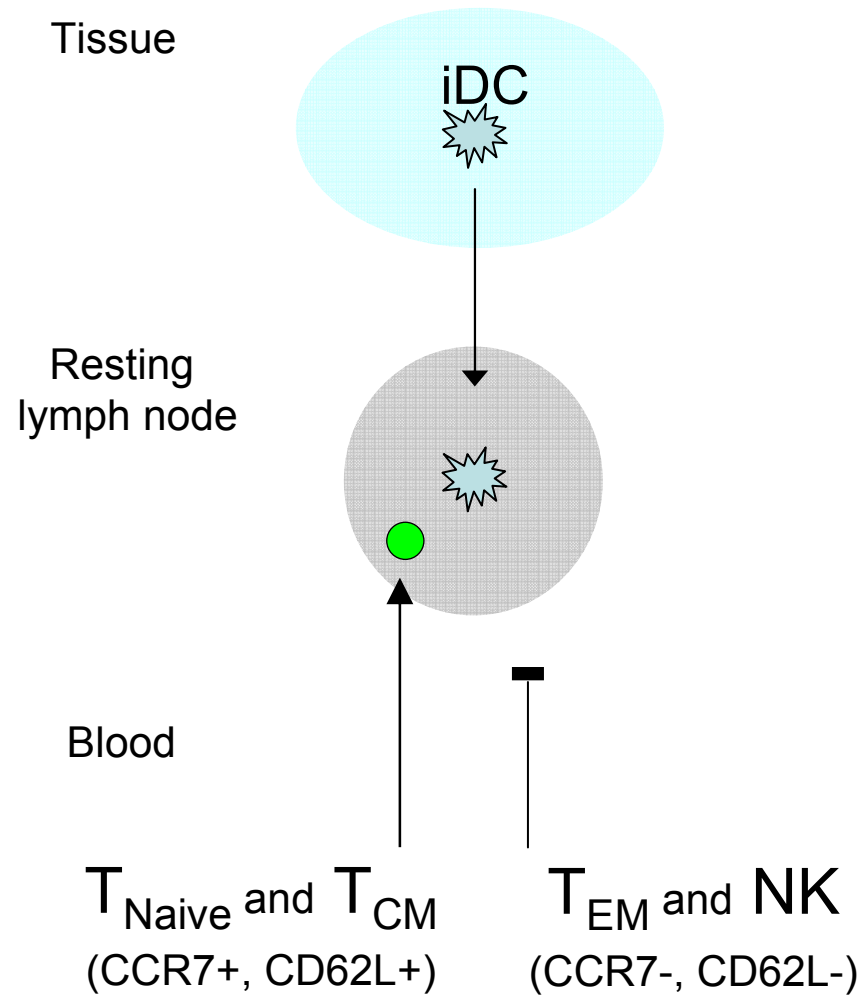
# Applications

- Predict antigenicity of complex molecules (even whole pathogens)
- Assess immunocompetence (elderly, HIV)
- Assess memory in different T cell subsets
- Generate T cells for cellular immunotherapy

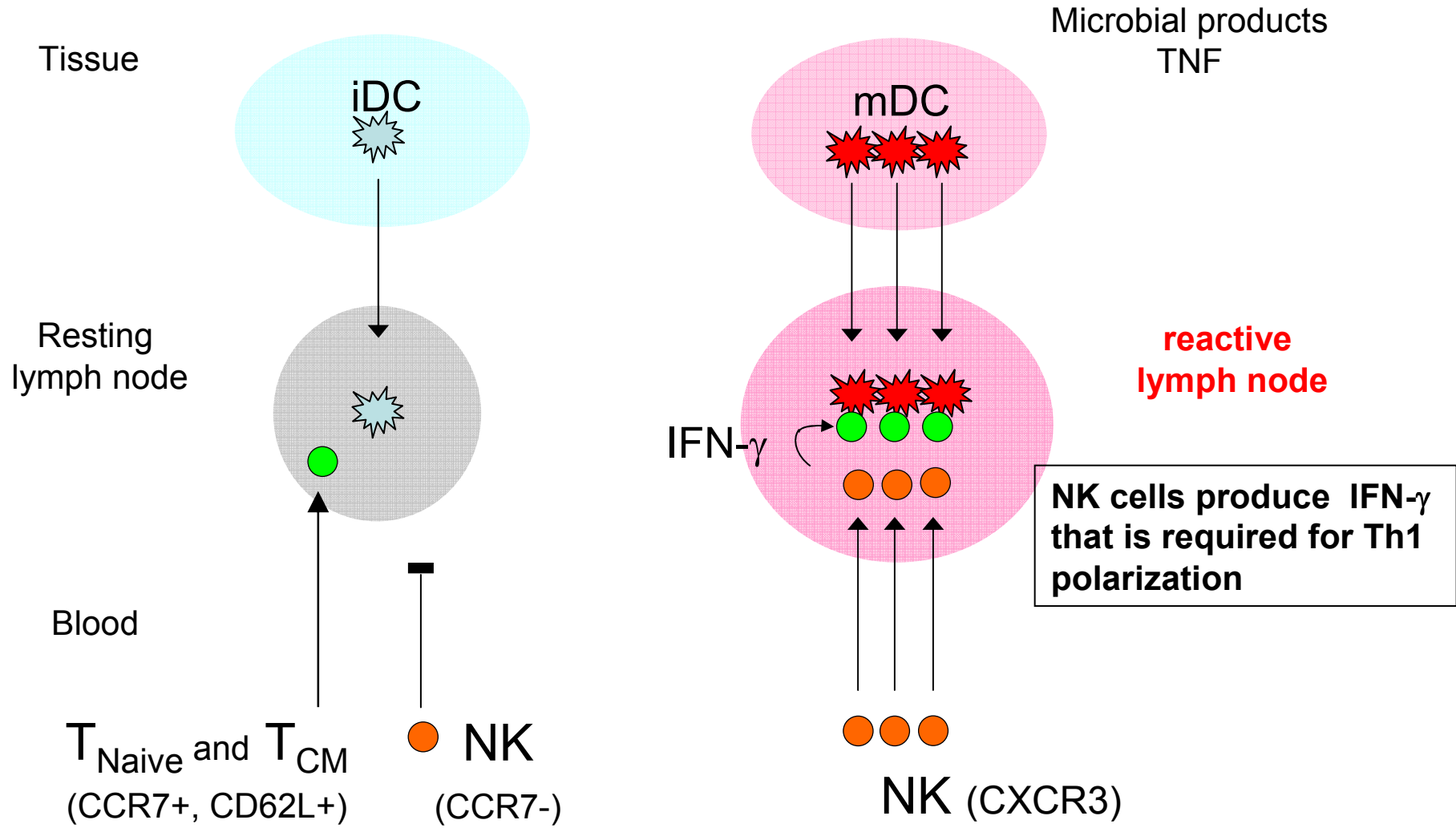
1. T cell activation and fate determination
2. Human effector/memory T cell subsets  
( $T_{CM}$ ,  $T_{EM}$ , Th1, Th2, Th17, etc)
3. T cell traffic in steady state and inflammatory conditions (mouse)

# Paradigm: migration from blood to lymph nodes requires CCR7 and CD62L

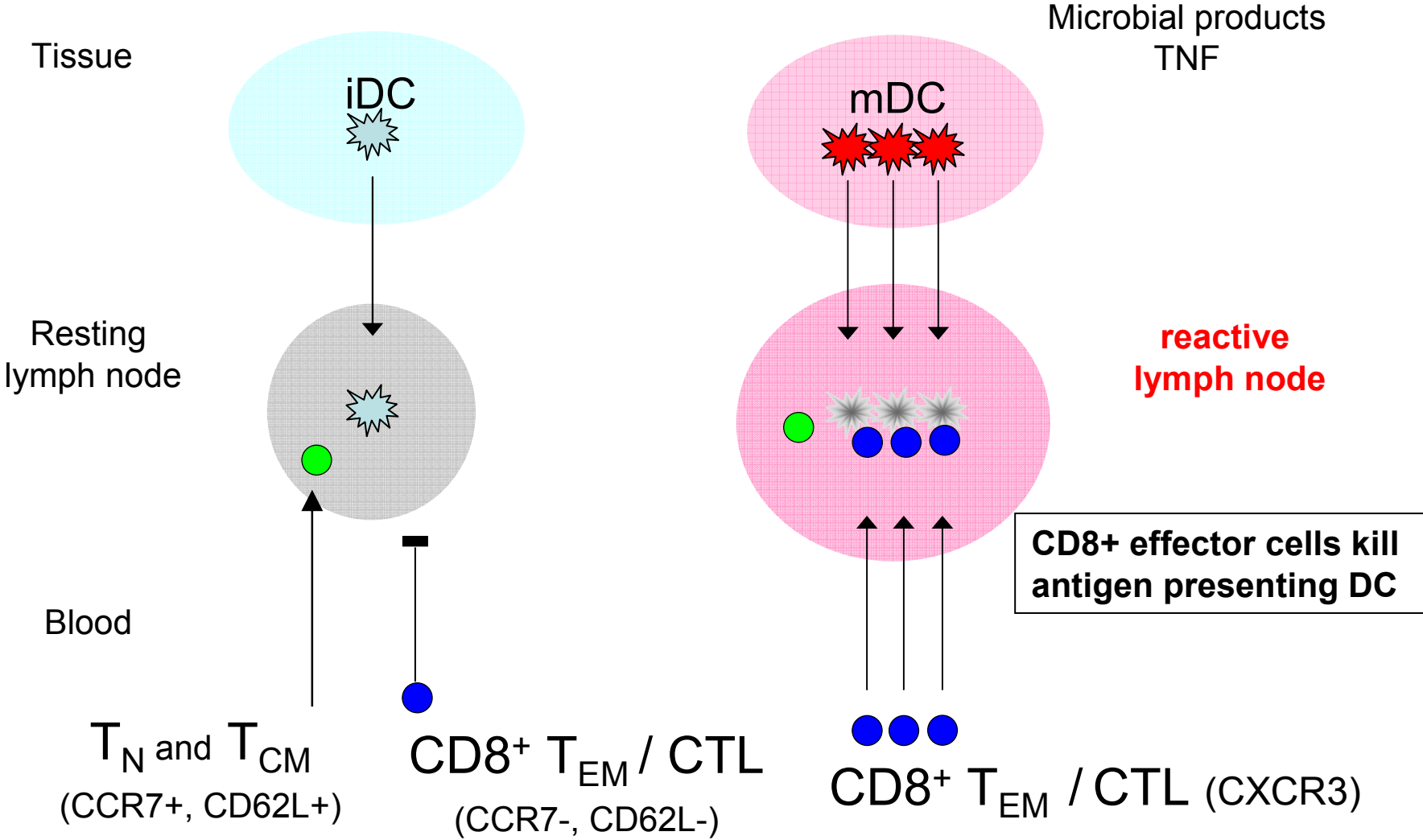
---



# NK cells migrate to reactive lymph nodes in a CXCR3-dependent fashion



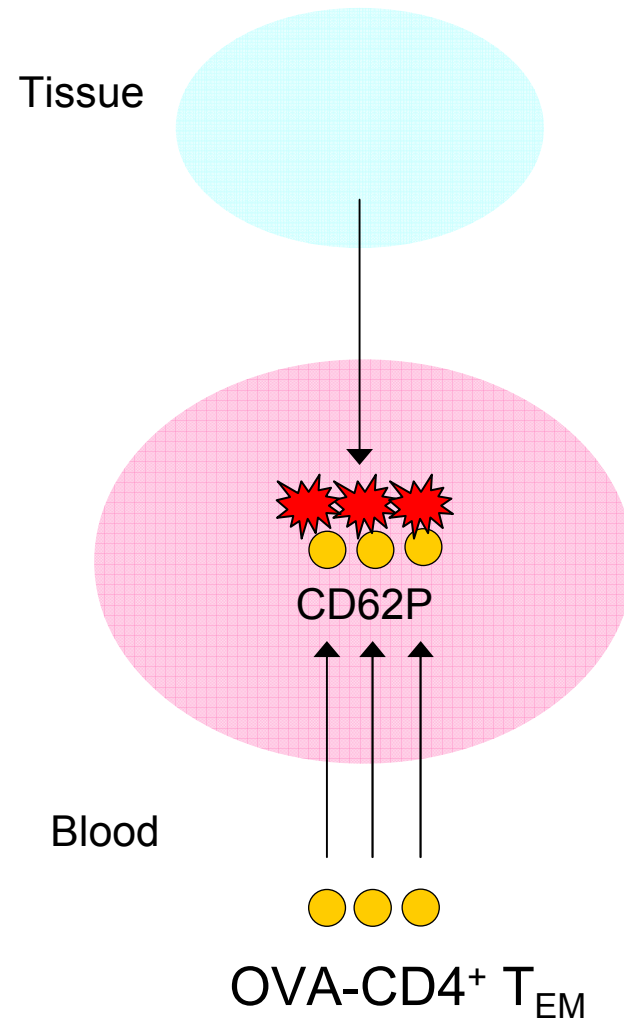
# Effector and effector memory CD8 T cells migrate to inflamed lymph nodes in a CXCR3 dependent fashion and kill antigen presenting dendritic cells





# Effector memory CD4 T cells license DC in chronically inflamed lymph nodes

---

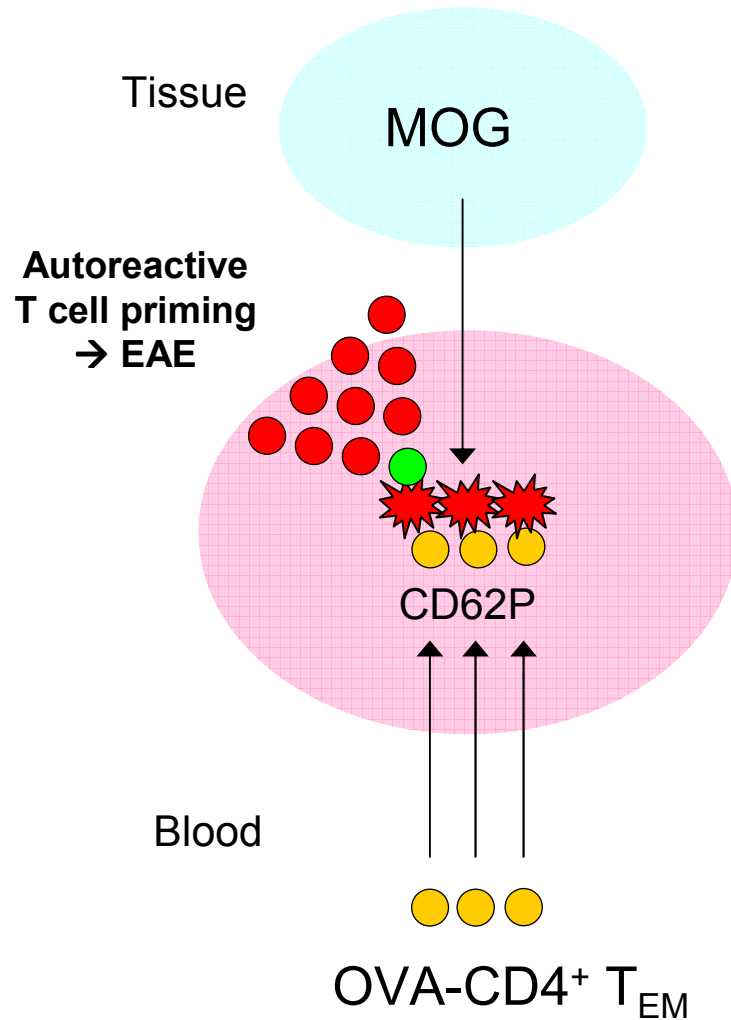


## CD4<sup>+</sup> effector memory T cells

- migrate to chronically reactive lymph nodes via CD62P
- Constitutively express CD40L
- Trigger DC maturation

# Effector memory CD4 T cells license DC in chronically inflamed lymph nodes

---

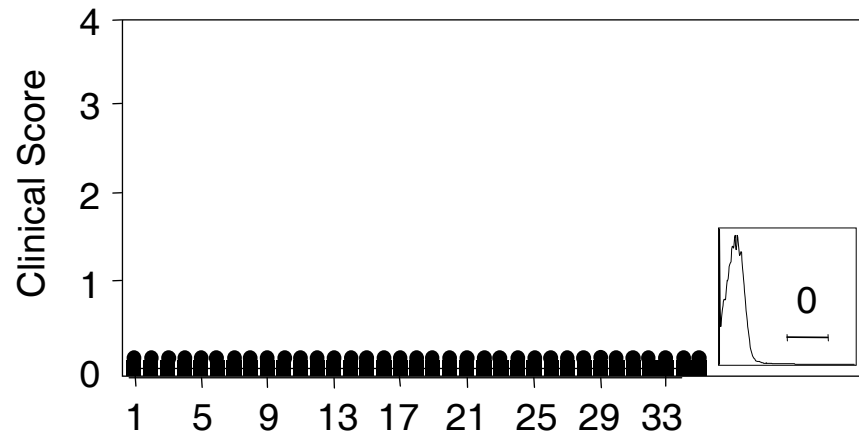


## CD4<sup>+</sup> effector memory T cells

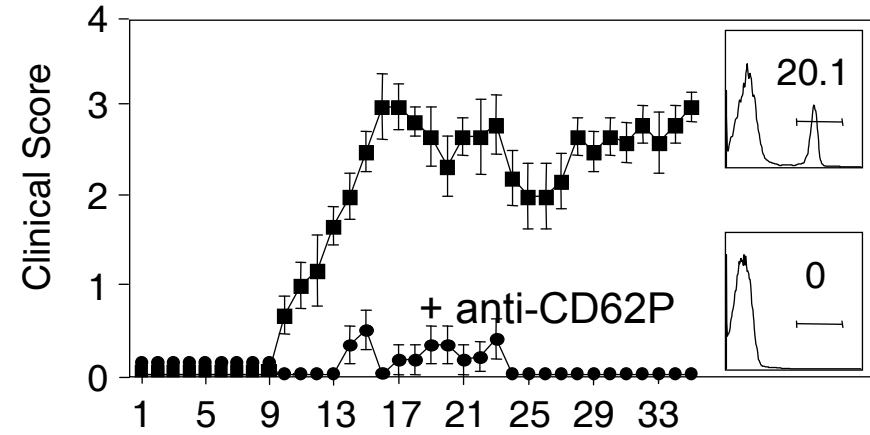
- migrate to chronically reactive lymph nodes via CD62P
- Constitutively express CD40L
- Trigger DC maturation
- Cause EAE in the absence of adjuvant

# Induction of EAE by MOG in PBS draining to a chronic lymph node

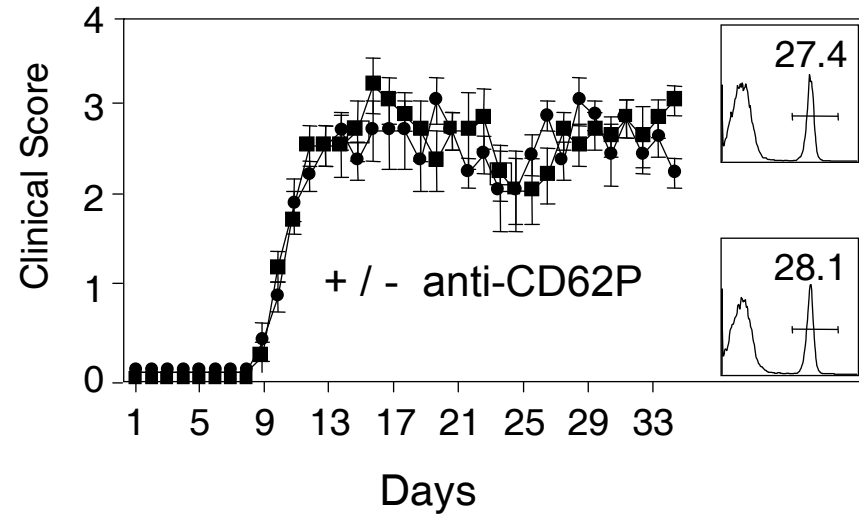
### MOG in PBS / Resting LN



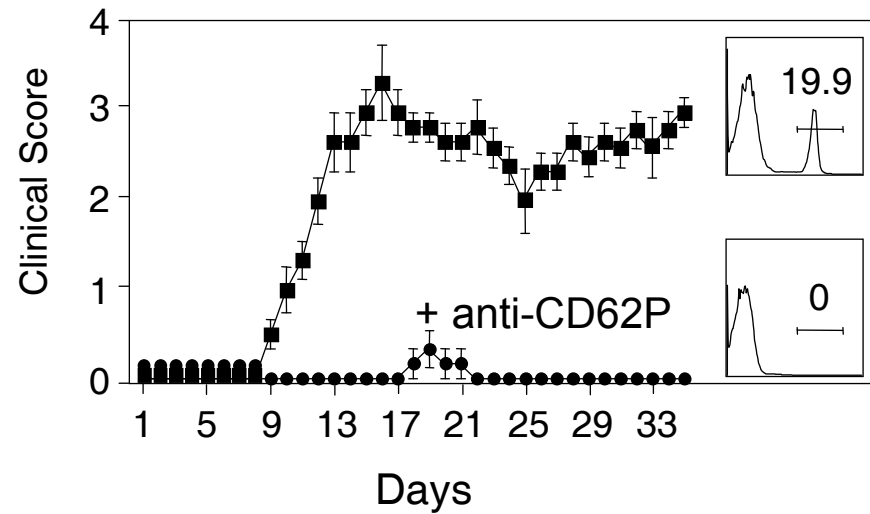
### MOG in PBS / Chronic LN (CFA)



### MOG in CFA / Resting LN



### MOG in PBS / Chronic LN (2X DC)



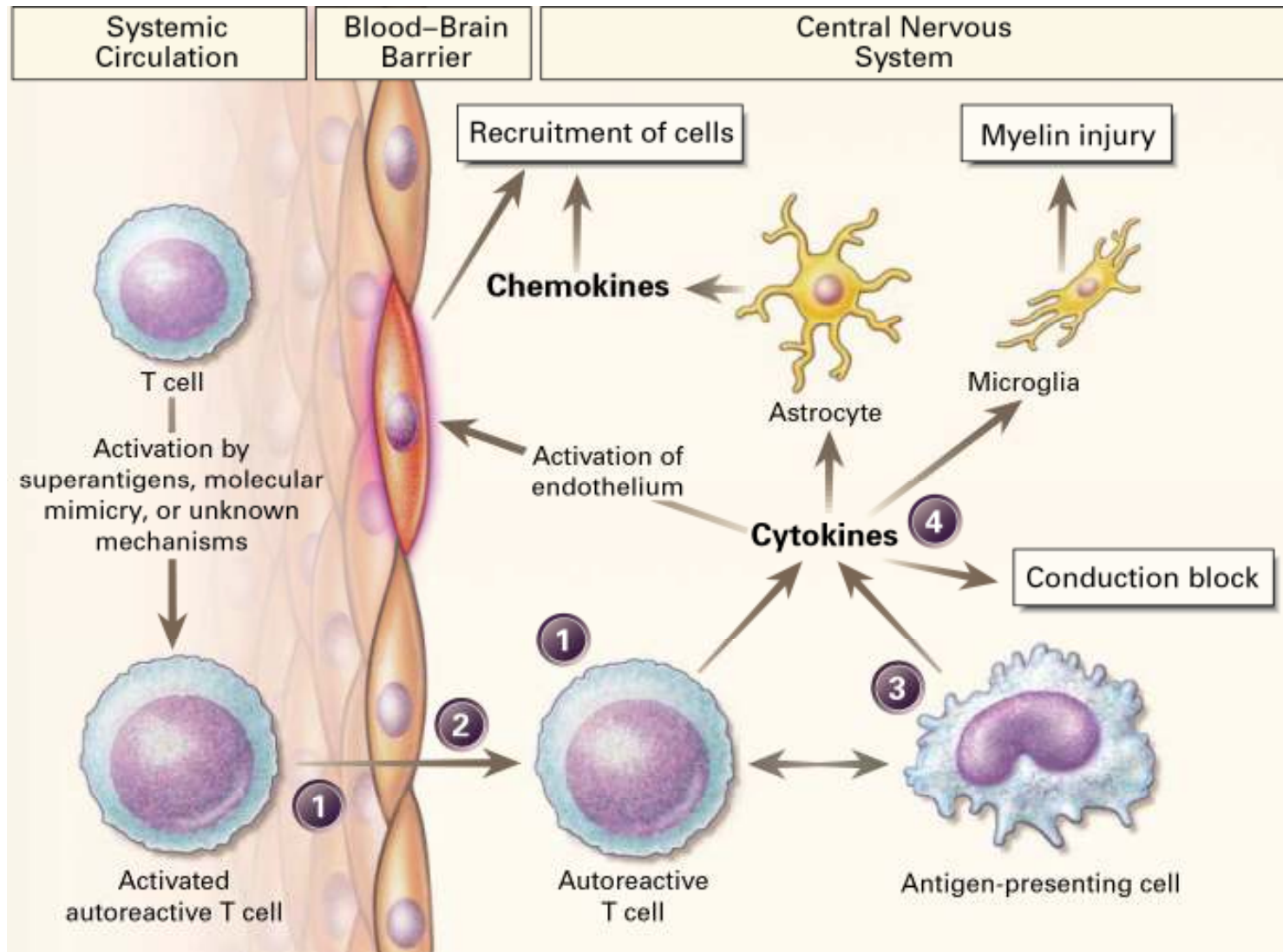


## The role of CCR6 in experimental autoimmune encephalomyelitis

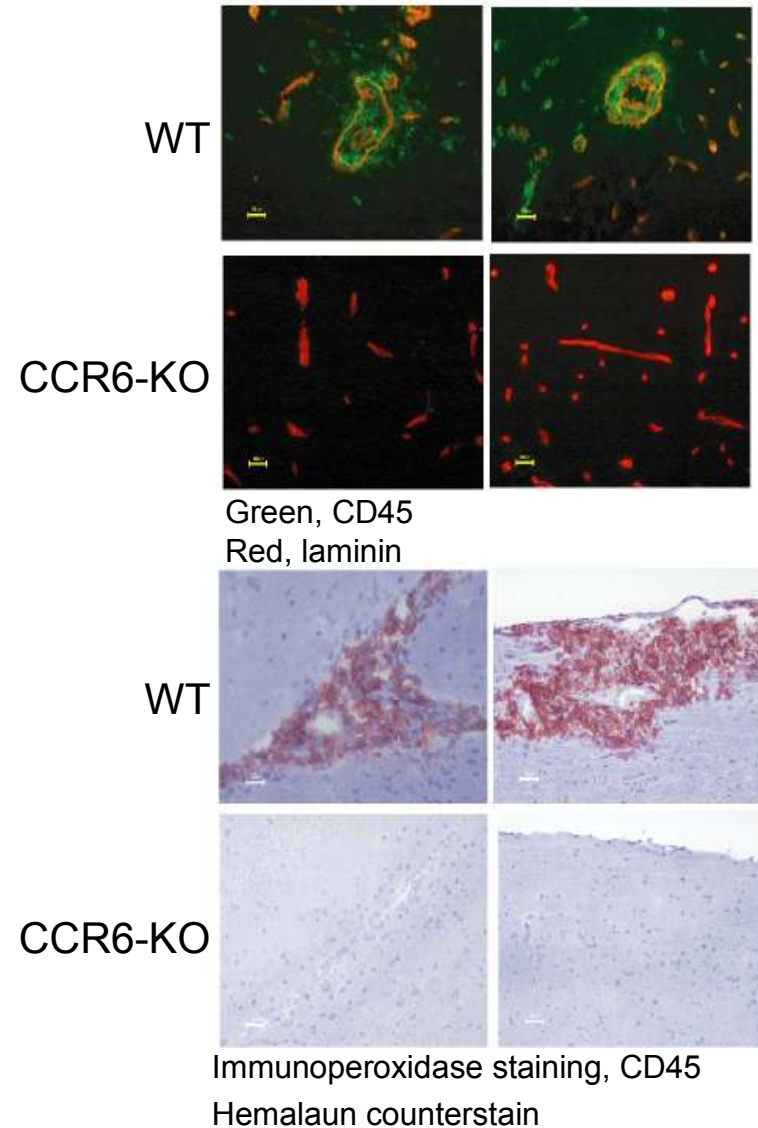
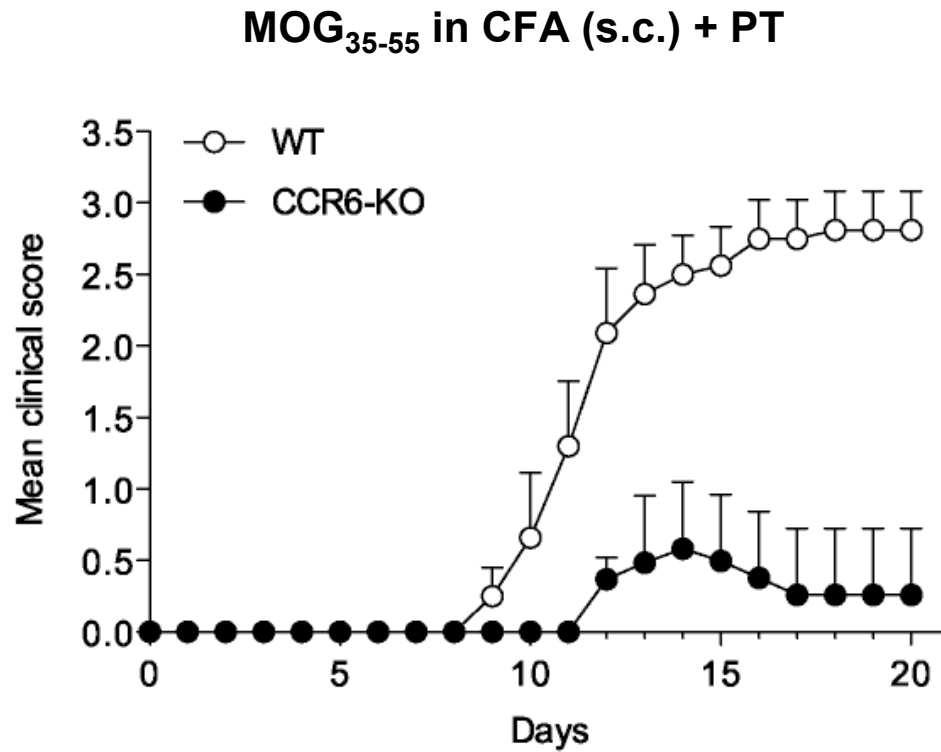
Andrea Reboldi

CCR6-KO mice (*Cook et al, Immunity 2000*)

# Pathogenesis of Multiple Sclerosis



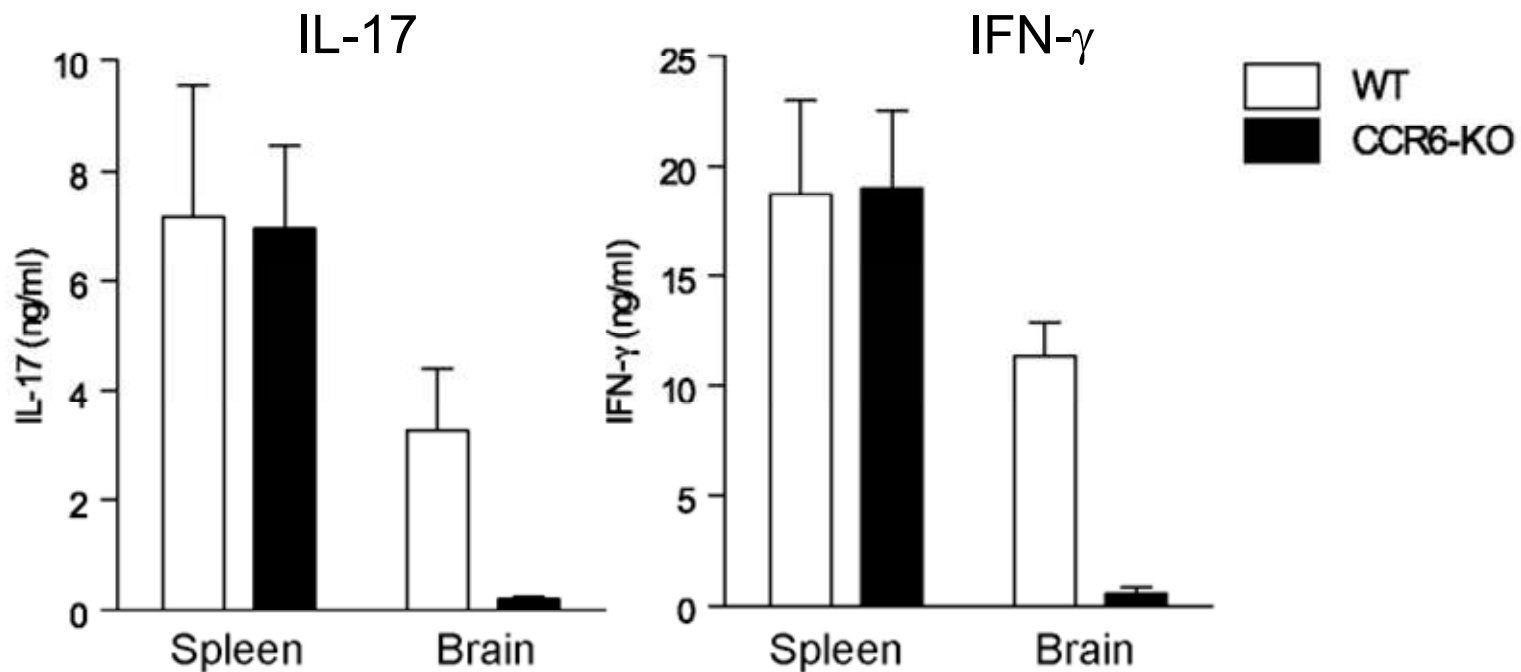
# CCR6-KO mice do not develop EAE



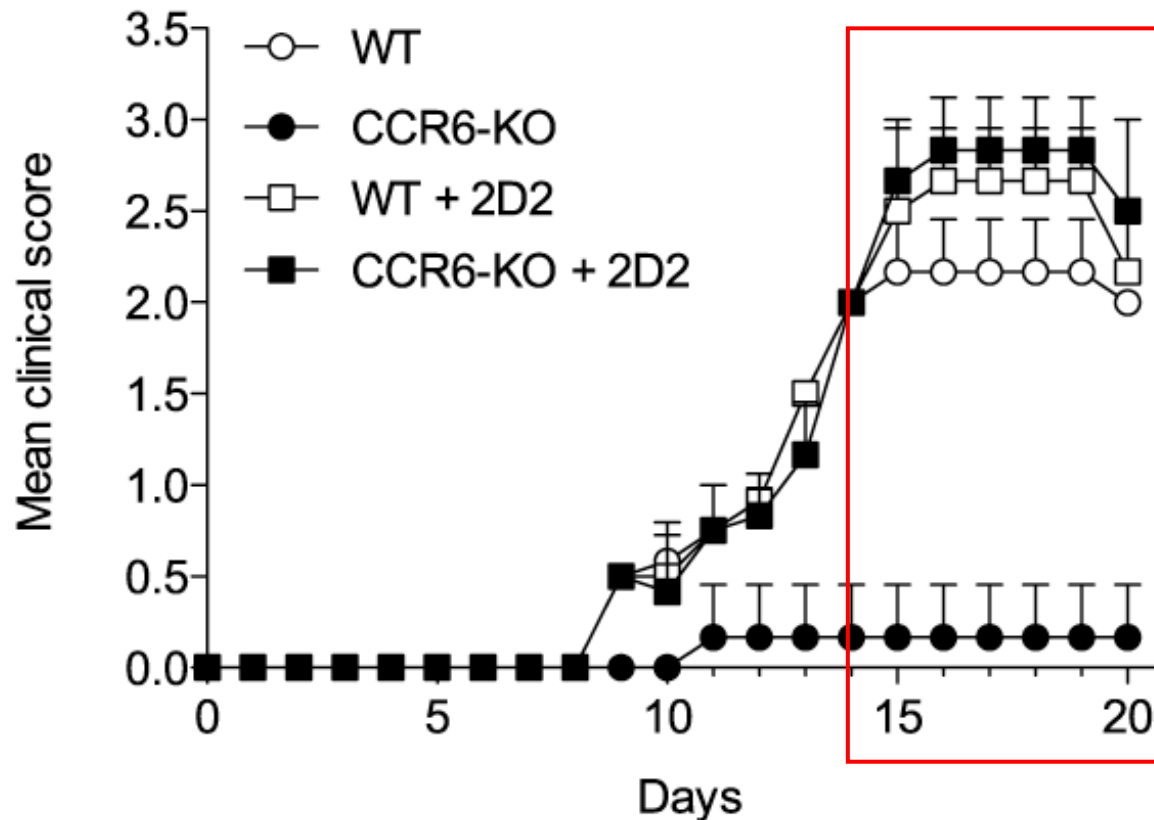
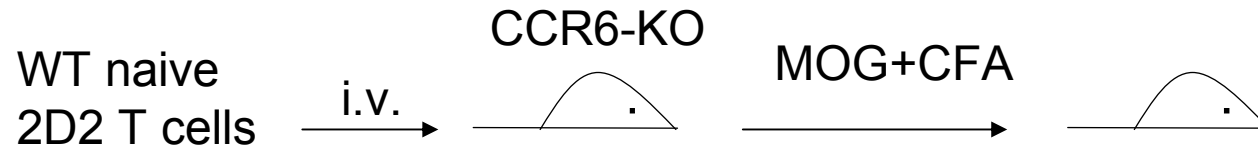
# In CCR6-KO mice MOG-reactive Th17 and Th1 cells are primed but do not migrate into the CNS

---

Cytokine production following restimulation with MOG peptide:



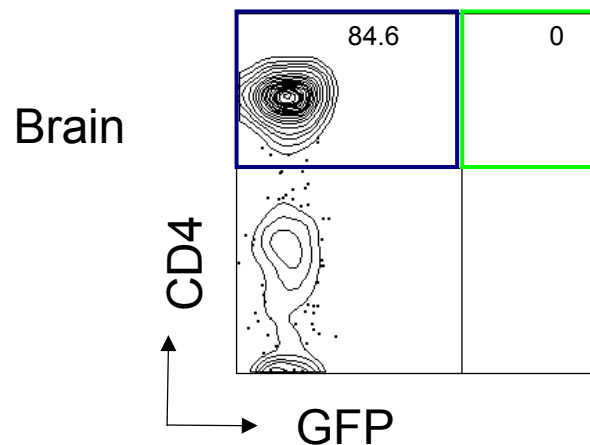
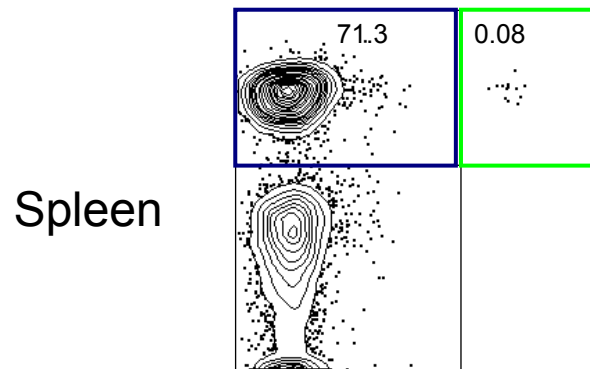
# Transfer of wild-type 2D2 T cells reconstitutes disease susceptibility in CCR6-KO mice



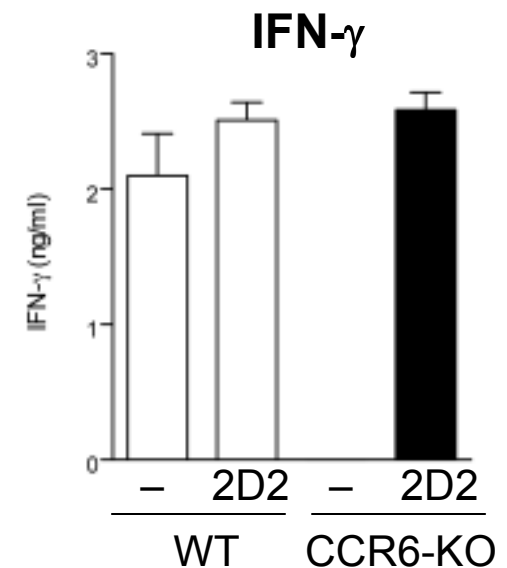
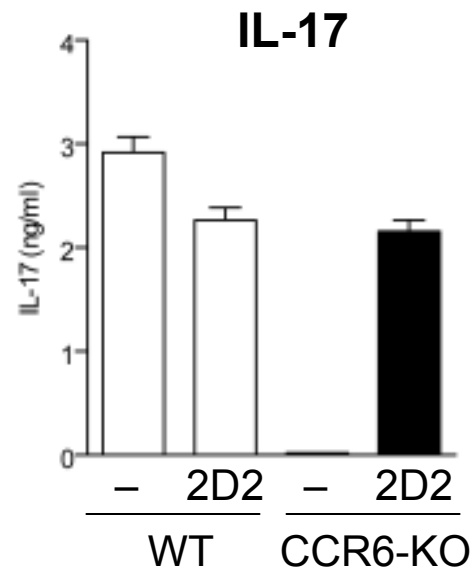


# ... but on day 20 the T cells in the CNS are endogenous CCR6<sup>-/-</sup> Th1 and Th17

CCR6-KO mice transferred with CCR6<sup>+/+</sup> GFP<sup>+</sup> 2D2 T cells (Day 20)

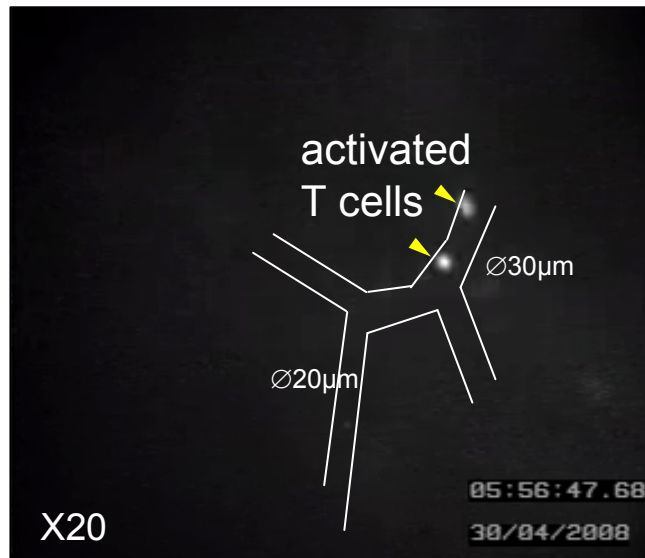


Cytokine production following restimulation with MOG peptide:

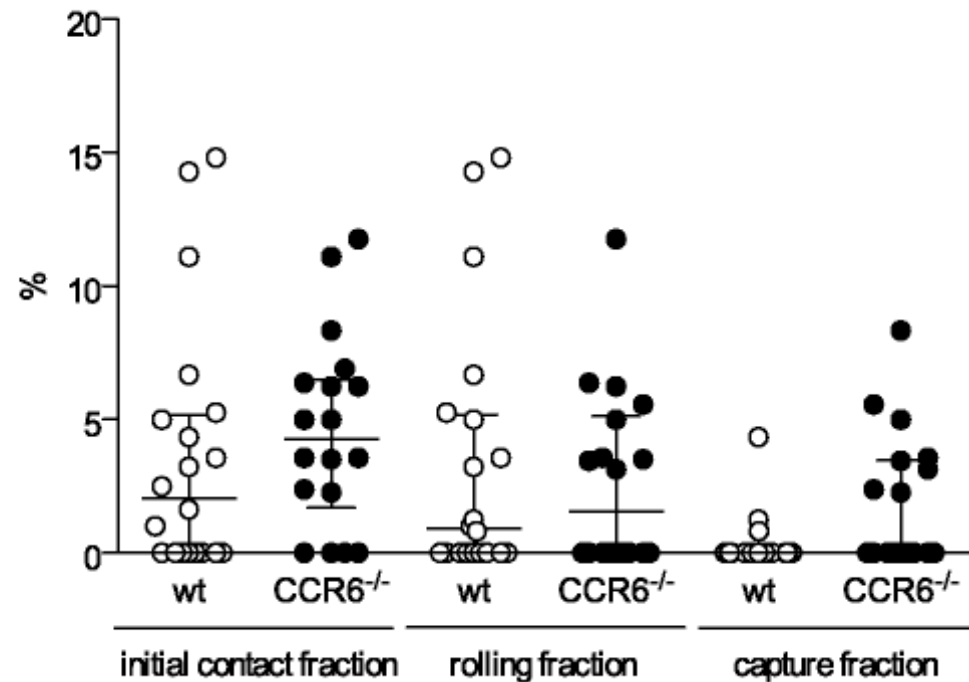


# CCR6 is not required for rolling and adhesion to inflamed endothelial cells of CNS parenchyma

## Intravital microscopy



Firmly adherent CCR6-KO T cells  
10min after injection  
in score 2 EAE mice



# CCR6<sup>+</sup> T cells as gate keepers for entry in an intact CNS?

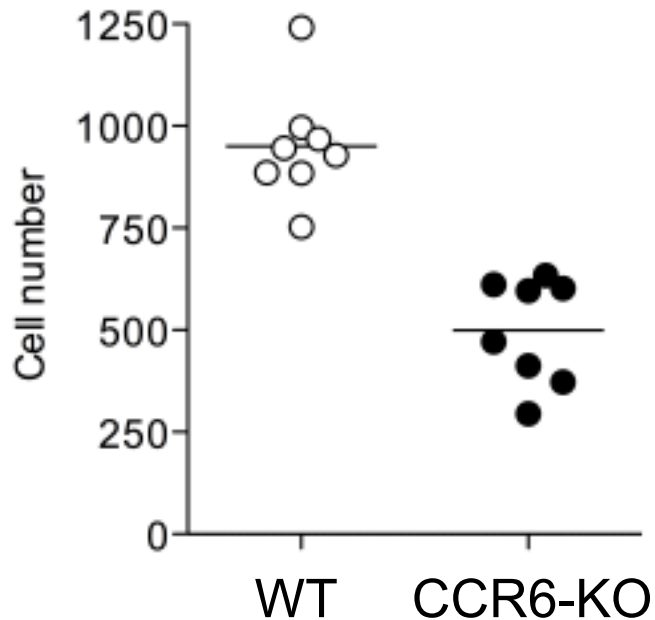
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*In EAE, effector T cells generated upon immunization have to enter a normal non inflamed brain.*

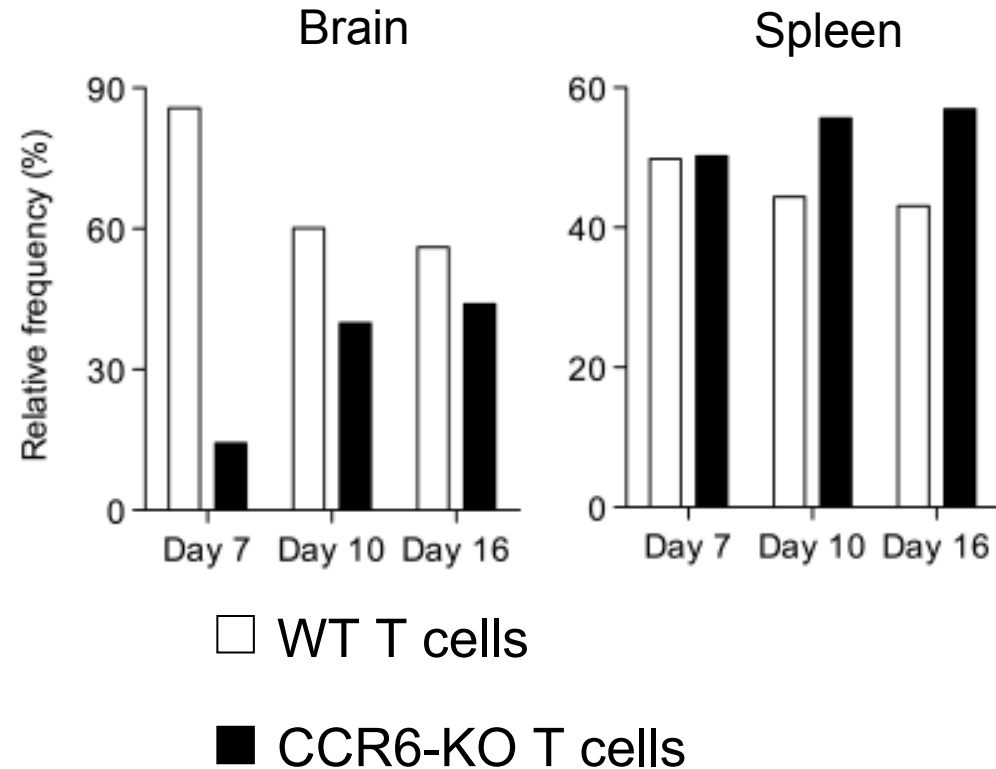
*Initial entry of CCR6<sup>+</sup> T cells by a constitutive pathway may be required to trigger subsequent recruitment of effector T cells by an inflammatory pathway through activated endothelial cells of the blood brain barrier.*

# CCR6 requirement under steady state conditions and at early time points during EAE

### Brain CD4+ T cells in steady state conditions

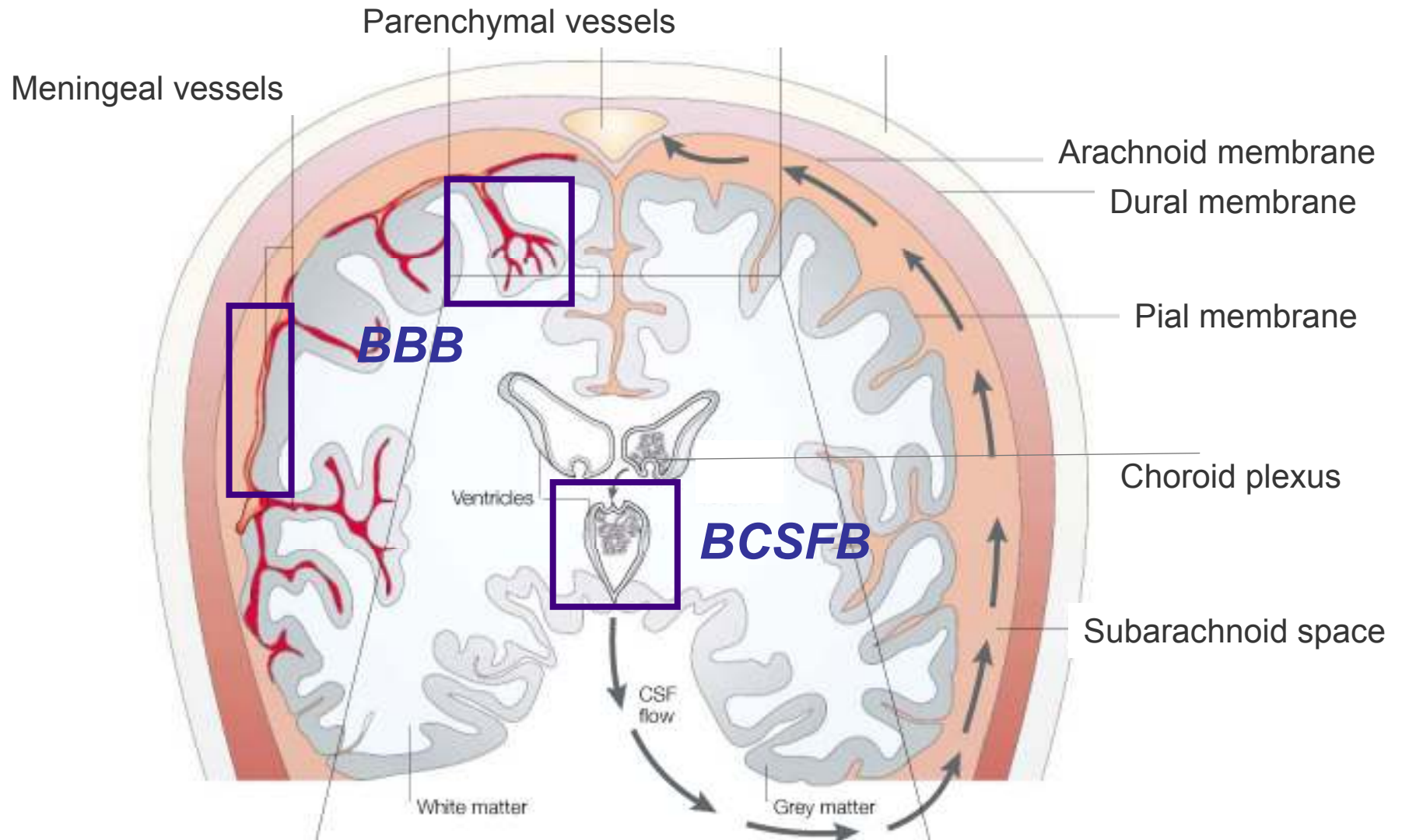


### Migration of WT and CCR6-KO T cells



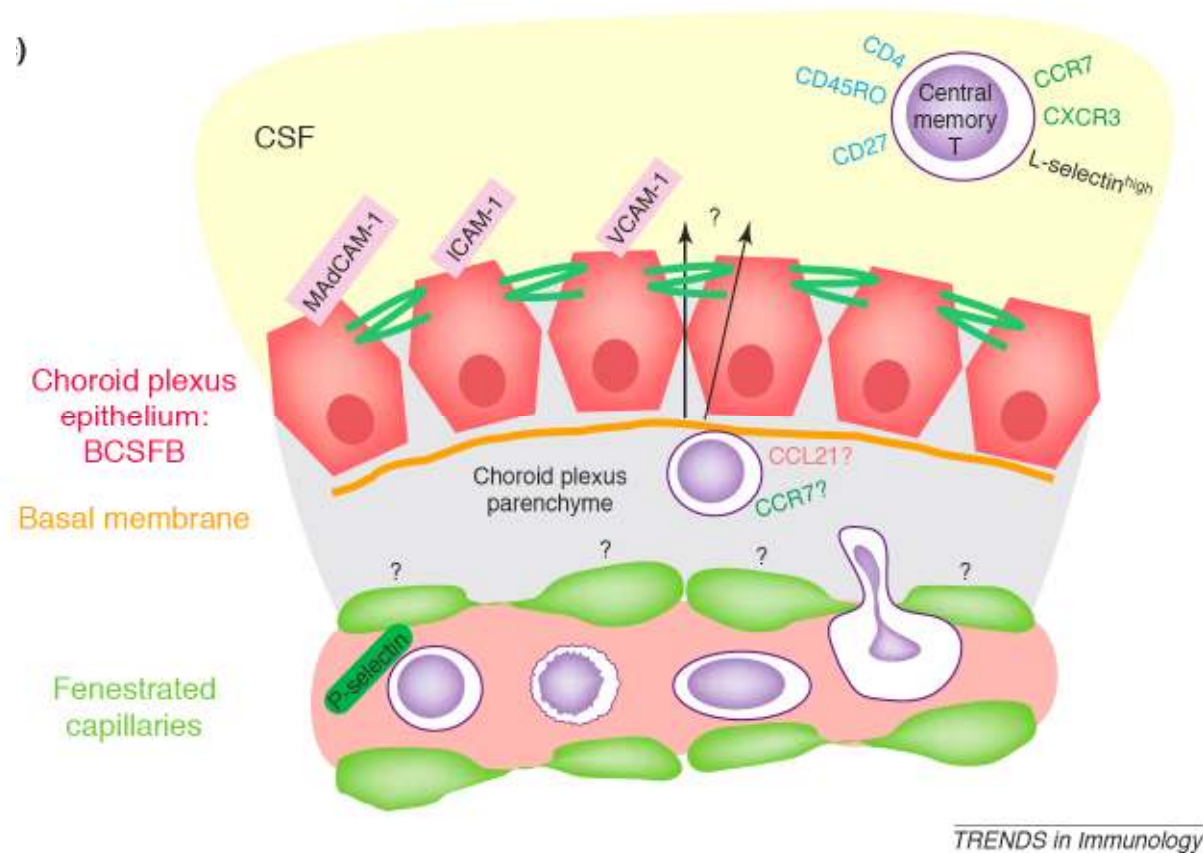
*Which is the initial port of entry of CCR6<sup>+</sup> T cells?*

# Routes for leukocyte migration into the CNS



Modified from Ransohoff et al, *Nat Rev Immunol* 2003

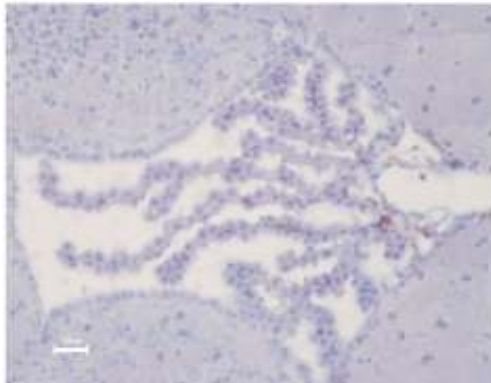
# Is the choroid plexus the port of entry of CCR6<sup>+</sup> Th17 cells?



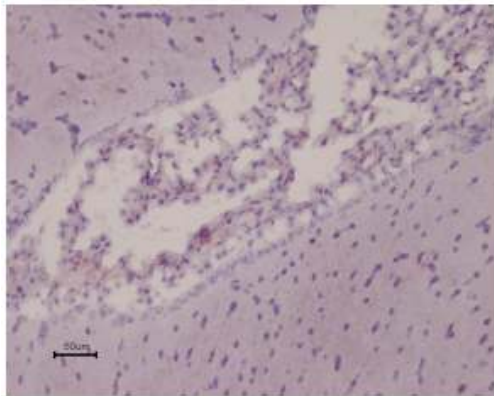
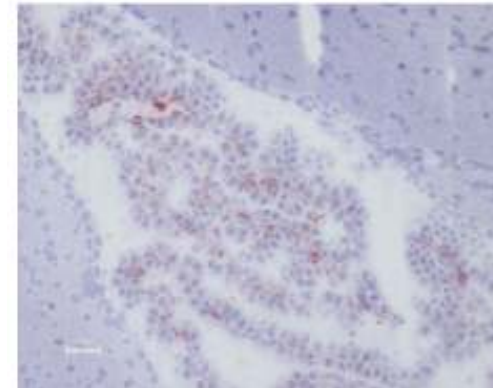
# In CCR6-KO mice immunized with MOG+CFA lymphocytes are trapped in the choroid plexus

---

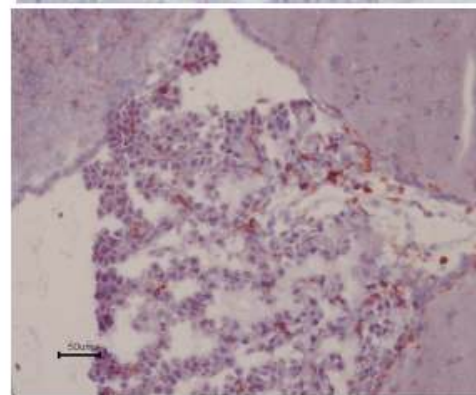
**Wild-type**



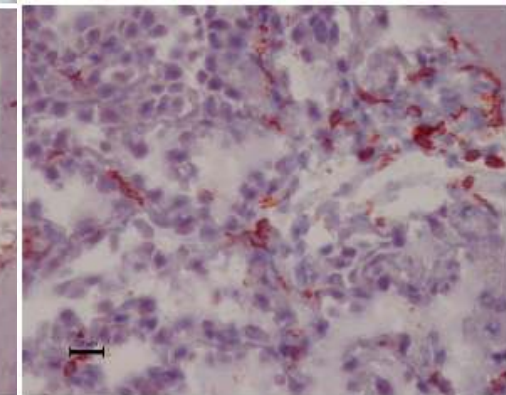
**CCR6-KO**



CD45 Ab



CD45 Ab

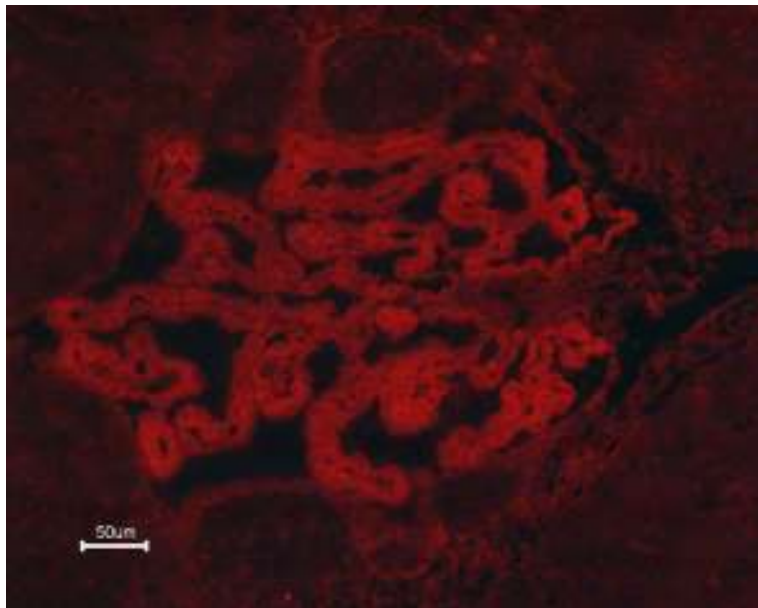




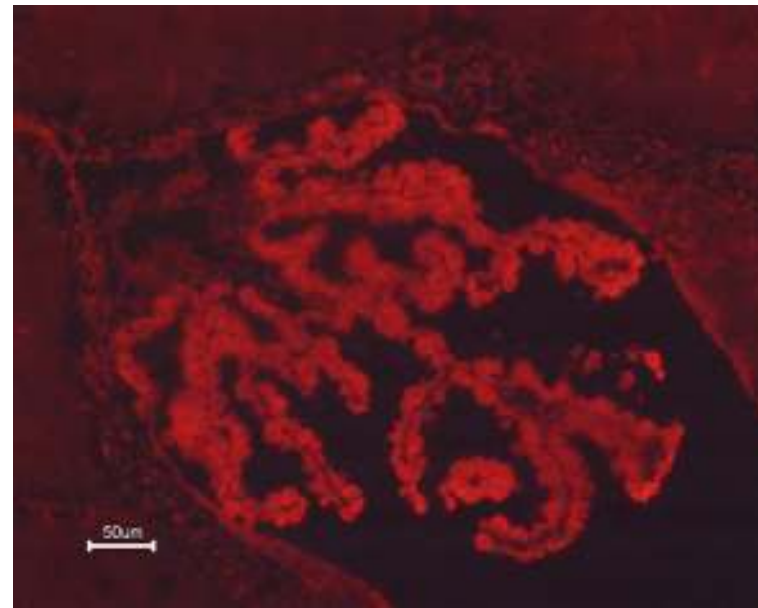
# The CCR6 ligand CCL20 is highly expressed in epithelial cells of the mouse choroid plexus

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**Wild-type**



**CCR6-KO**

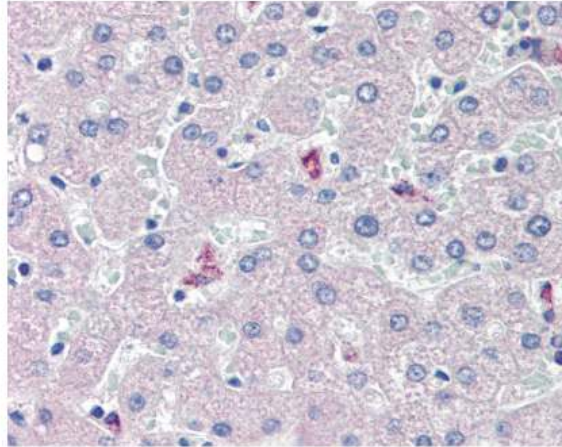


CCL20 Ab

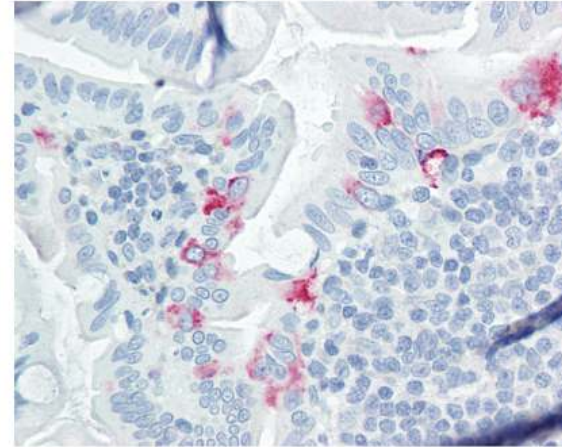
# The CCR6 ligand CCL20 (LARC) is constitutively expressed in **human** choroid epithelium

---

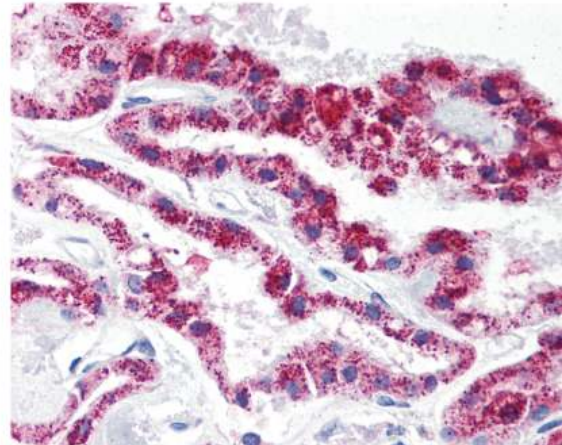
Liver



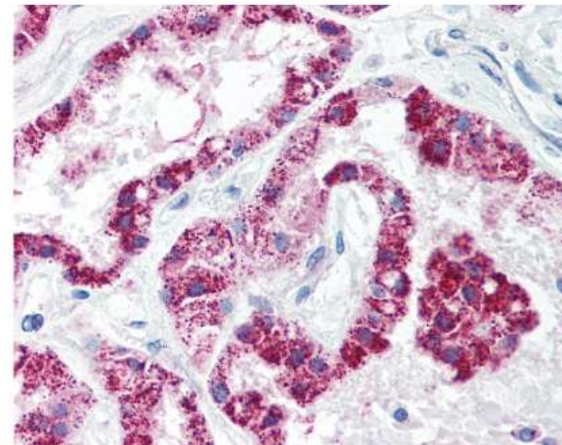
Ileum



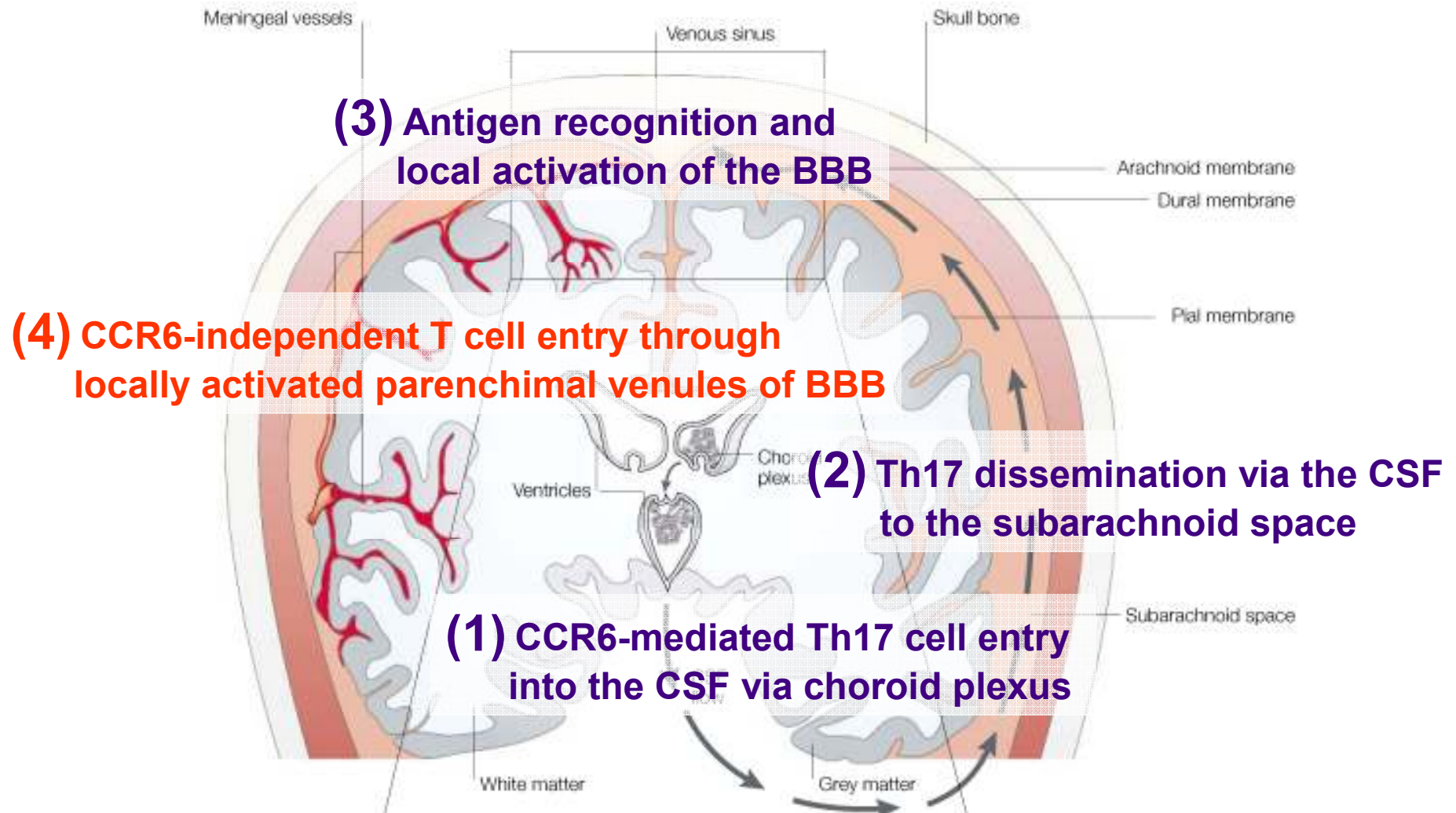
Choroid plexus



Choroid plexus



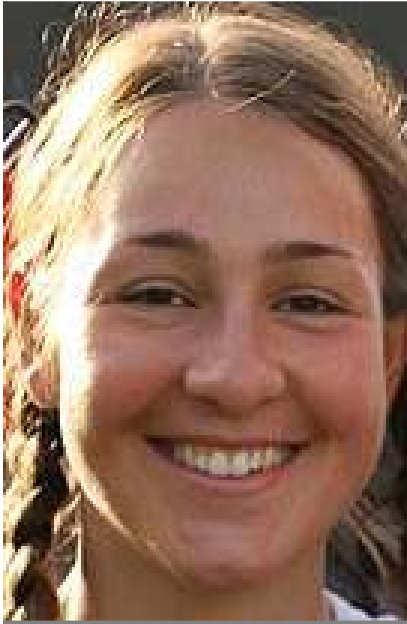
# Constitutive and **inflammatory** routes of entry into the CNS: a two step model of EAE pathogenesis



# CCR6<sup>+</sup> T cells as gate keepers for CNS entry

---

- *How do CCR6<sup>+</sup> T cells trigger leukocyte recruitment?*
- *Which are the chemokine receptors involved in late EAE?*
- *What is the relative contribution of Th17 and Th1 cells at different stages of the disease?*
- *Does CCR6-blockade have any therapeutic effect (relapsing-remitting SJL model)?*
- *Do CCR6<sup>+</sup> T cells play a role in surveillance of the CNS?*



## Human T cell repertoire analysis using amplified T cell libraries

Rebekka Geiger

# Challenges in analyzing the human naïve T cell repertoire

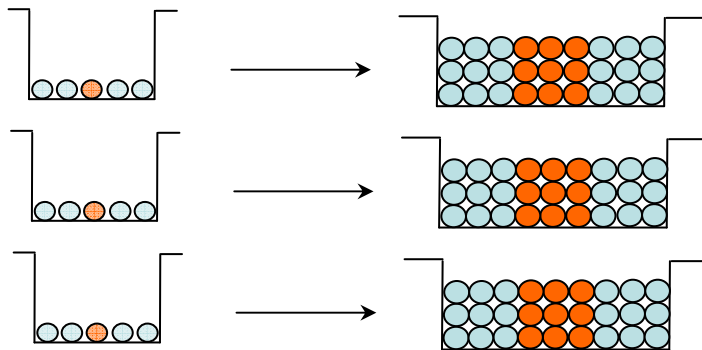
- Low frequency of antigen-specific naïve T cells
- High activation threshold of naïve T cells
- Broad spectrum of avidities
- Limitations of peptide-based and tetramer-based approaches
- Need to measure T cell responses to complex naturally processed antigens

# Analysis of naïve and memory T cell repertoires using amplified T cell libraries

## *1<sup>st</sup> step: Amplification*

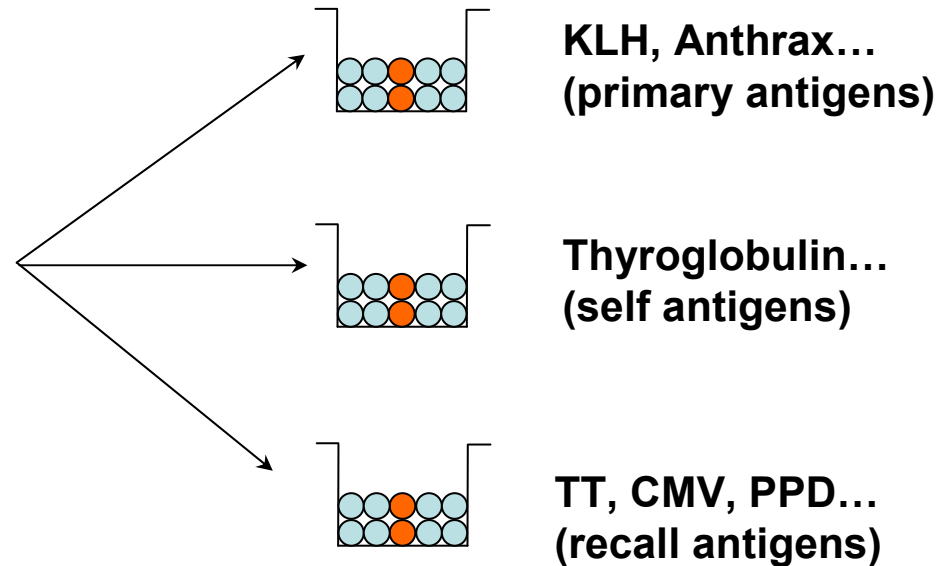
Multiple cultures (96 up to 384)  
each containing 1000 cells  
are expanded with PHA and IL-2

1 naïve T cell → 5,000 T cell blasts



## *2<sup>nd</sup> step: Interrogation*

Each culture is tested  
for proliferation in response  
to Ag and autologous APC

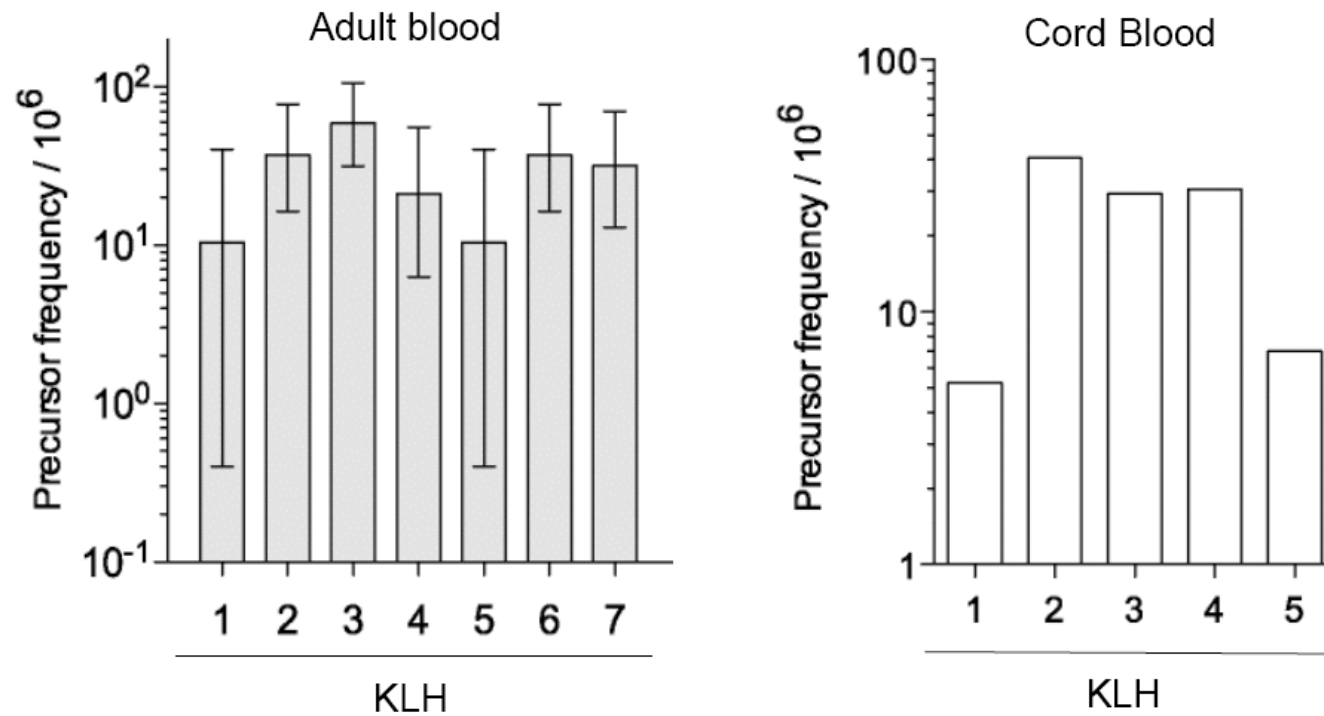




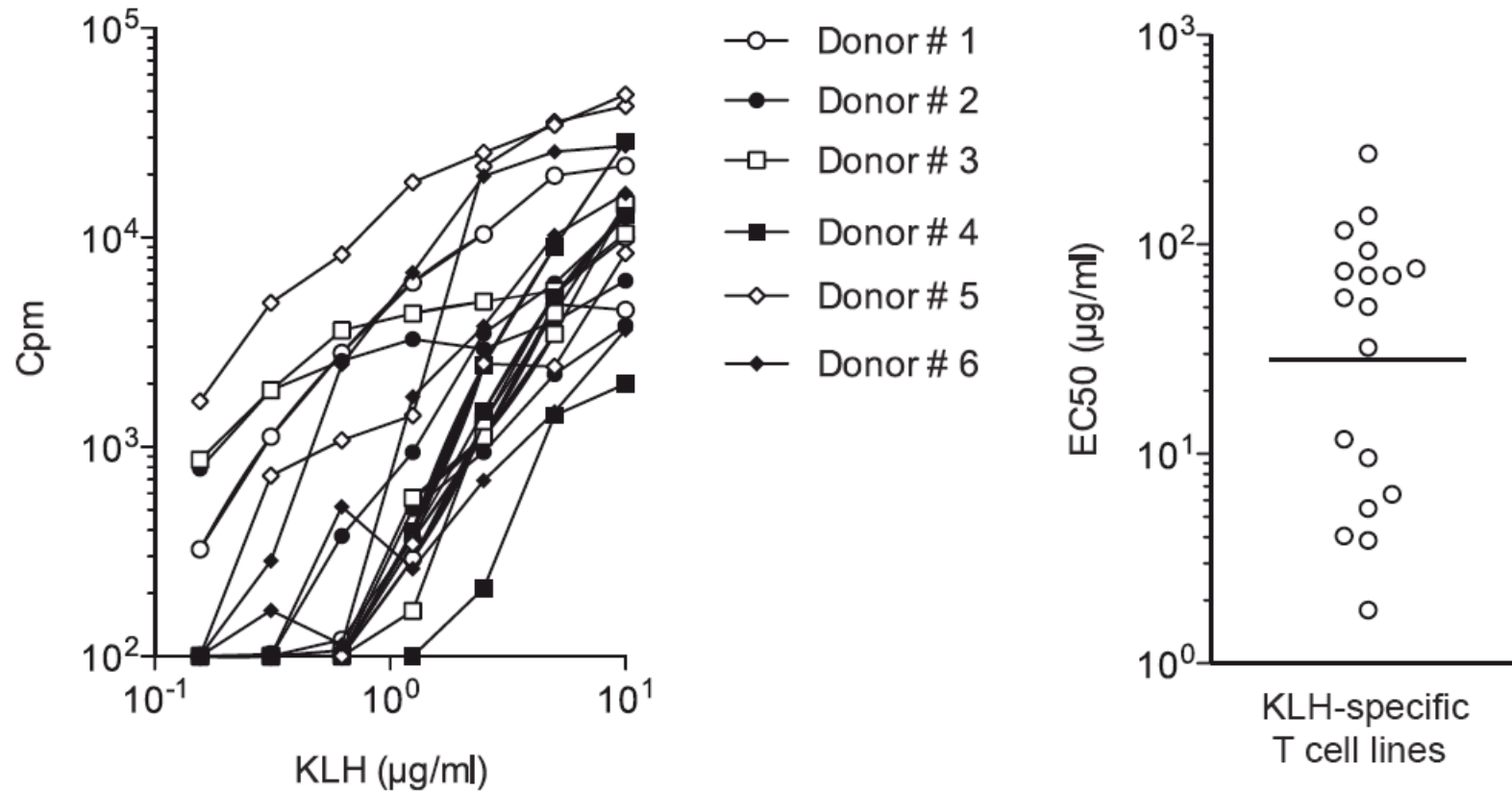


# Frequency of KLH-specific naïve T cells

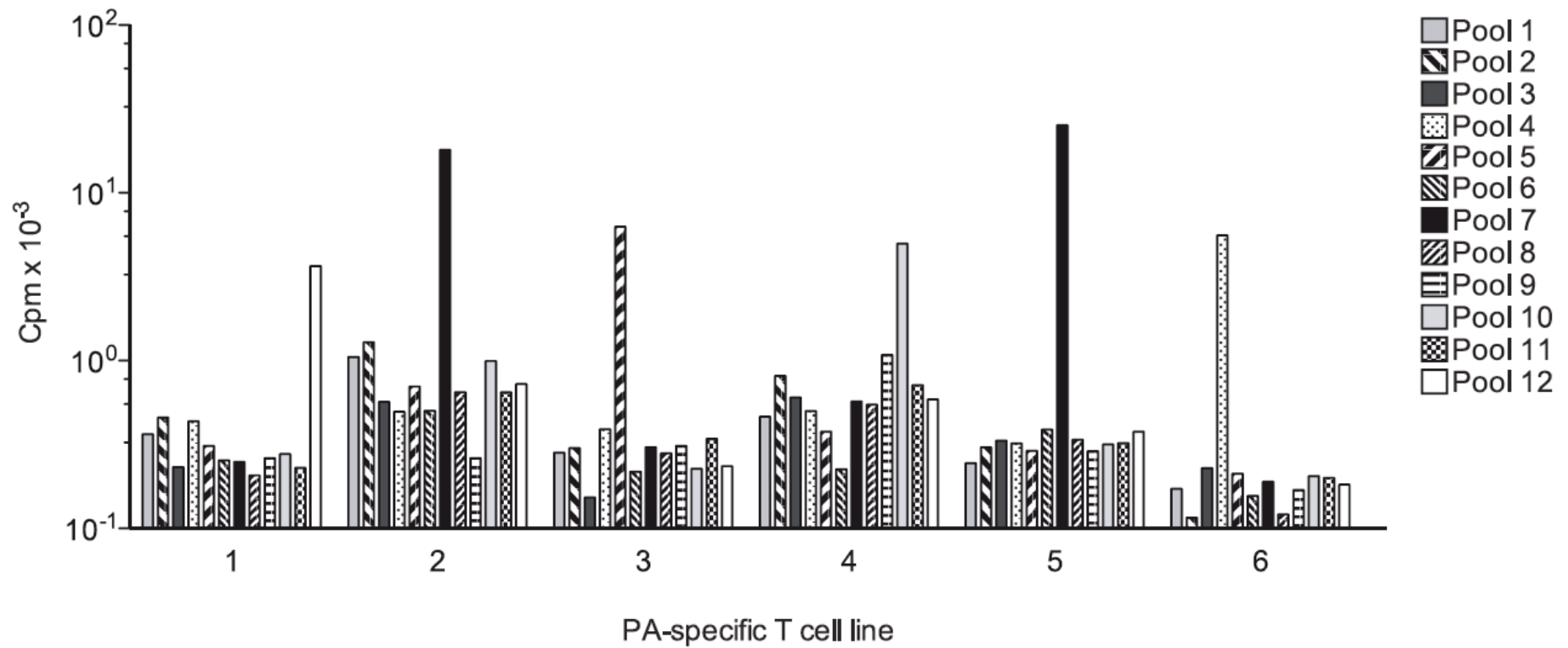
“Amplified T cell libraries” from naïve CD45RA<sup>+</sup> CD45RO<sup>-</sup> CCR7<sup>+</sup> CD4<sup>+</sup> T cells



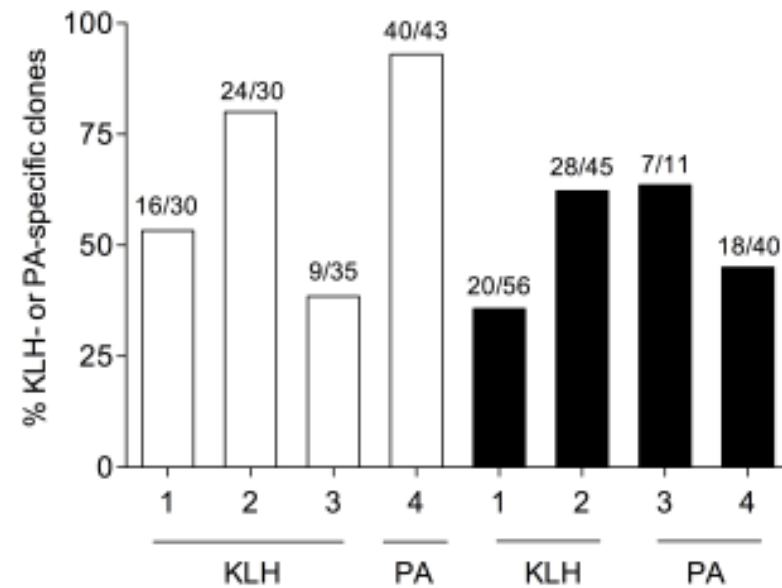
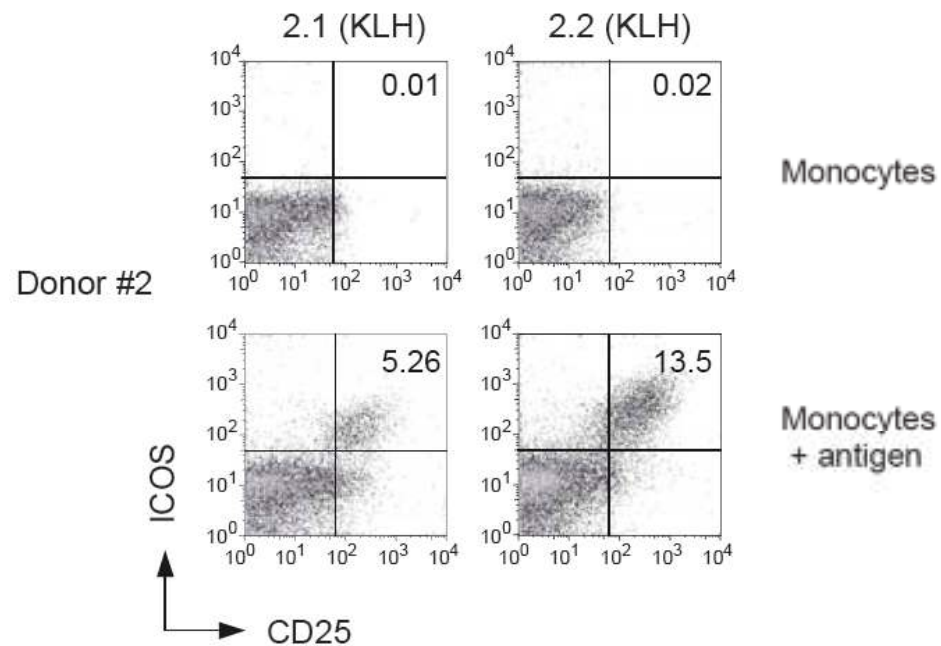
# Broad range of responsiveness



# Broad range of epitope specificities

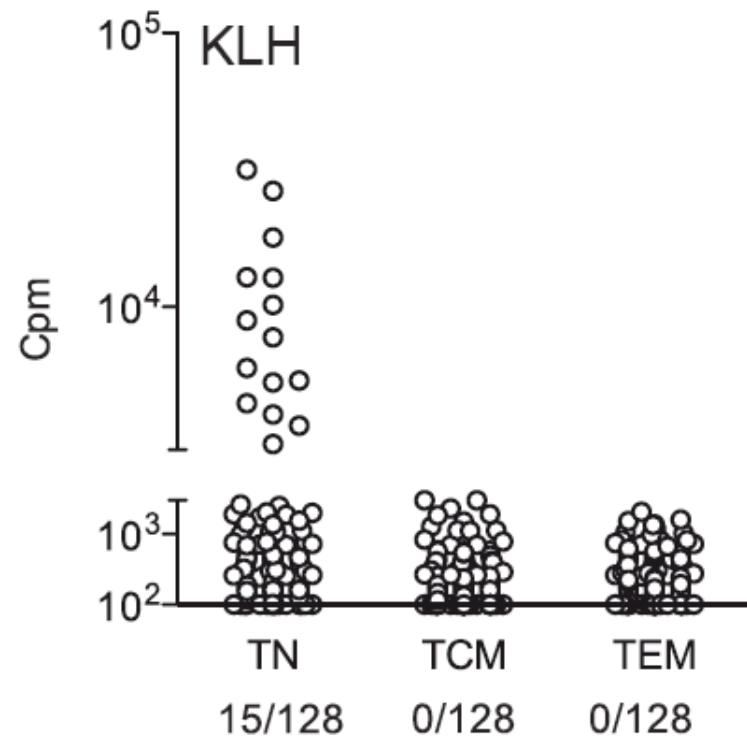


# Isolation of Ag-specific T cell clones from the naïve repertoire

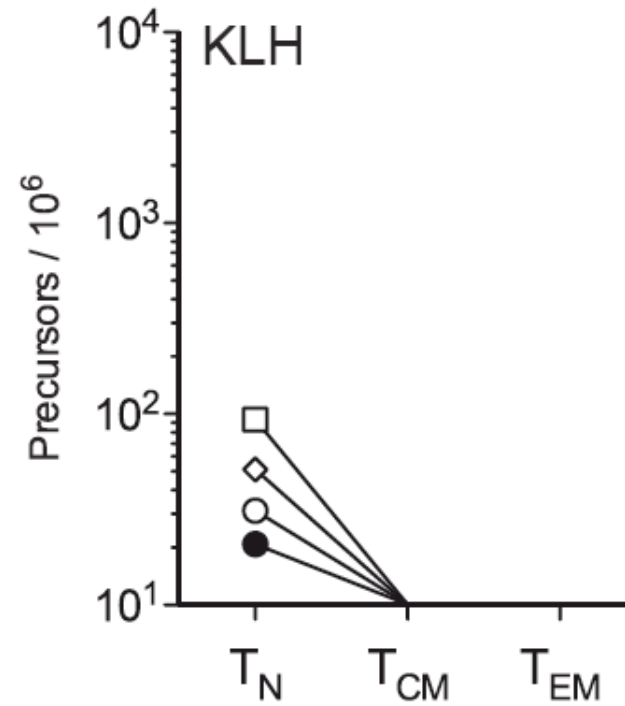


# Analysis of memory repertoires

**C**

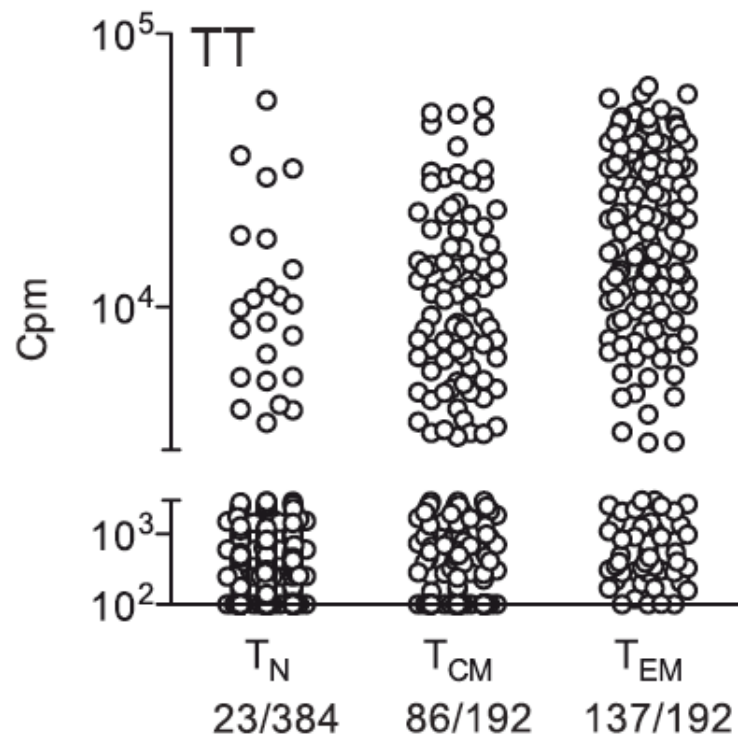


**D**

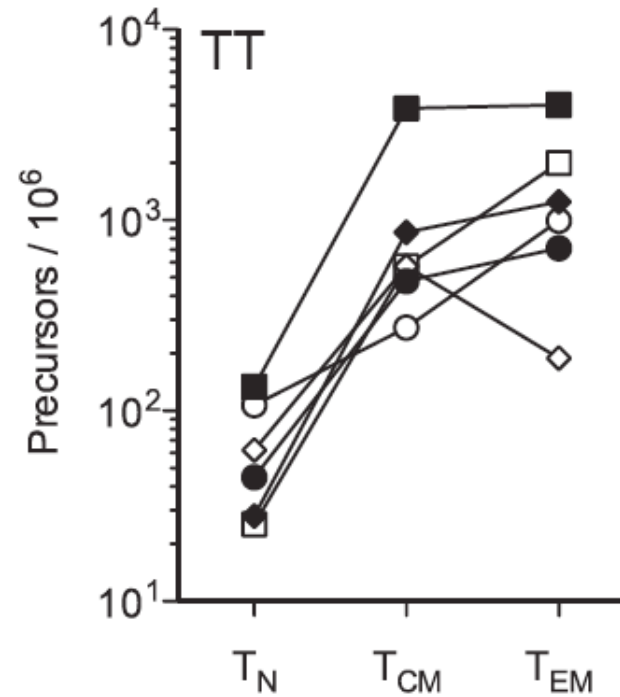


# Increased frequencies in the memory compartment (Tet Tox)

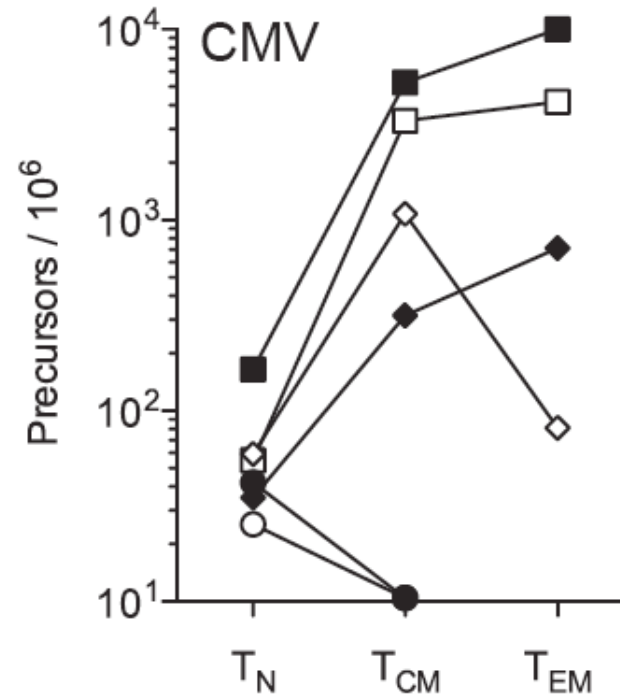
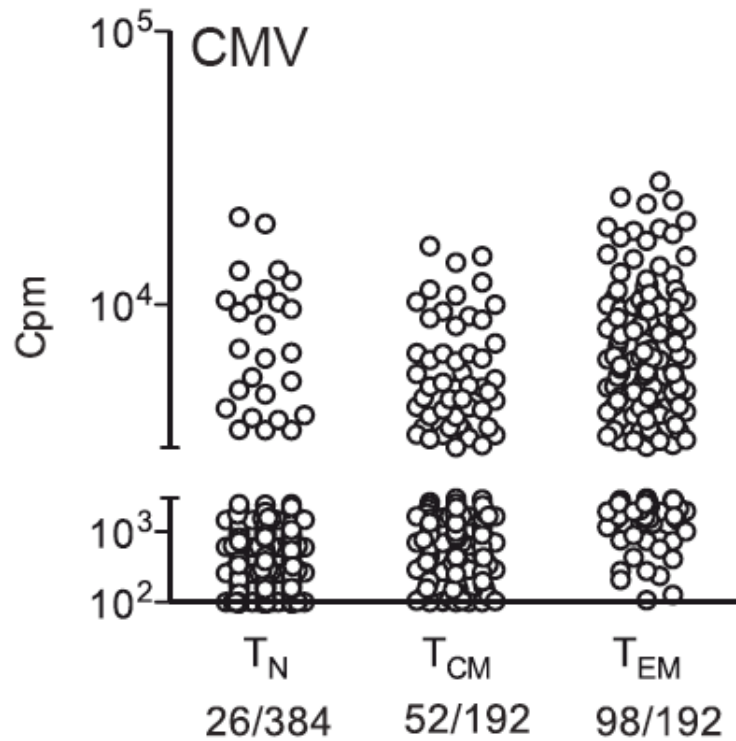
**A**



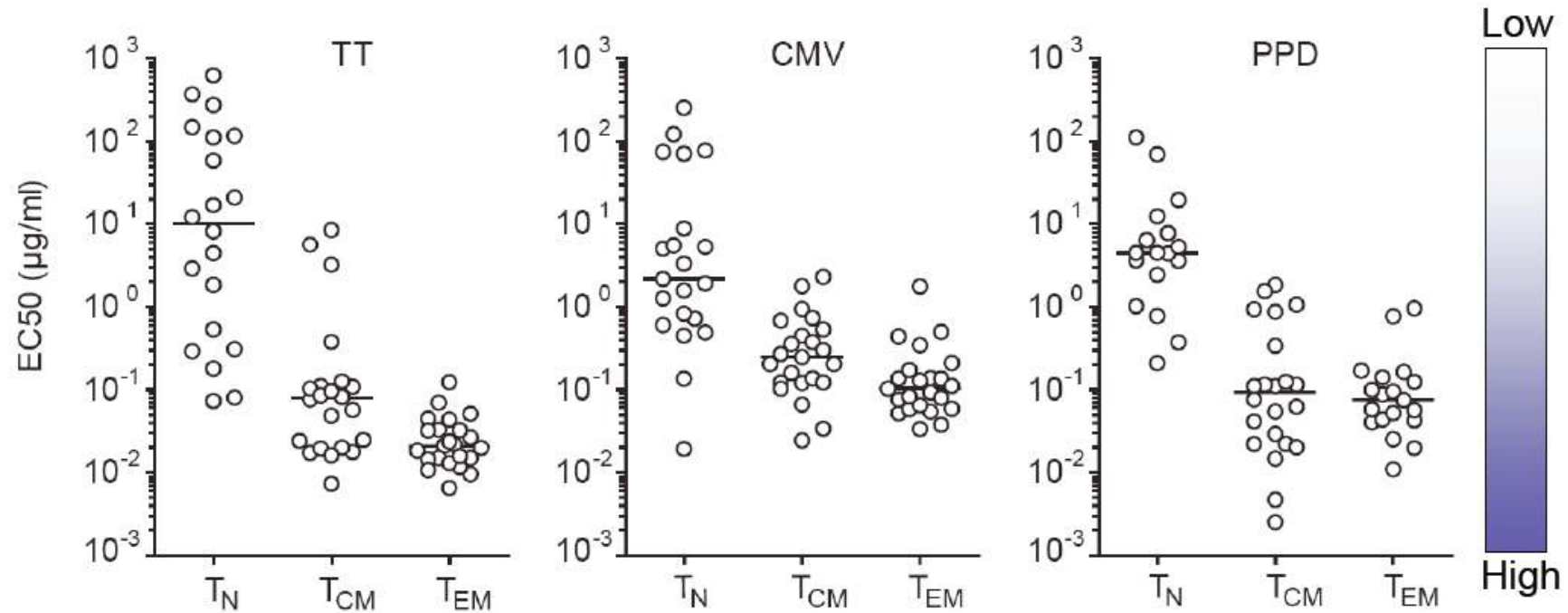
**B**



# Increased frequencies in the memory compartment (CMV)



# Selection of high avidity T cells in the memory pool





# Applications

- Predict antigenicity of complex molecules (even whole pathogens)
- Assess immunocompetence (elderly, HIV)
- Assess memory in different T cell subsets
- Generate T cells for cellular immunotherapy

## The Lanzavecchia & Sallusto Lab

**Giorgio Napolitani** (Th17)

**Andrea Reboldi** (EAE)

**Thomas Duhén** (Th22)

**Rebekka Geiger** (T cell libraries)

**Martina Beltramello** (dengue)

**Davide Corti** (influenza)

**David Jarrossay** (plasma cells)

Dirk Baumjohann

Annalisa Macagno

Janine Stubbs

Debora Pinna

Blanca Fernandez

Chiara Silacci

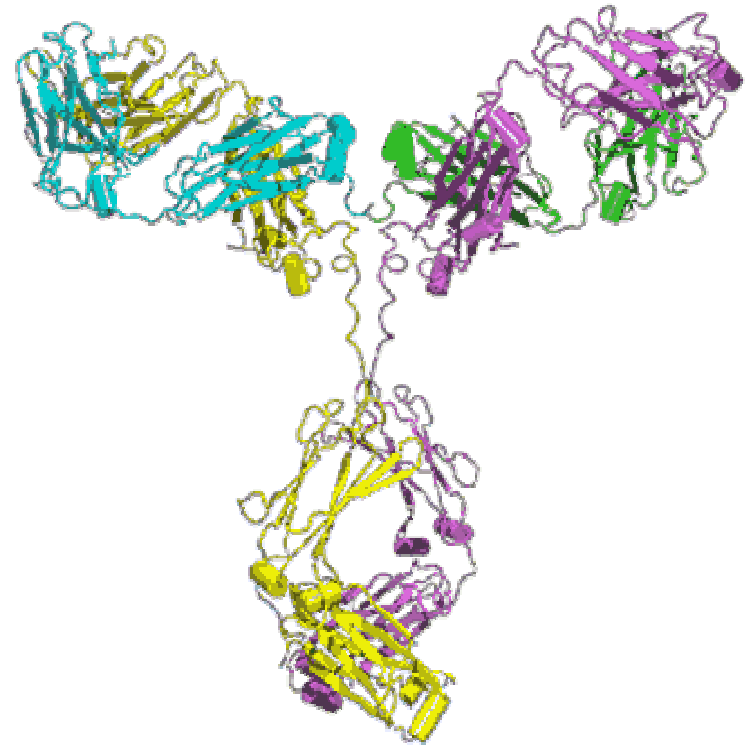
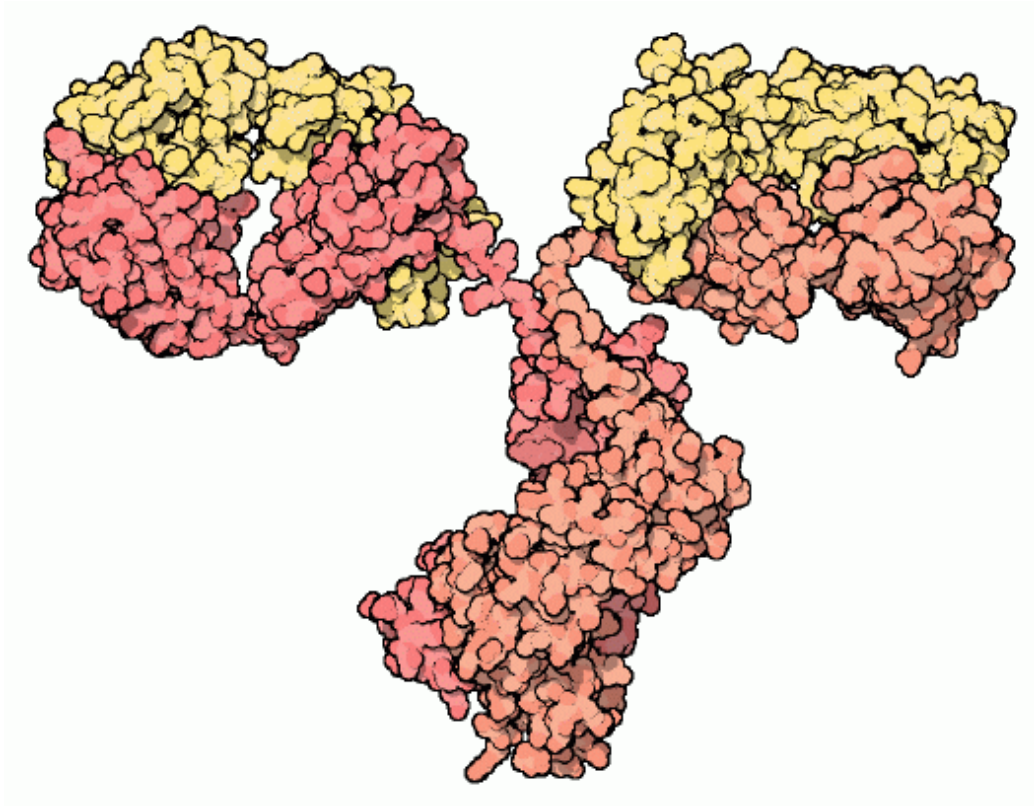
Fabrizia Vanzetta

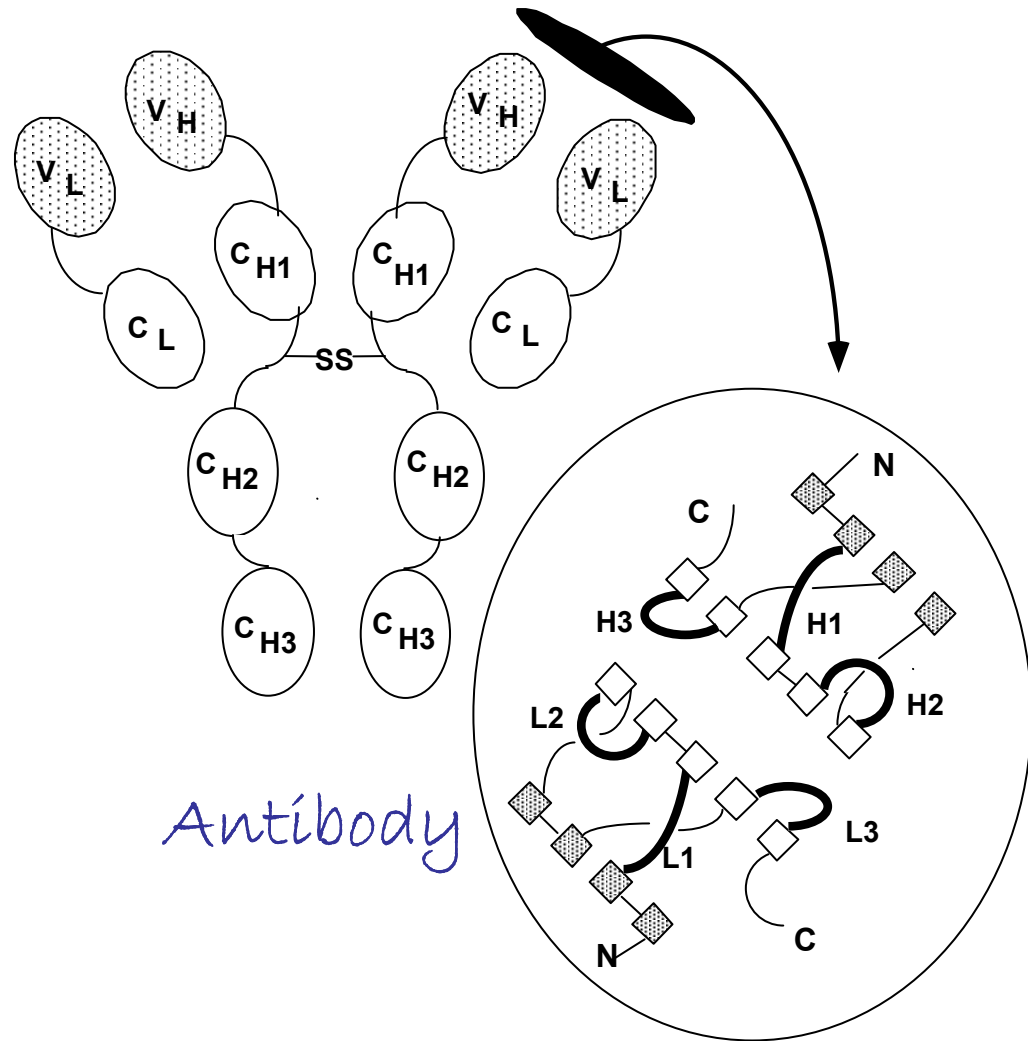
Eva Acosta-Rodriguez, Cordoba - Jens Geginat, Berlin -

Mara Messi, Basel - Amanda Gett, Basel - Laura Rivino,

Berlin

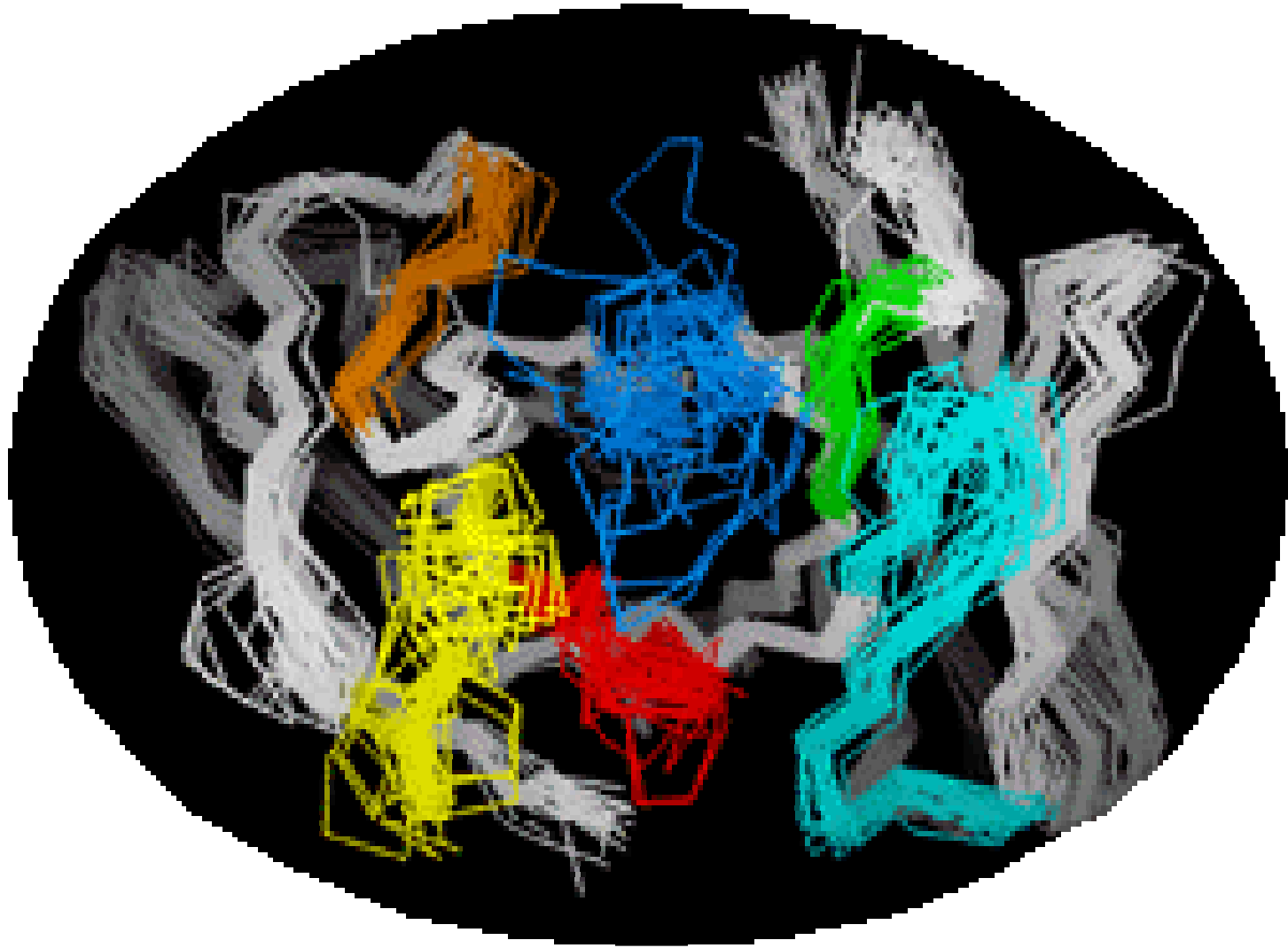






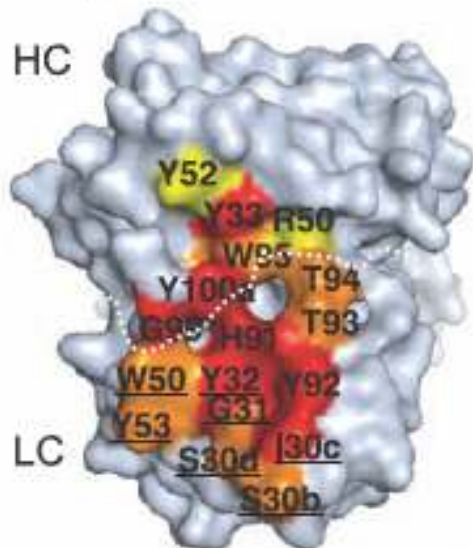
Antibody

Antigen binding site

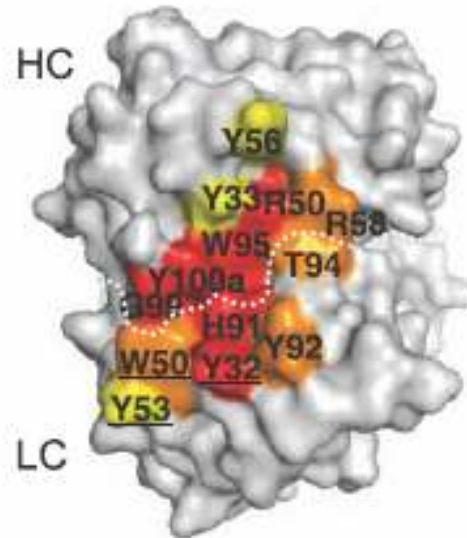


# Multispecificity / Crossreactivity / Promiscuity / Degeneracy

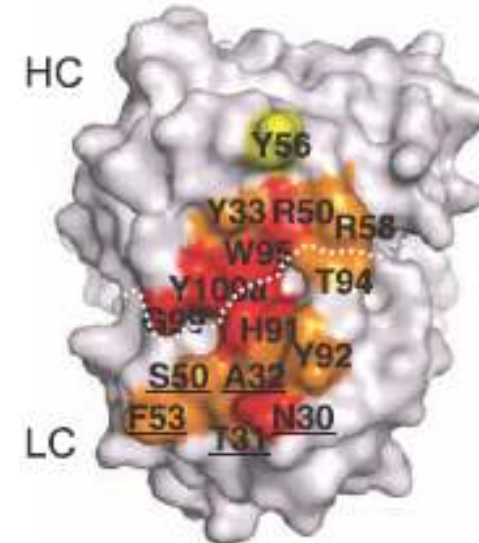
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bH1 (VEGF bound)



bH1 (HER2 bound)



Herceptin (HER2 bound)

# Antiviral activities of antibodies

Average number  
of antibodies  
bound per virion  
at neutralization = 4

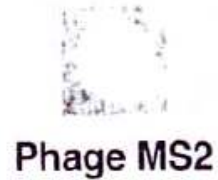
4-5

38

70

225

Antibody



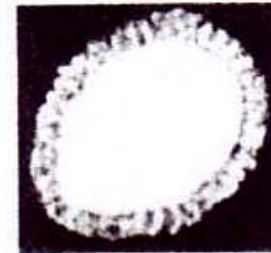
Phage MS2



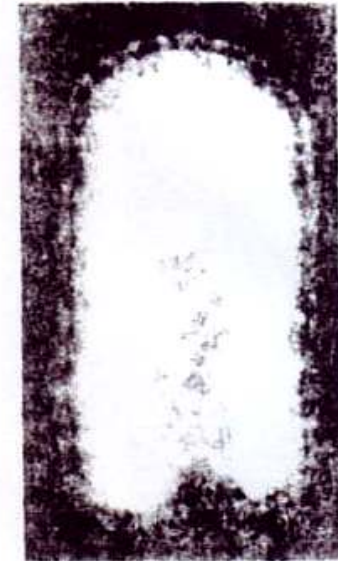
Poliovirus



Papillomavirus



Influenza Virus A

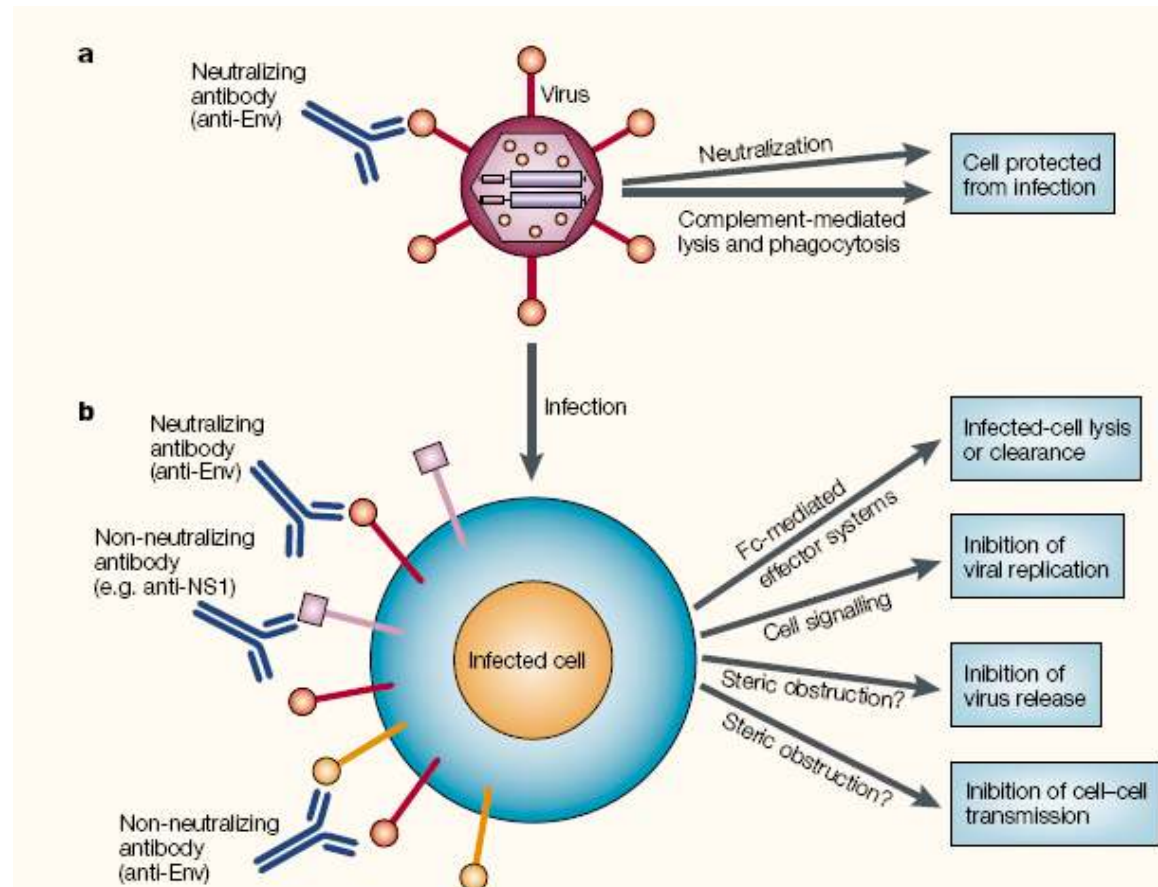


Rabies Virus

100 nm

**affinity / avidity / accessibility**

# Antiviral activities of antibodies





# The cellular basis of immunological memory

---

## “Effector memory”

**Plasma cells (Ab)**

**Effector memory T cells**

- Fully differentiated
- In peripheral tissues
- Immediate protection

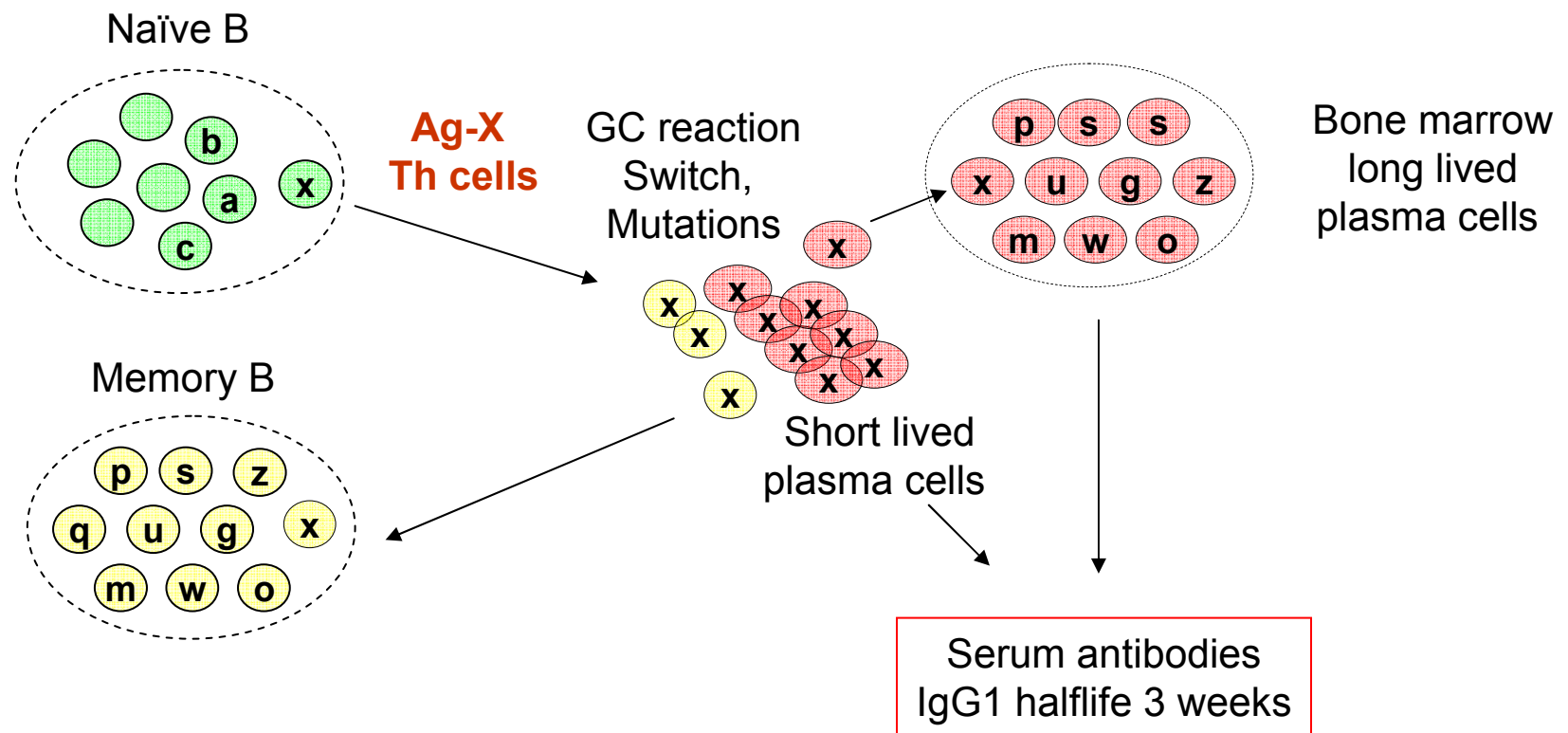
## “Central memory”

**Memory B cells**

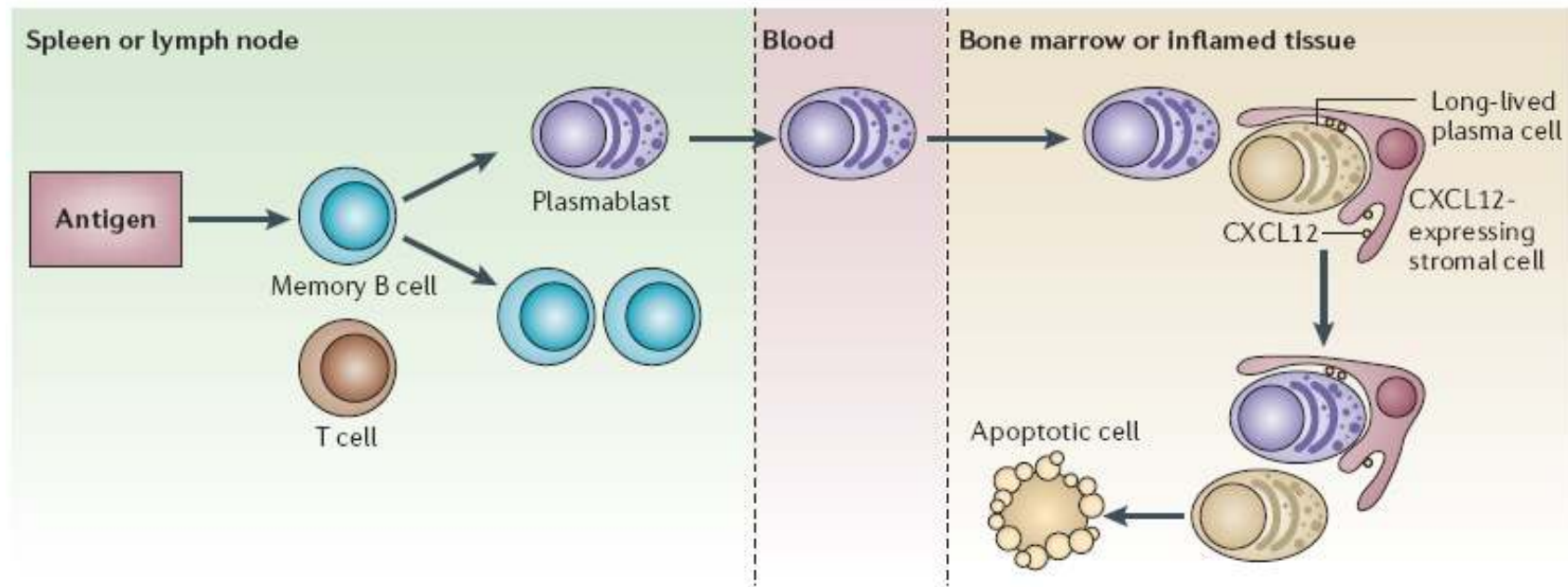
**Central memory T cells**

- Differentiation intermediates
- In secondary lymphoid organs
- Recall responses

# B cell clonal dynamics: primary responses

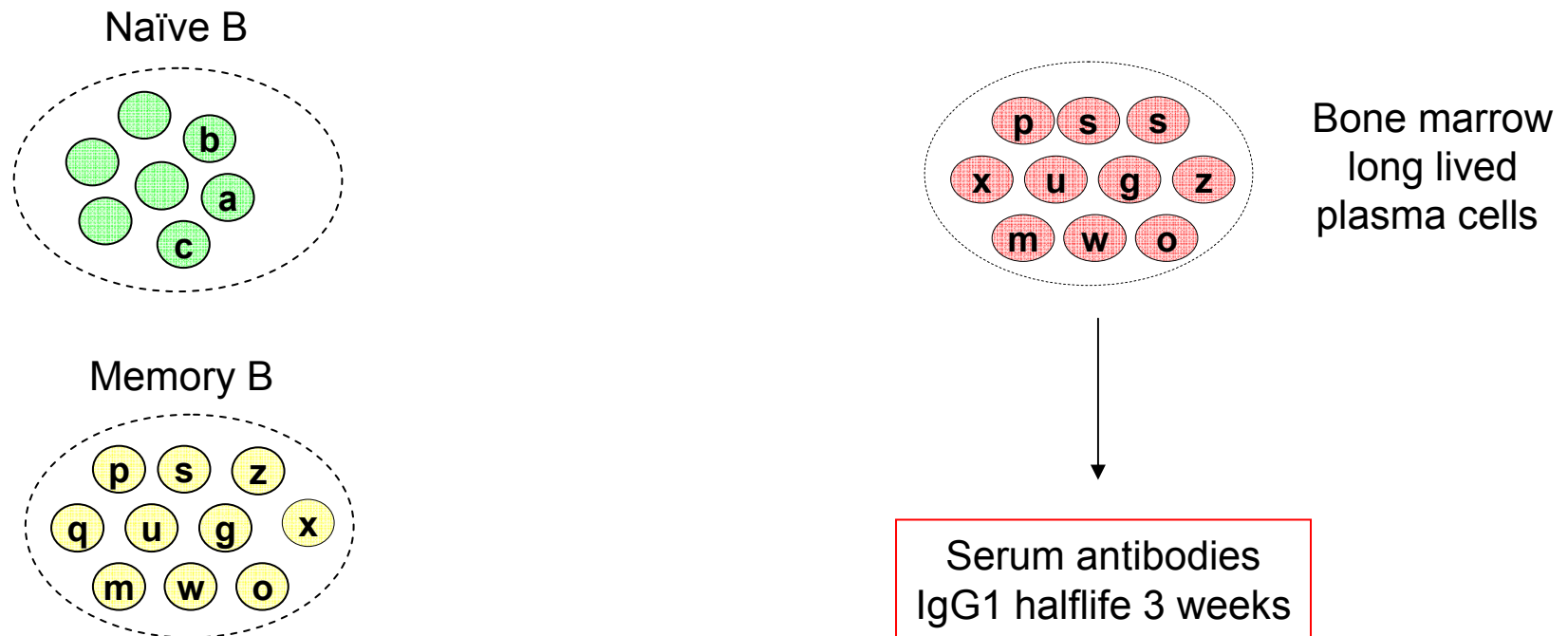


# Bone marrow niches for long lived plasma cells



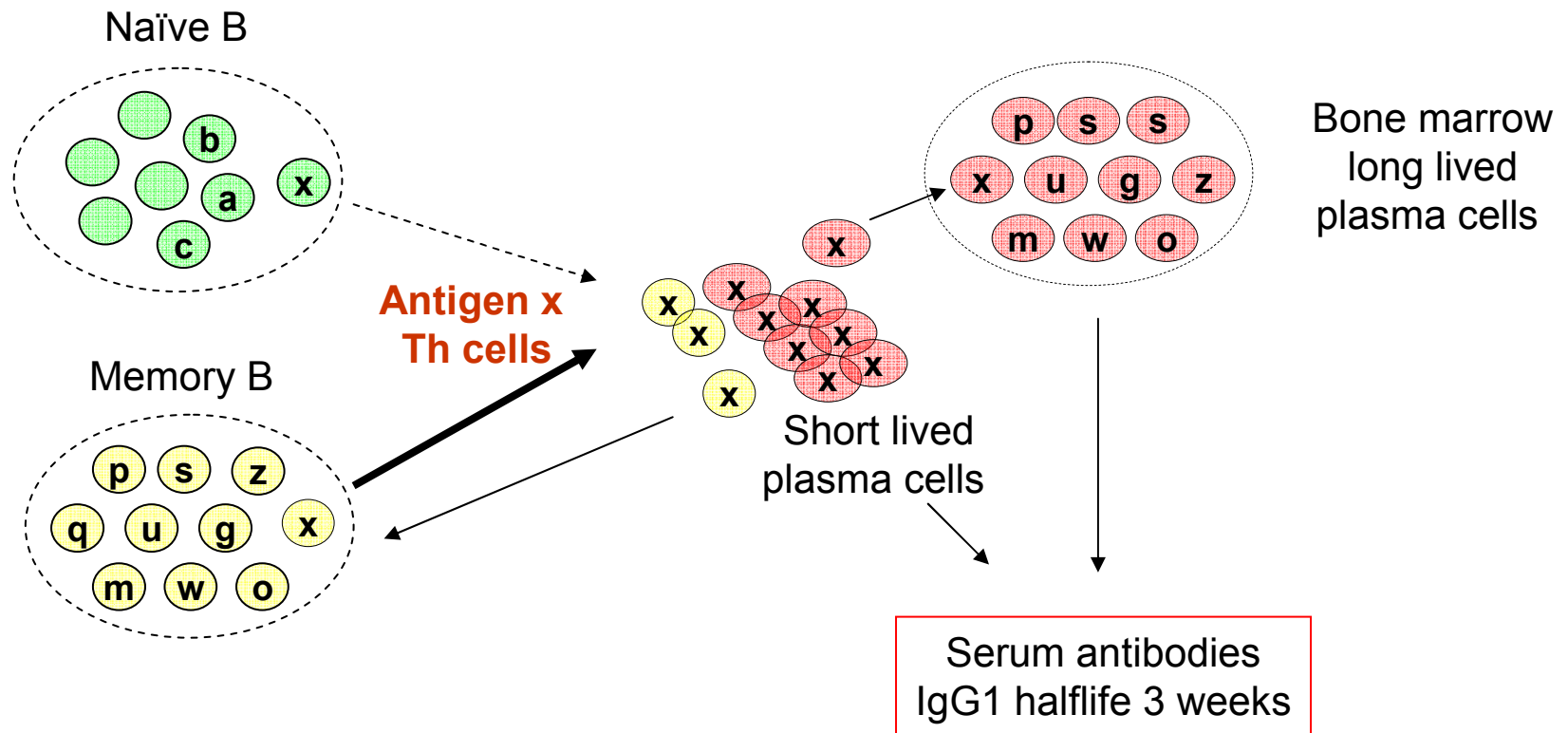
Radbruch et al Nat Rev Immunol, 2006

## B cell clonal dynamics: sustained serum antibodies



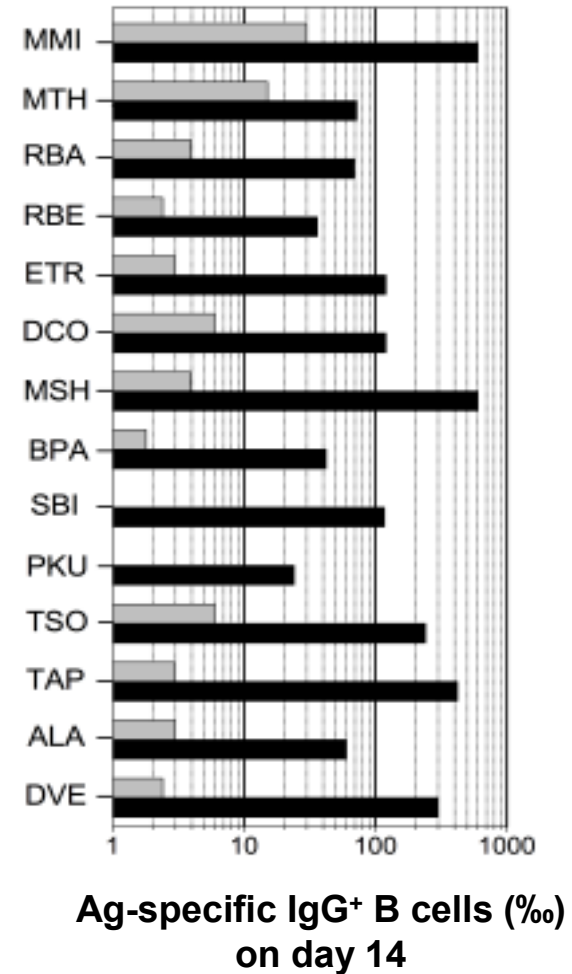
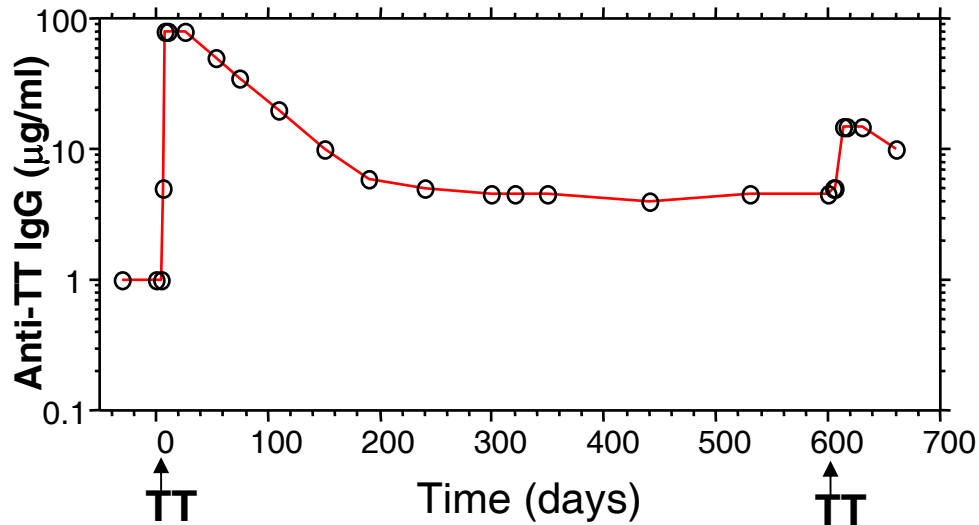
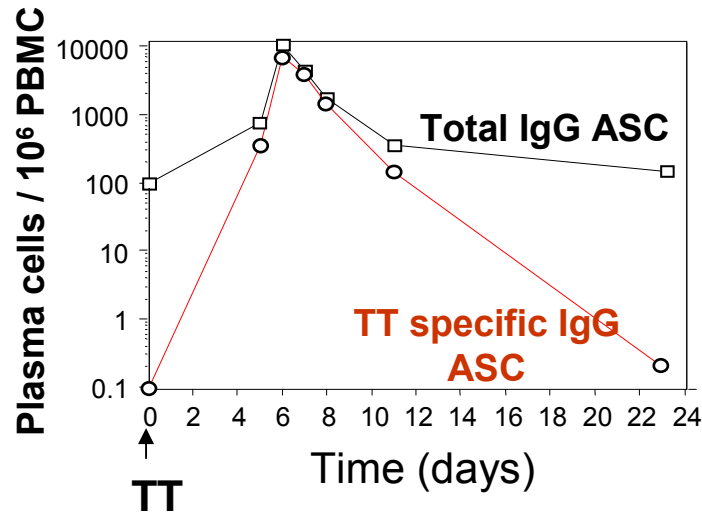
**Serum antibodies to vaccinia virus are maintained constant for a lifetime**

# B cell clonal dynamics: secondary responses



“Original antigenic sin”

# Kinetics of circulating plasma cells, memory B cells and serum antibodies



# Human memory B cell subsets

