

**Combining protein and peptide biomaterials
with laser technologies
towards tissue engineering scaffolds**

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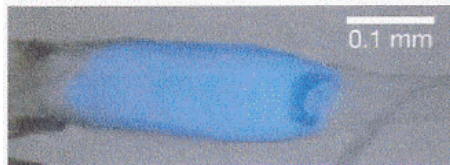
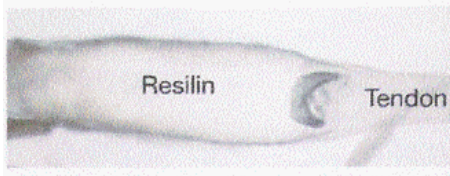




Can we exploit the self-assembly



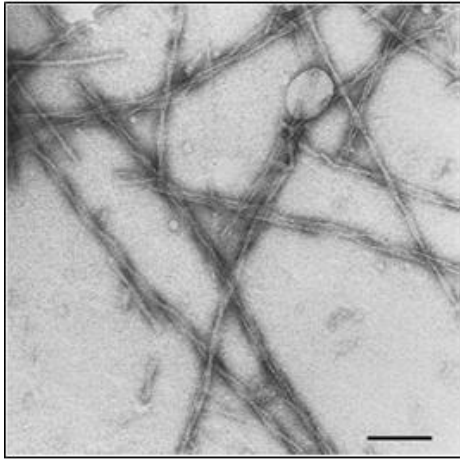
of natural biological materials



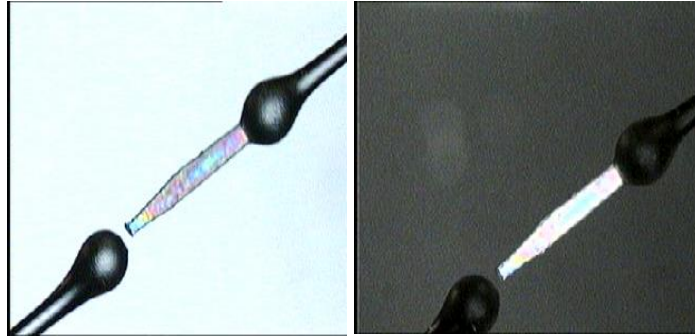
Towards the design of novel biomaterials from the nano- to the macro-scale??



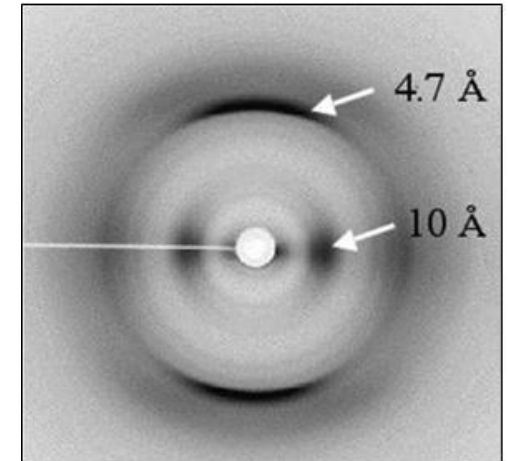
Protein and peptide fragments can self-assemble into “amyloid-type” fibrils



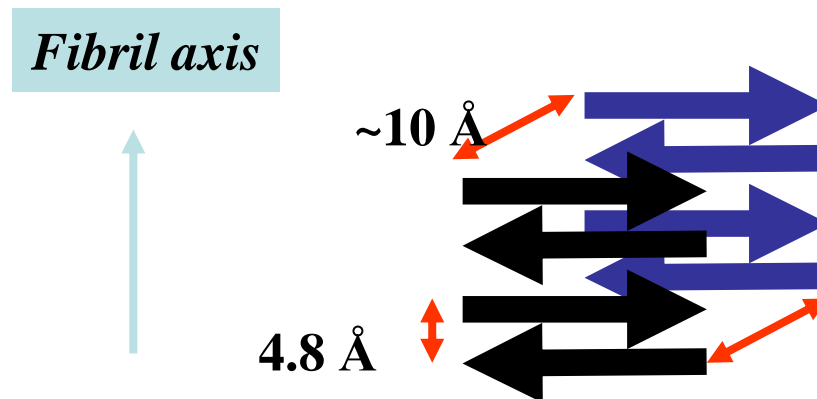
TEM Analysis
(bar:100nm)



Cross-beta model
for amyloid fibrils



X-ray fiber diffraction
pattern for amyloid fibrils



Amyloid fibrous assemblies

- Amyloid assemblies can also be formed by proteins and peptides that are not associated with disease

They are also found in natural fibers / materials (insect chorions, curli fibers, egg stalks, etc)



Chrysopa (Green lacewing fly)
egg stalks (Geddes et al. 1968)

Fibrous nanostructured objects:

**-promising for potential integration
in future generations of nano-micro devices**

-carbon nanotubes most investigated

**-however, limitations:
(high production costs, extreme manufacturing
conditions)**

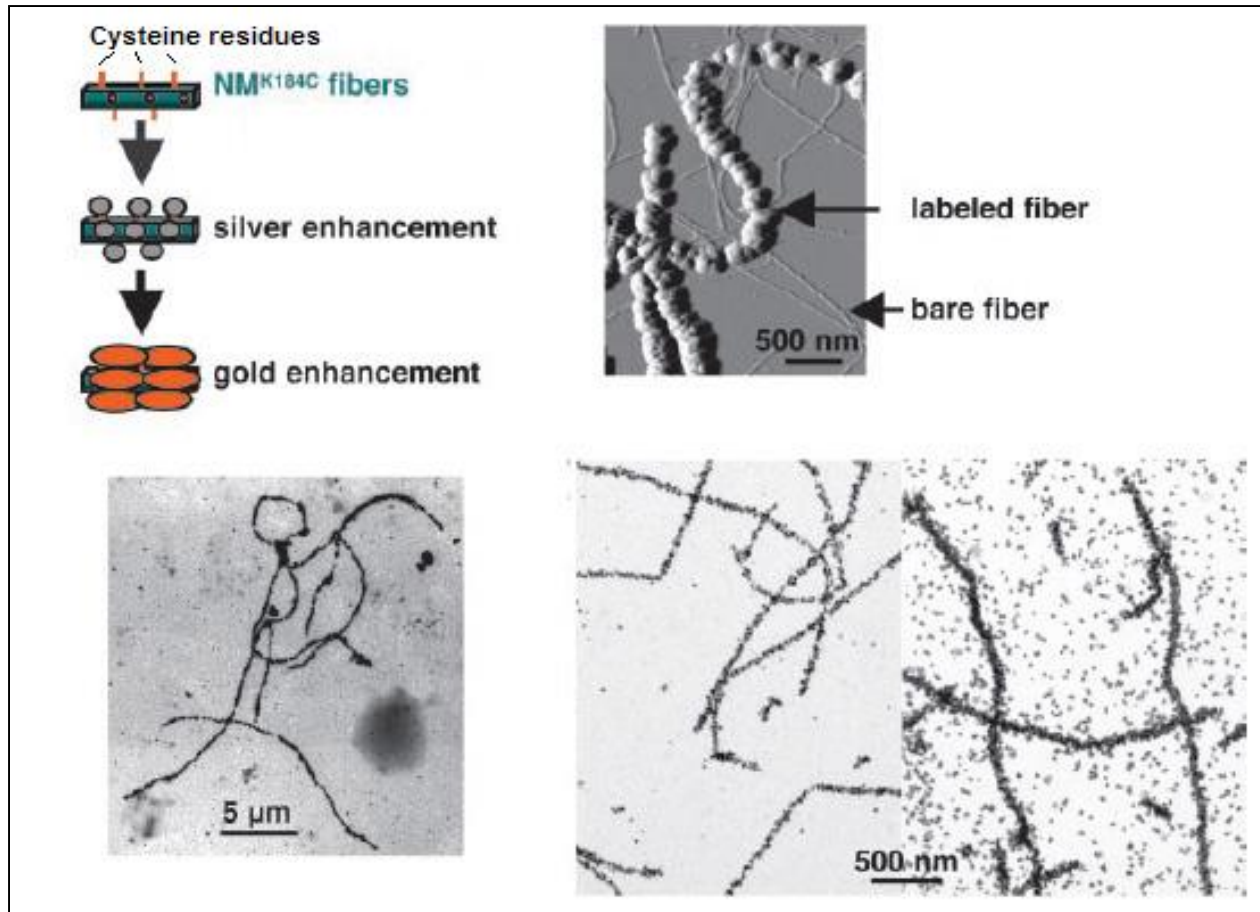
Advantages of biological counterparts (DNA, peptides, proteins):

-Spontaneous self-assembly in mild conditions

**-ease of functionalization
(possibility of modifications *at the sequence level*)**

Self-Assembling peptides as templates for technological applications

A fragment from the yeast *Saccharomyces cerevisiae* Sup35 protein that forms amyloid fibrils was used for nanowire fabrication



Susan Lindquist group

*Scheibel et al.,
PNAS 2003, 100: 4527*

Natural assemblies as a source of inspiration for technological applications

Natural assemblies can inspire

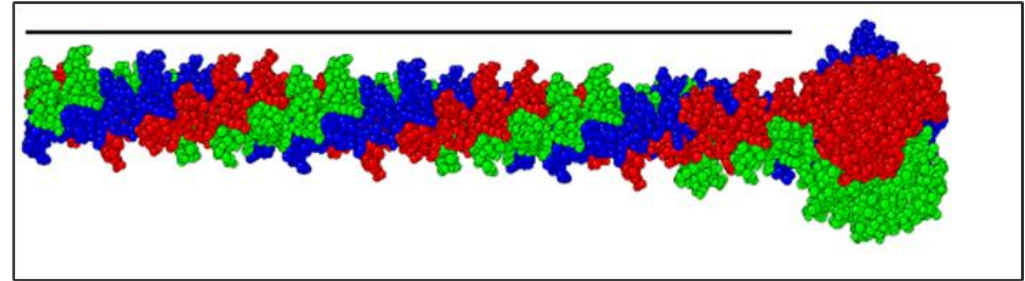
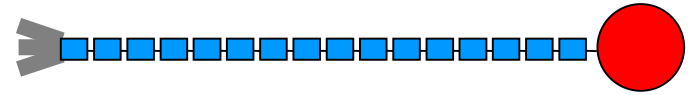
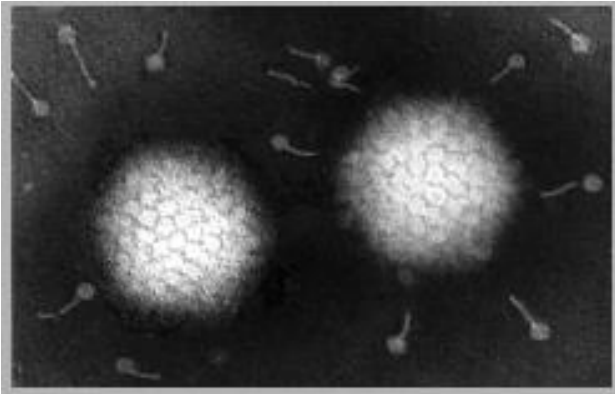
the specific design of

biological building blocks in order to

fabricate supramolecular structures

with advanced functionalities

Adenovirus fiber as a self-assembly system



Model for the full-length fiber based on the crystal structure of a stable fragment with 4 repeats

[van Raaij et al. Nature 1999, 401:935]

Properties of adenovirus fiber:

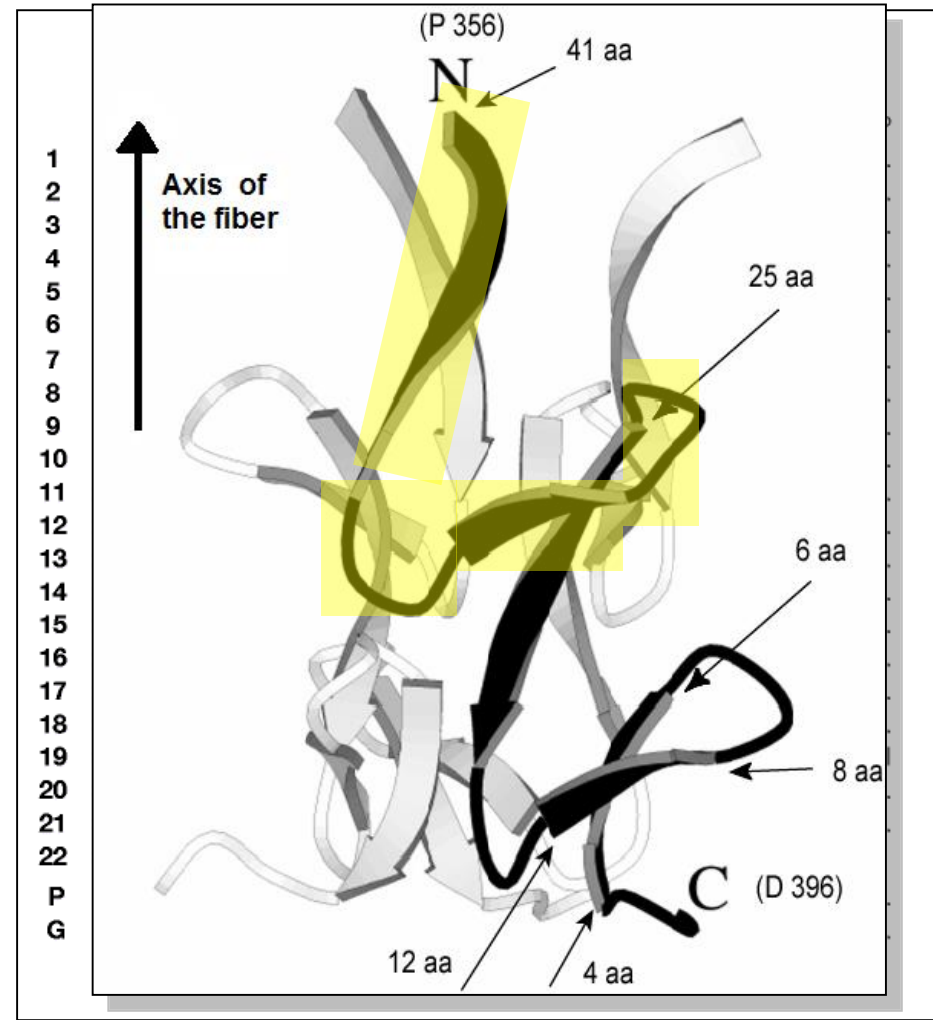
- 1. Long and thin shaft (30 nm) gives the virus 'reach'**
- 2. Trimeric, beta-structured**
- 3. Extremely stable proteins: resistant to SDS, temperature, denaturants**
- 4. Fibrous shaft built from sequence repeats (blue squares)**

Adenovirus fiber as a self-assembly system

Each fiber monomer contains 582aa

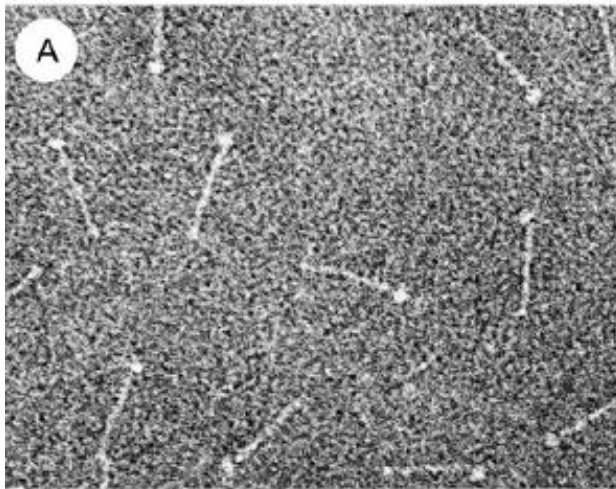
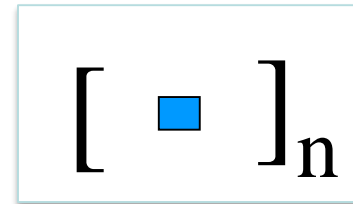
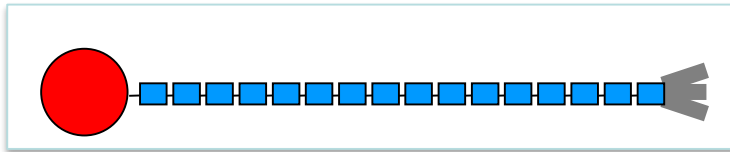
1. The fibrous shaft is built from a repeating sequence motif with a **hydrophobic** aminoacids (**ORANGE** and **GREEN** residues) alternating with **hydrophilic** ones and glycine or proline at conserved positions (**PURPLE** residues)

2. The basic repeat fold contains a beta-strand almost parallel to the axis of the fiber followed by a beta turn and another beta strand which runs at an angle of 45° relative to the fiber axis. The repeats are joined by a solvent exposed loop

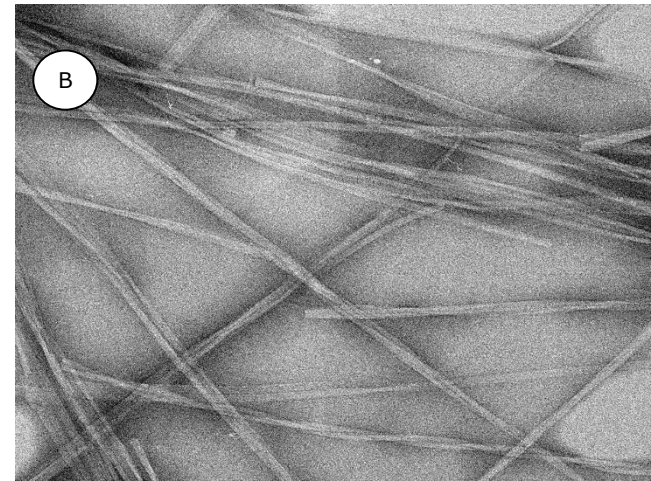


Peptide sequences as a self-assembly system

Identification of minimal peptides corresponding to natural building blocks (blue squares, octapeptide NSGAIIG) that can self-assemble into amyloid-type fibrils



50 nm

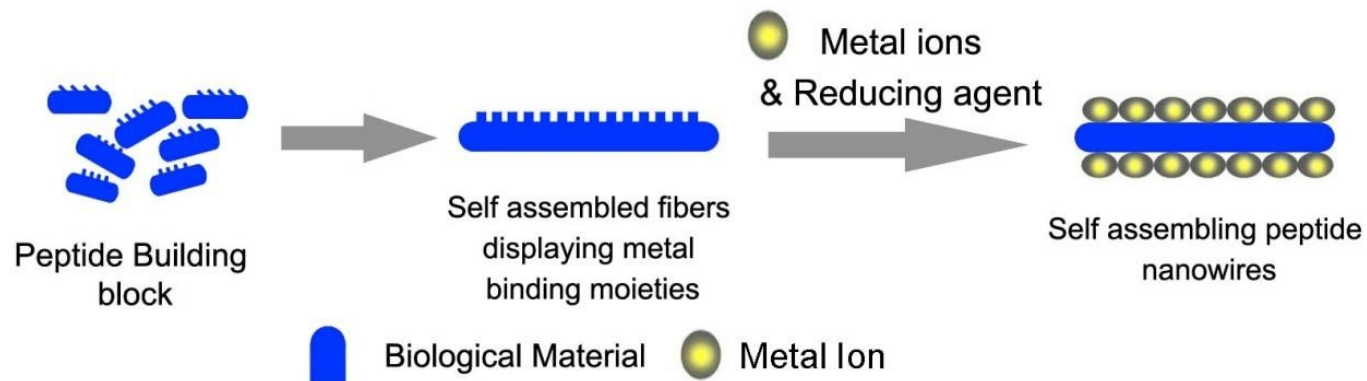


100 nm

Can we rationally modify these fibrous building blocks towards specific functions?

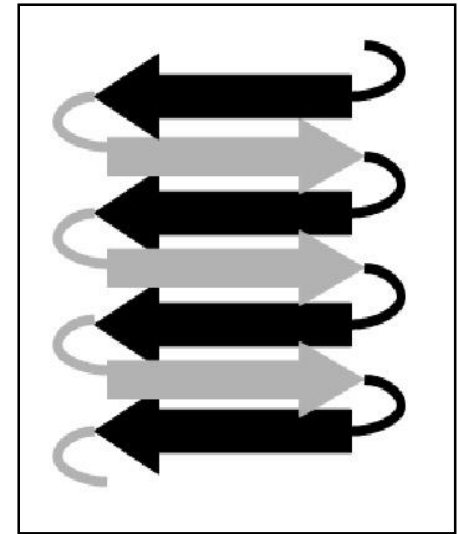
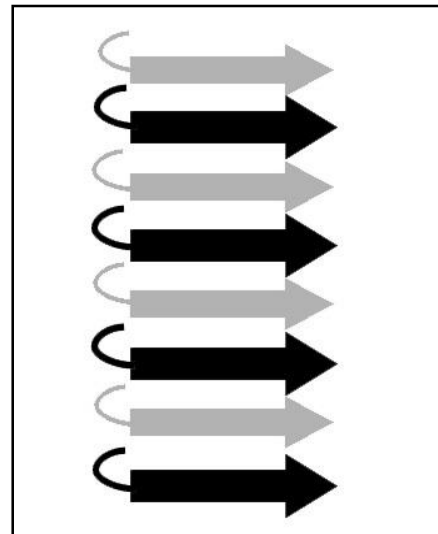
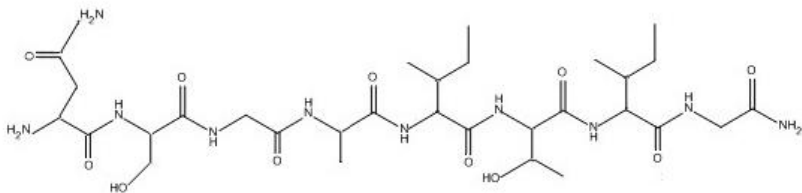
eg. templating of inorganic materials

**targeting sequences for metallization,
Calcium binding,
cell attachment etc**



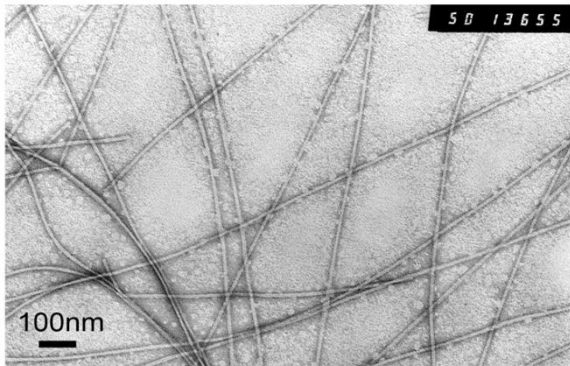
- The N-S-G-A-I-T-I-G peptide self-assembles into amyloid fibrils with cross-beta structure
- The beta strands are perpendicular to the fibril axis
- The peptide arrangement can be parallel or anti-parallel
- Molecular Dynamics simulations suggest that the N-S residues stay exposed out of the amyloid fibril core

(*Tamamis et al., J. Phys. Chem. B 2009, 113:15639*)

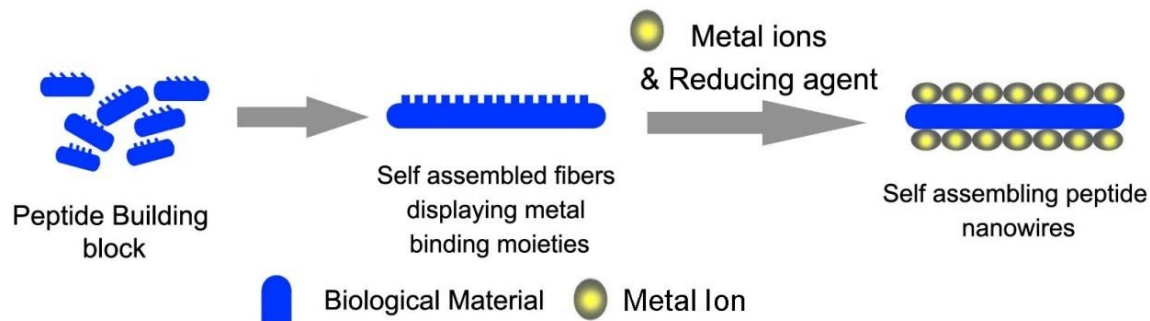
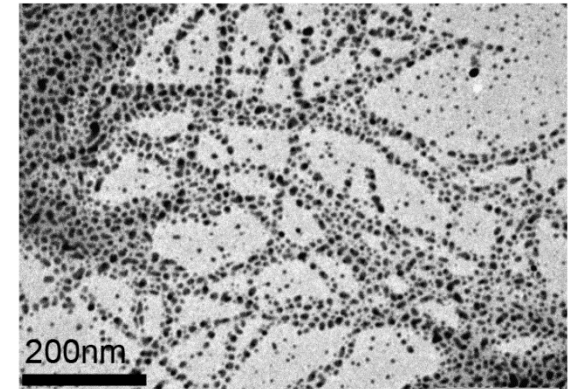


Rational modification of these building blocks towards specific functions

eg. templating of inorganic materials
(metal nanoparticles, silica, Qdots, calcium)
cell attachment *etc*



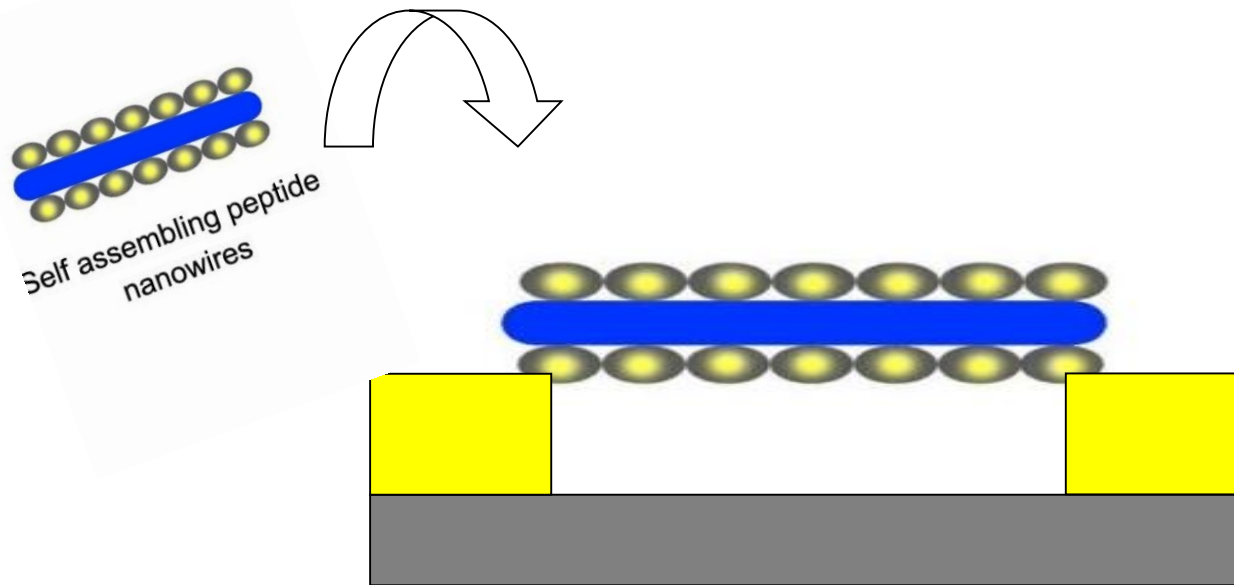
- 1) Kasotakis et al., 2009, *Biopolymers* 92:164-72
- 2) Kasotakis, Mitraki, 2012, *Biopolymers*, 98:501
- 3) Kasotakis et al., 2014, *App. Phys.A.*, 116:977



Can we control the positioning of fibers accurately, in 2- or 3-dimensions at the micro / nano scale???

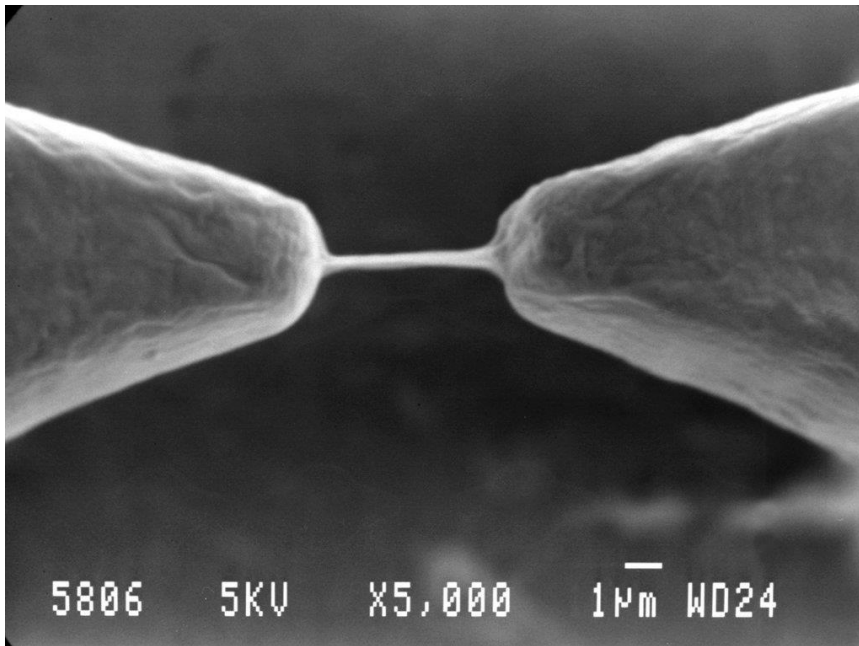
Directed positioning is a major challenge

For integration of self-assembling fibrous materials in devices



Development of inter-disciplinary approaches

**towards controlled positioning on surfaces
and integration in devices**



Example: peptide
fibrils positioned
on a hybrid
organic-inorganic
material
using femtosecond
Laser
Technologies

V. Dinca et al., NanoLetters 2008, 8:538

(collaboration with Dr. M. Farsari et al., IESL, FORTH)

Direct Laser Writing (DLW) Multi-photon polymerization technique (MPP)

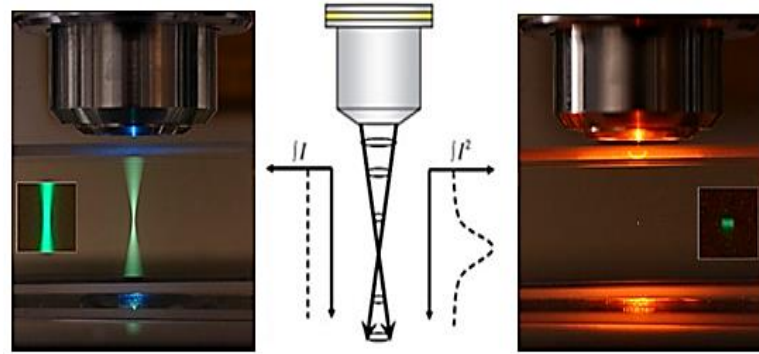
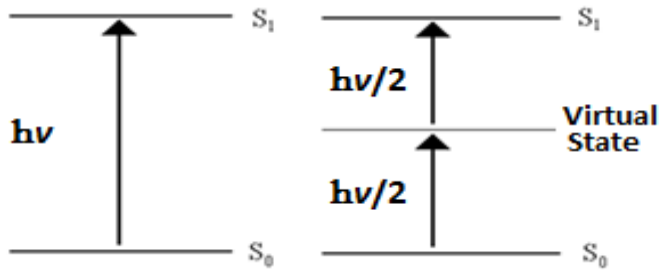
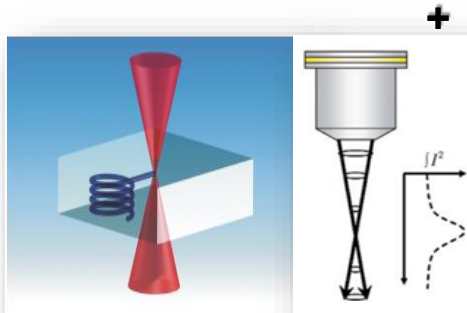
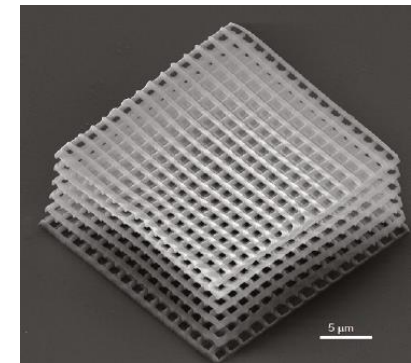


Image by Steve Ruzin and Holly Aaron, UC Berkeley

Hybrid Photostructurable Material



→
NIR fs pulses



- ❖ *Highly attractive and promising 3D microfabrication technology*
- ❖ *Real 3D Writing (no mask, mold or stamp)*
- ❖ *Complex Micro- & Nano- structures*

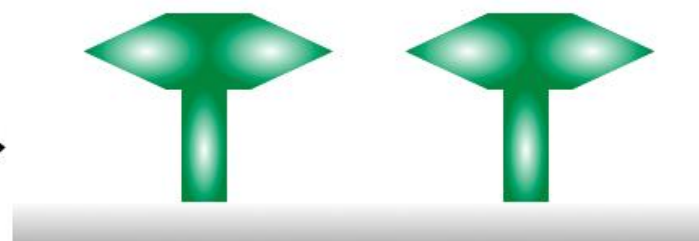
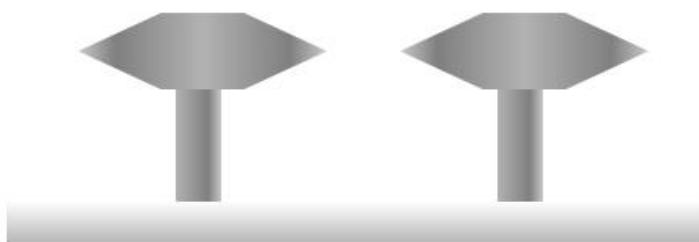
Sakellari et. al, 2012, ACSNano, 3: 2302

Positioning of metal-binding peptides on 3D structures fabricated using a composite sol-gel with metal binding sites

Metallization – Peptide deposition process:

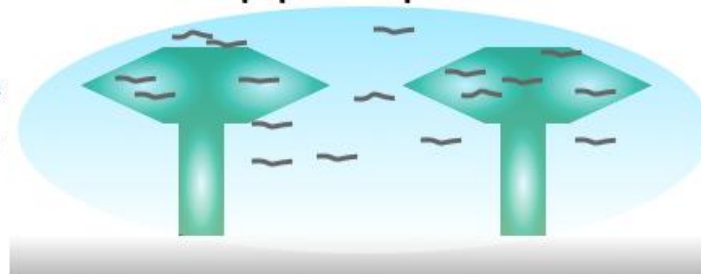
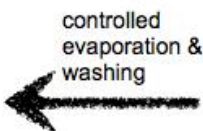
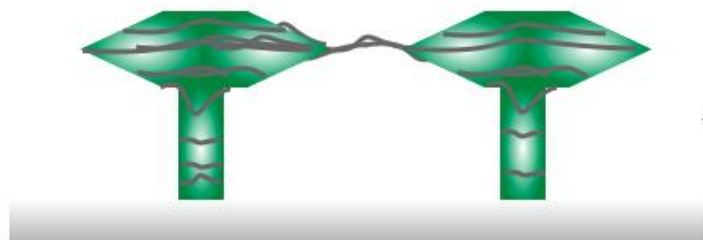
• 3D structure fabrication using 2PP

• H_{Au}Cl₄ seeding + reduction (sodium citrate or NaBH₄)



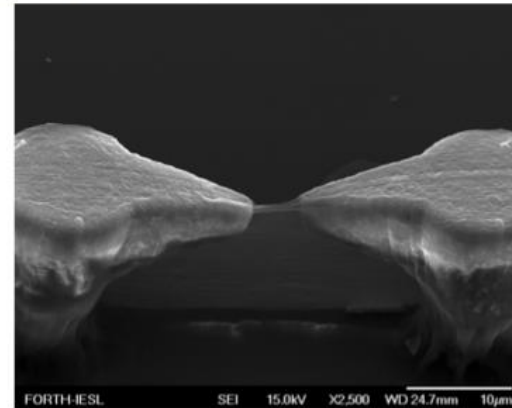
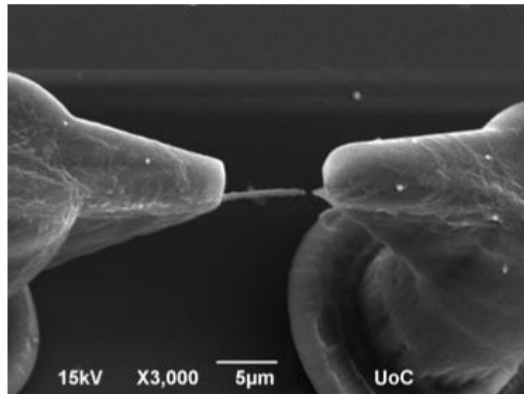
• formation of bridges

• peptide deposition

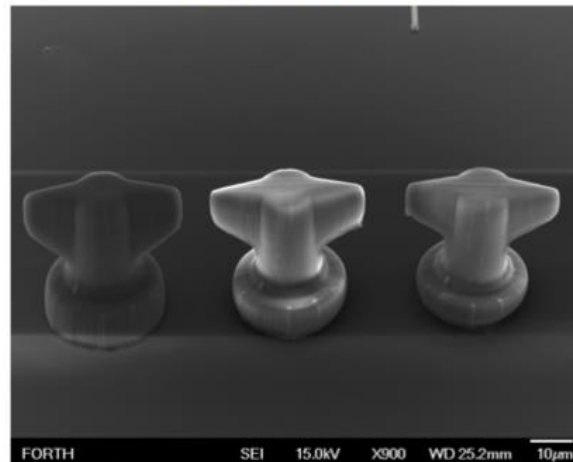


Positioning of Cysteine-containing peptides on 3D nanostructures fabricated using a composite sol-gel with metal binding sites

Cysteine containing peptide



control peptide



***Can we develop « scaffold-on-scaffold »
approaches?***

ie.

**Combine the advantages of top-down fabrication
(eg. control of scaffold geometry and micro-nanotopography)**

With

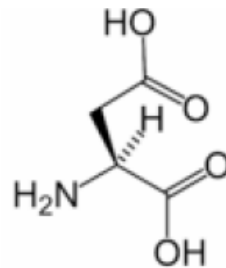
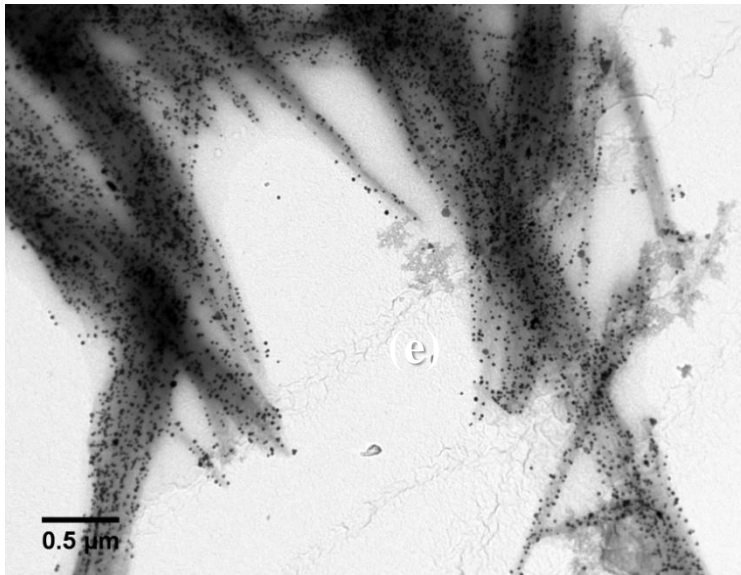
**Bottom-up design possibilities of
Self-assembling peptides?**

Design of bifunctional aspartate-containing self-assembled peptides

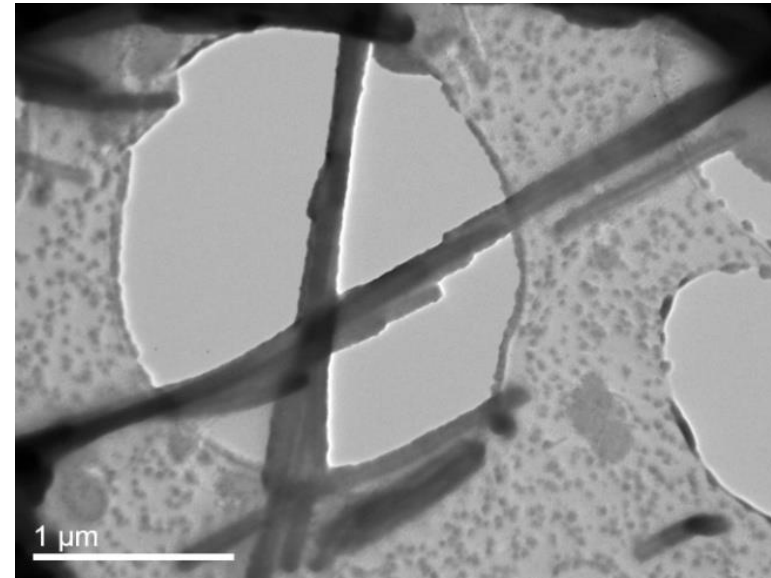
Peptide DDSGAITIG forms fibrils that:

a) Bind to metal nanoparticles

b) nucleate calcium phosphates on their surface (important for bone and teeth regeneration)



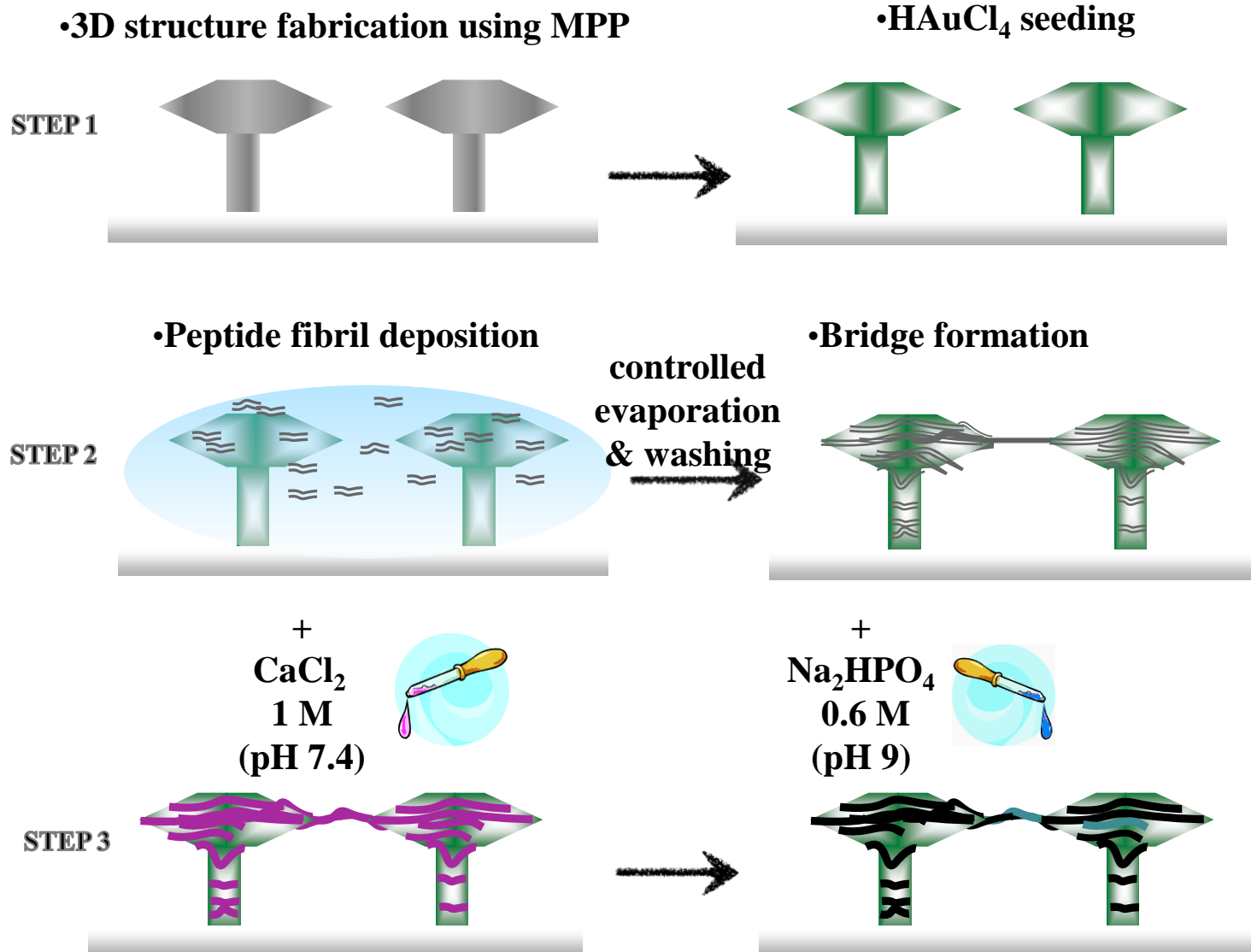
L-Aspartic acid
(Asp / D)



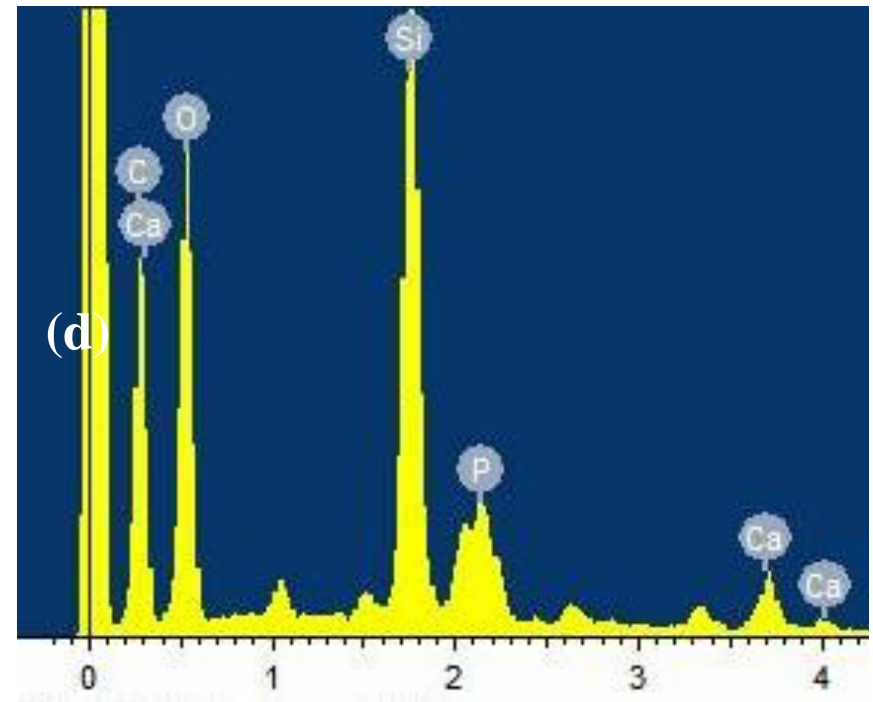
a) DDSGAITIG peptide fibrils after incubation with gold tetrachloroaurate solution and reduction with 1% citric acid

b) DDSGAITIG peptide fibrils after incubation with CaCl_2 and Na_2HPO_4 on a holey grid

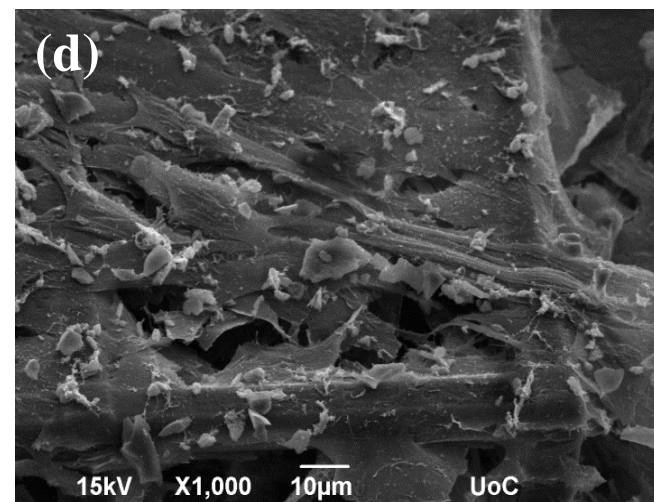
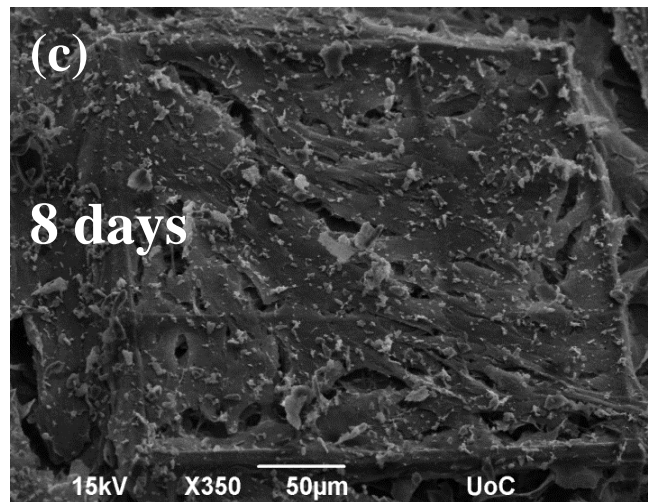
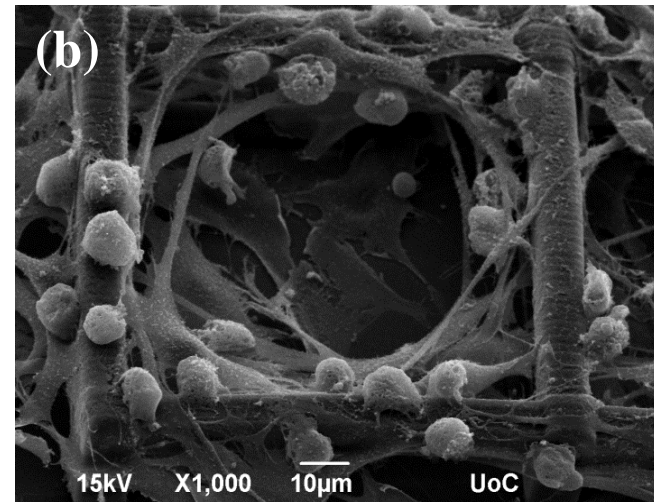
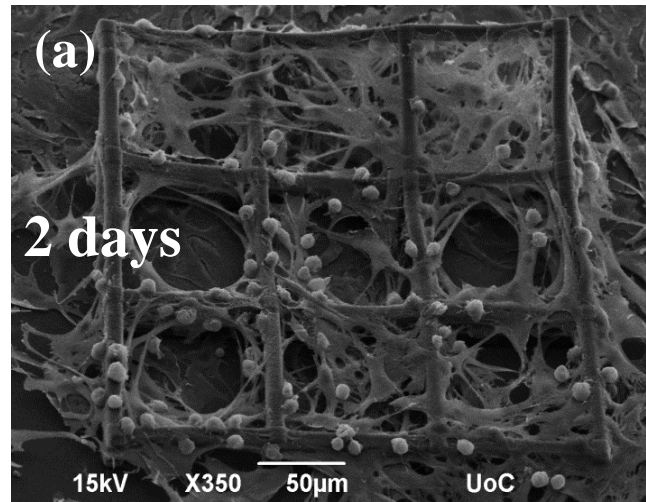
Development of a “Scaffold-on-Scaffold” strategy



**Peptide bridges positioned on the hybrid scaffold,
covered with calcium phosphates
And EDX pattern on the bridge**

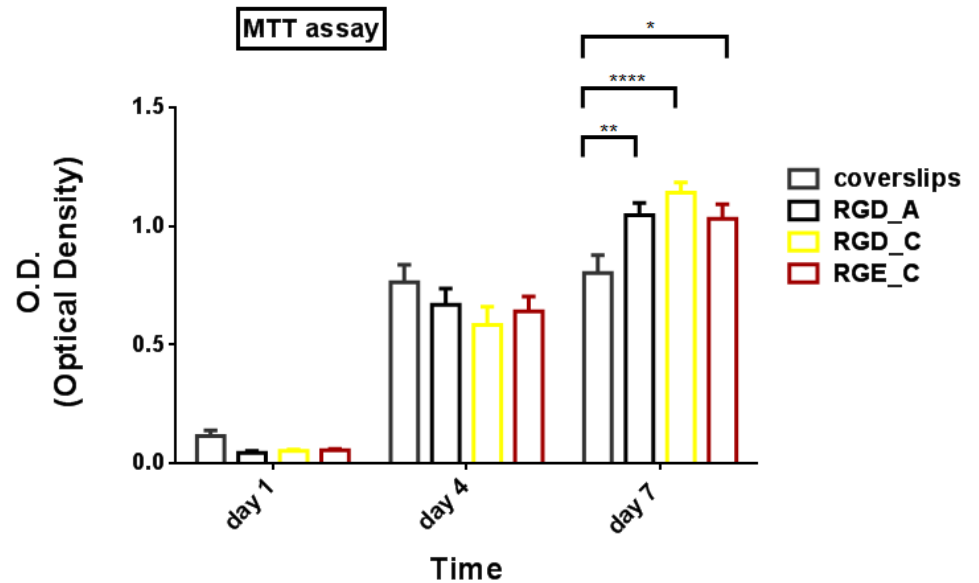
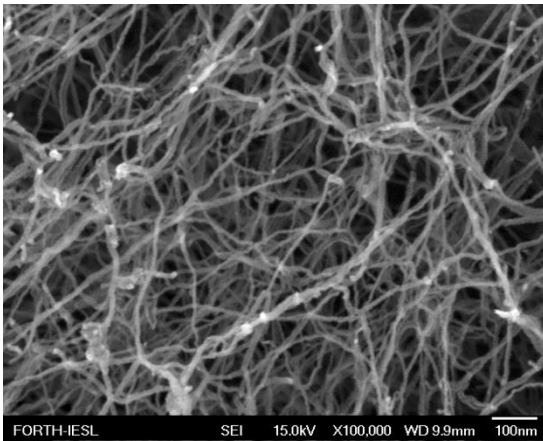


**SEM images of pre-osteoblastic cells cultured
onto DDSGAITIG peptide-functionalized
and Calcium mineralized 3D woodpile-shaped scaffolds**



Designer peptides containing cell attachment (RGD) motifs as scaffolds for tissue engineering: theoretical and experimental studies

RGD-SGAITIG-C self-assembling peptides



Loo et al., Adv. Healthcare Mat., 2015, 16: 1557

Deidda et al., ACS Biomat. Sci. and Eng. 2017, 7: 1404

Jonnalaggada et al., Mol. Syst. Design and Eng. 2017, 2: 321

Can we produce self-assembling fibrous materials in the macro-scale, using microbial cell factories (eg. recombinant production in bacteria)?

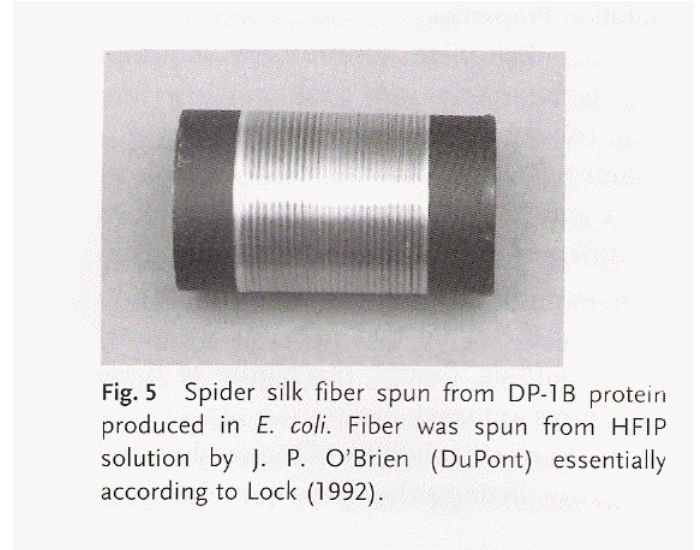
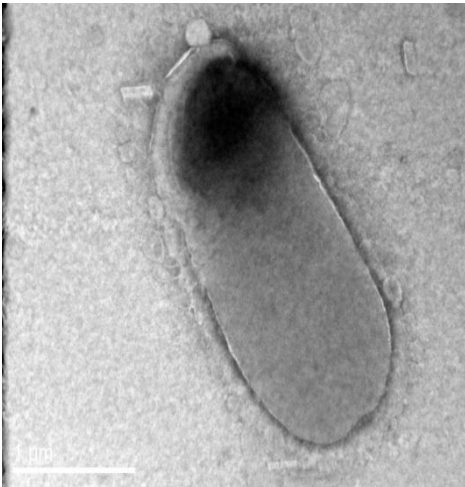


Fig. 5 Spider silk fiber spun from DP-1B protein produced in *E. coli*. Fiber was spun from HFIP solution by J. P. O'Brien (DuPont) essentially according to Lock (1992).

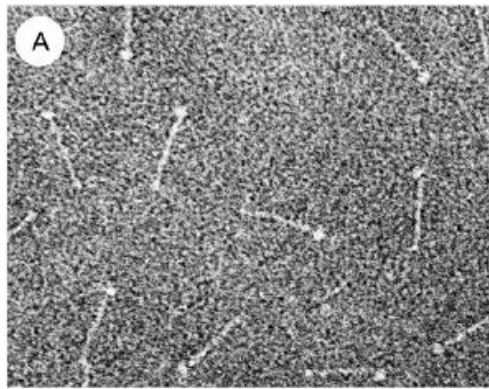
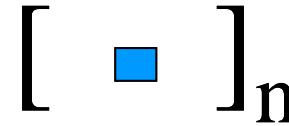
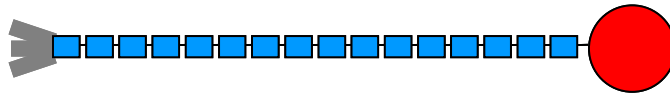


“RECOMBINAMER”

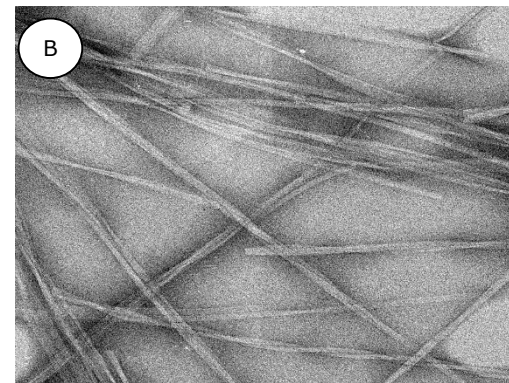
Concatamers, based on the minimal self-assembling building blocks

production in quantities will allow to use electrospinning and other methods used mainly for polymers

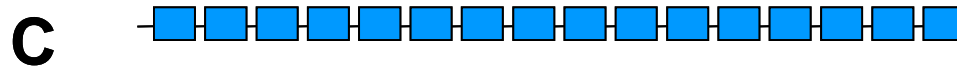
Next important challenge: recombinant production of self-assembling fibrous materials



— 50 nm

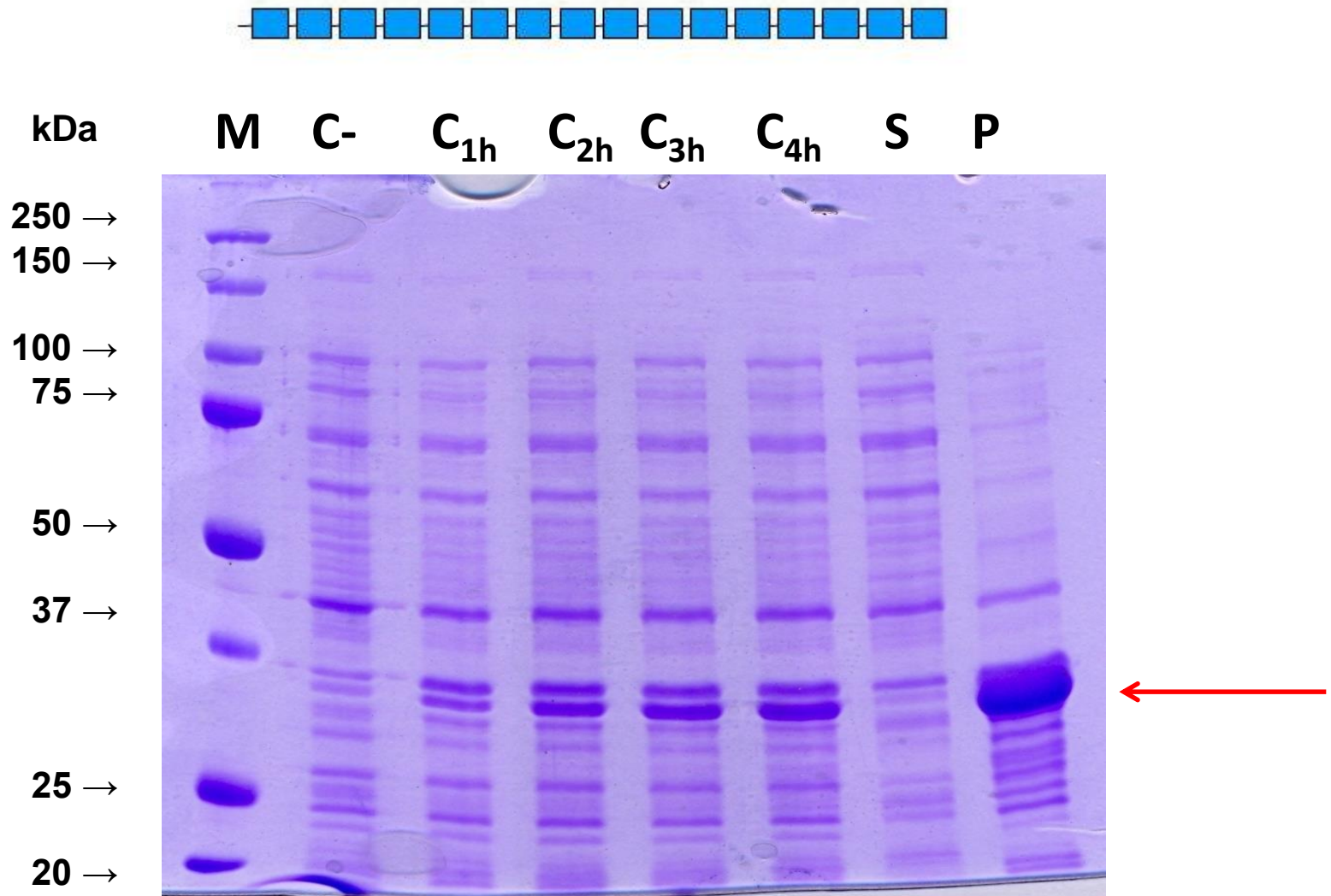


— 100 nm



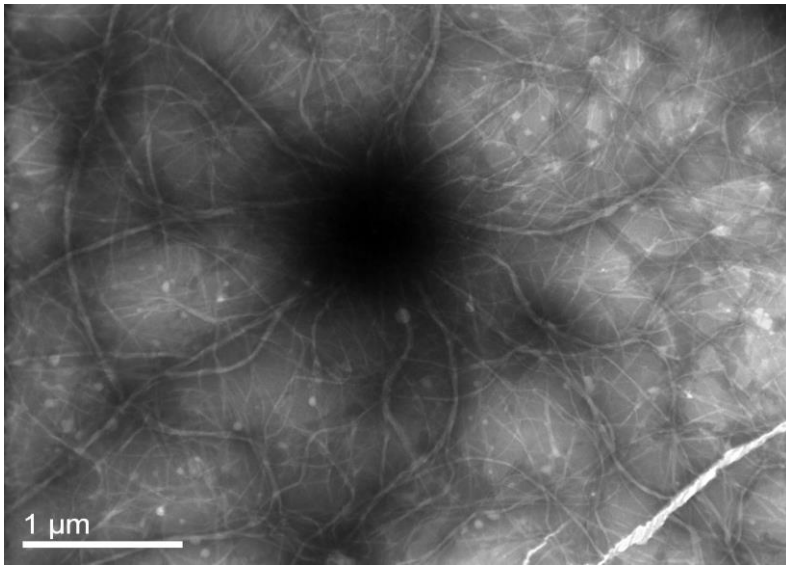
- 1) *Proteins composed of various lengths of the natural shaft*
- 2) *Concatamers based on the minimal self-assembling building blocks*

Expression of the full-length shaft in *E.coli* (residues 61-392)

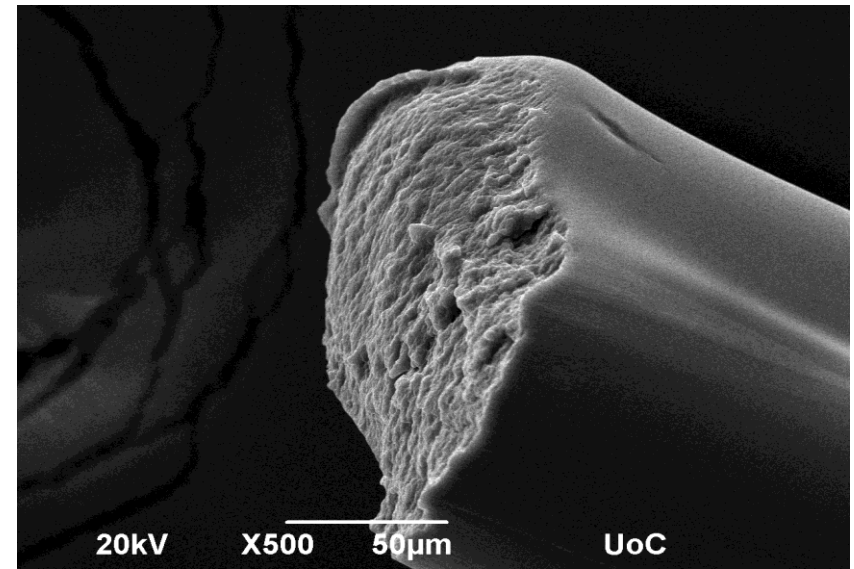


Lane M: molecular mass marker, **Lane C-:** non induced bacteria, **Lane C_{1h}:** bacteria after 1h of induction, **Lane C_{2h}:** bacteria after 2h of induction, **Lane C_{3h}:** bacteria after 3h of induction, **Lane C_{4h}:** bacteria after 4h of induction, **Lane S:** Supernatant of lysates, **Lane P:** pellet of lysates.

Processing of fibers from the purified proteins that self-assemble into amyloid-type fibrils



Protein is produced in form of “Inclusion bodies” consisting of amyloid-type fibrils



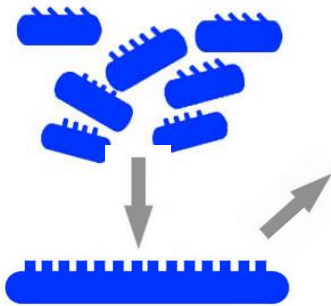
Fiber rod drawn from purified protein



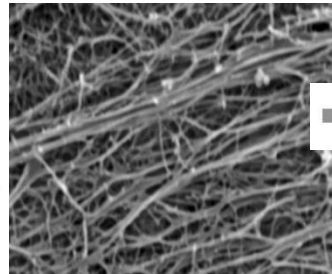
Protein fiber drawn between two glass rods, viewed under crossed polars

Ariadni Prigipaki, PhD work

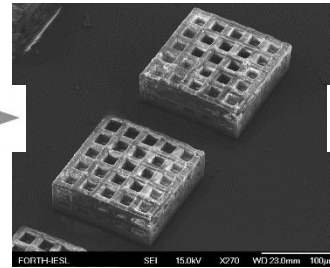
Can we photostructure protein and peptide-only materials and use them as scaffolds?



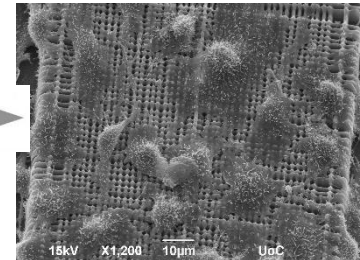
Peptide and protein
Building blocks



self-assembled
matrices



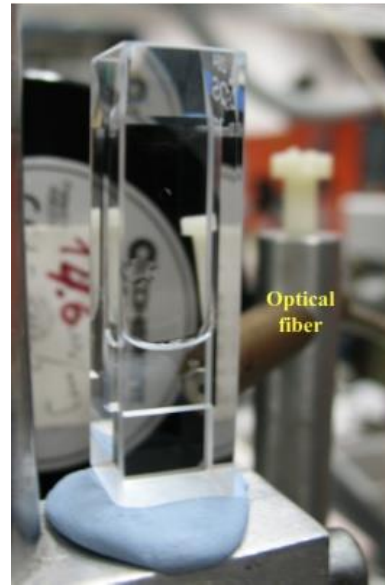
3D photofabrication



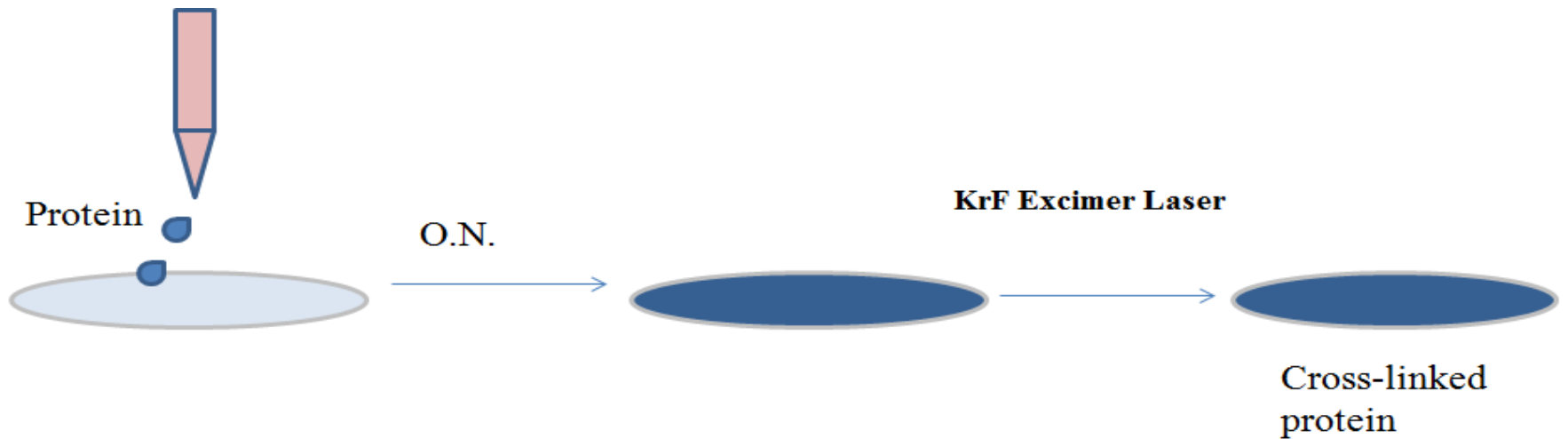
cell attachment
on scaffolds

(collaboration with Dr. A. Selimis and Dr. A. Ranella, IESL, FORTH)

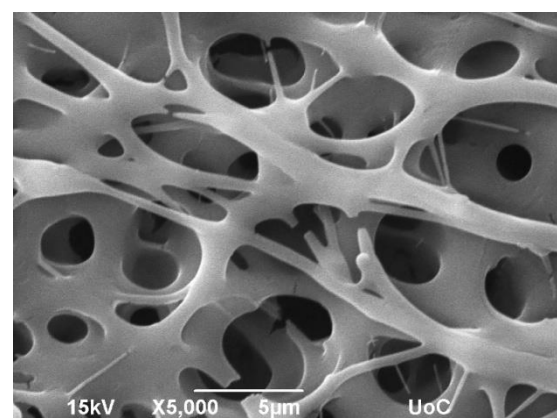
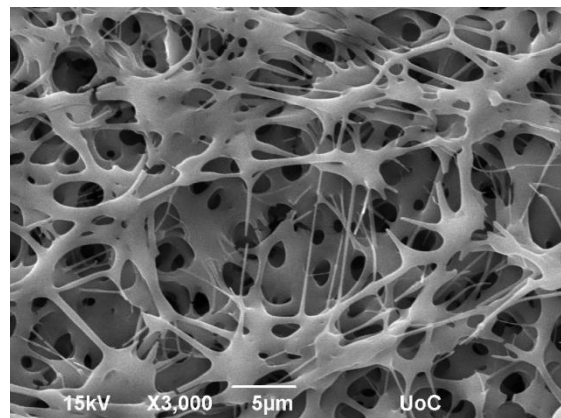
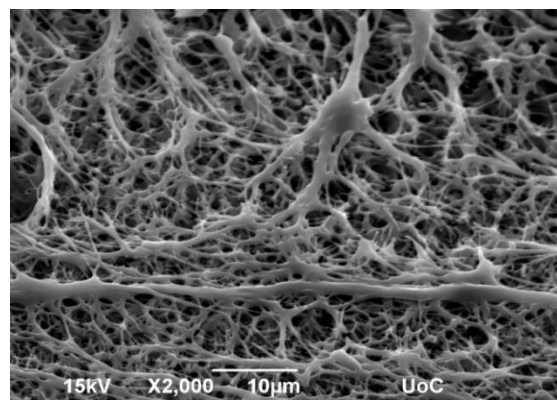
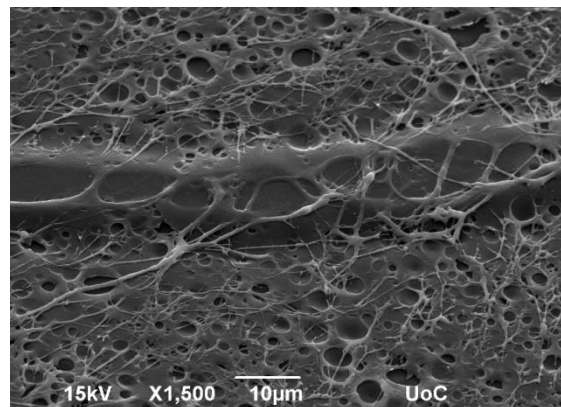
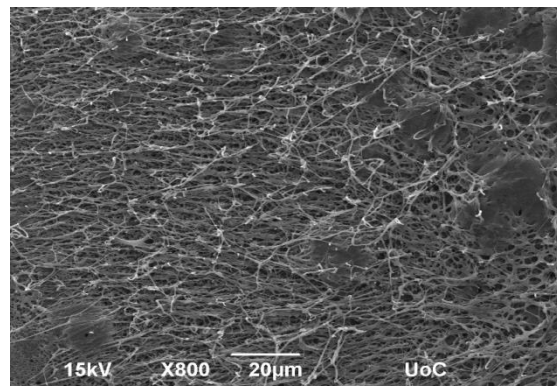
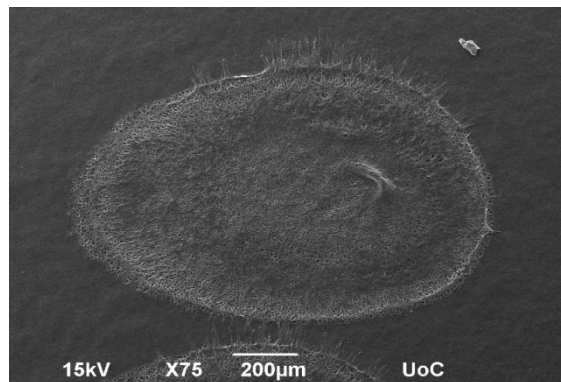
Cuvette Preparation



Film Preparation

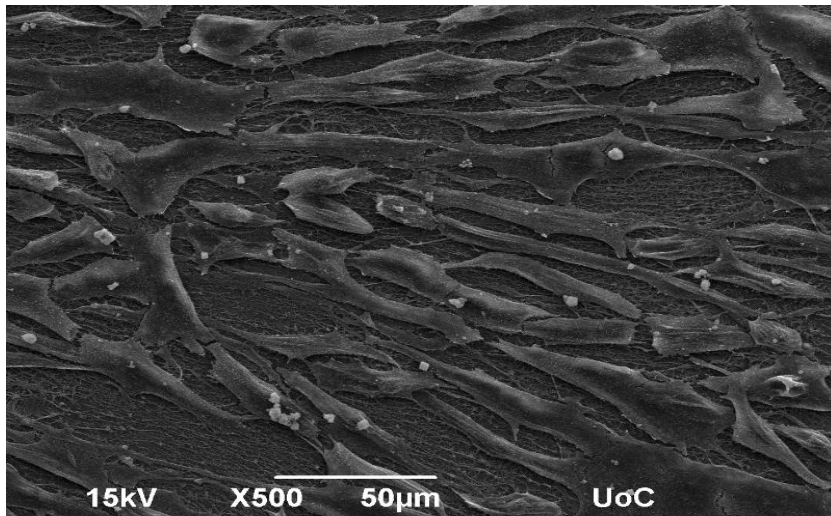
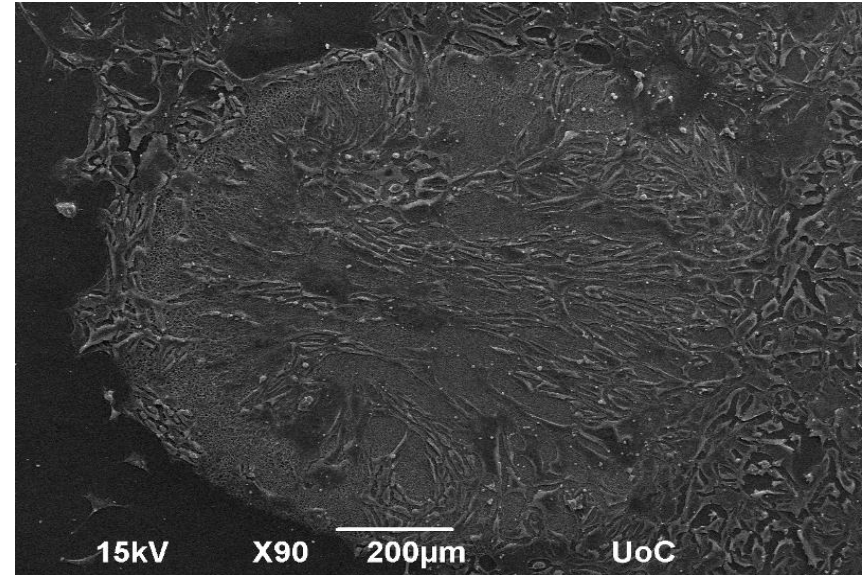
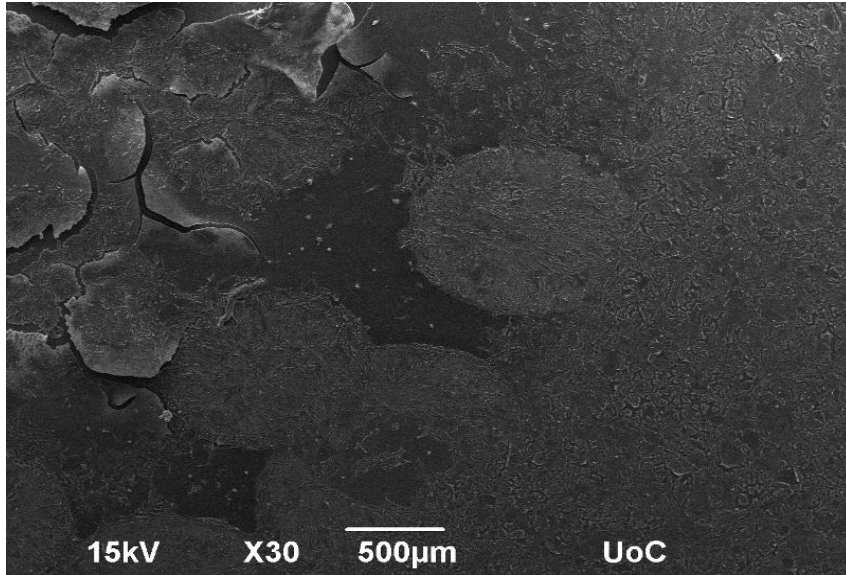


Irradiated area 300
mJ/cm²



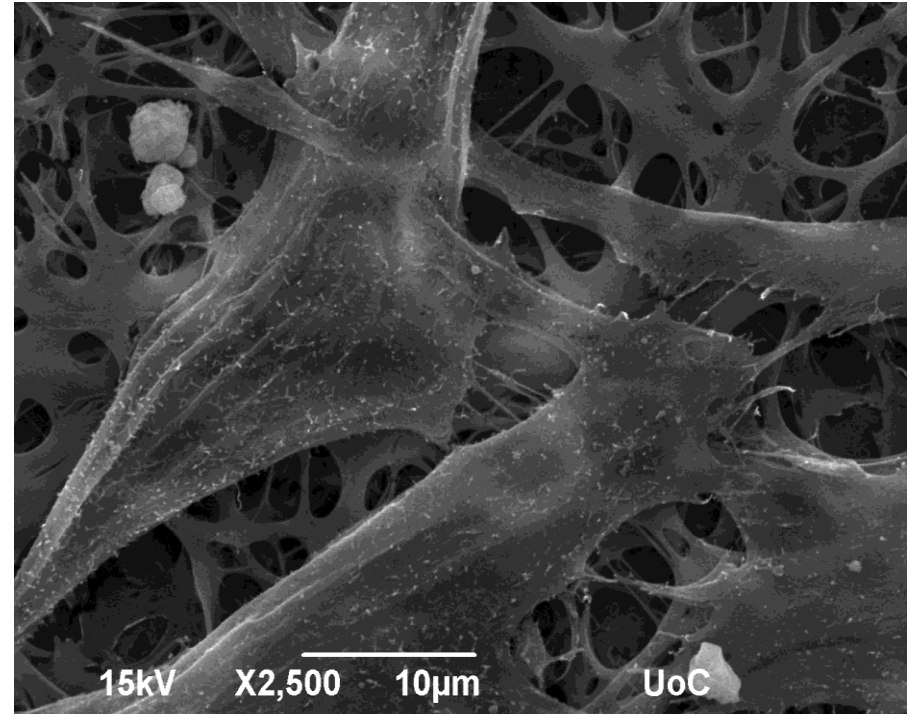
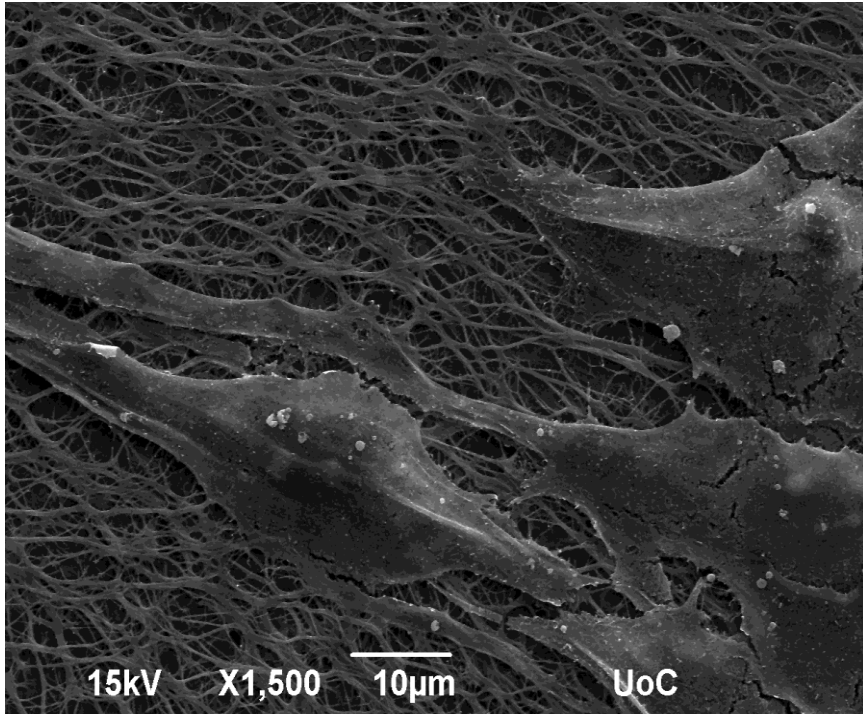
protein (2.5mg/ml)

Protein Scaffolds with Fibroblasts



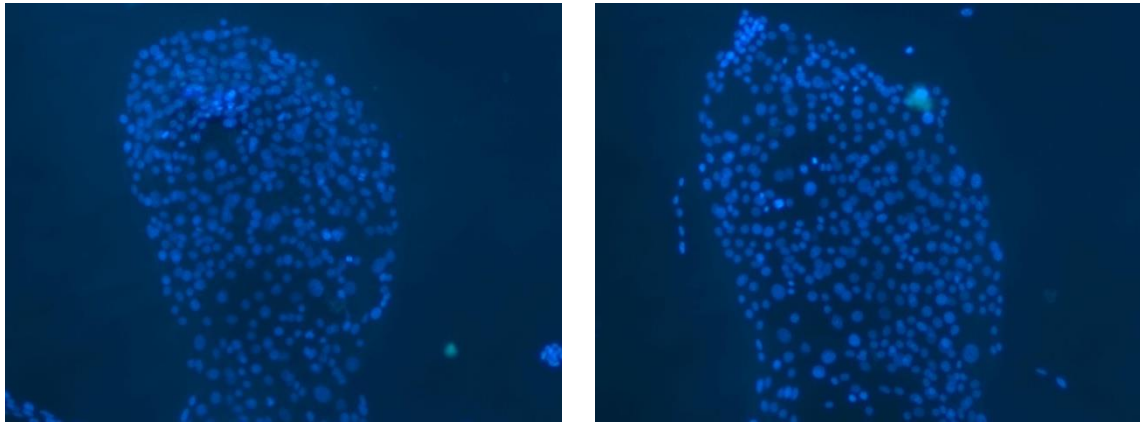
SEM image of the irradiated area of the protein film with one pulse (30ns) of the KrF excimer laser (248nm) at a fluence of 300 mJ/cm² (2.5mg/ml of protein - 40μl deposited, o/n drying at RT), covered with fibroblast cells (NIH/3T3 cells, 10⁵ cells/ml, 72h of culture).

Protein Scaffolds with Fibroblasts

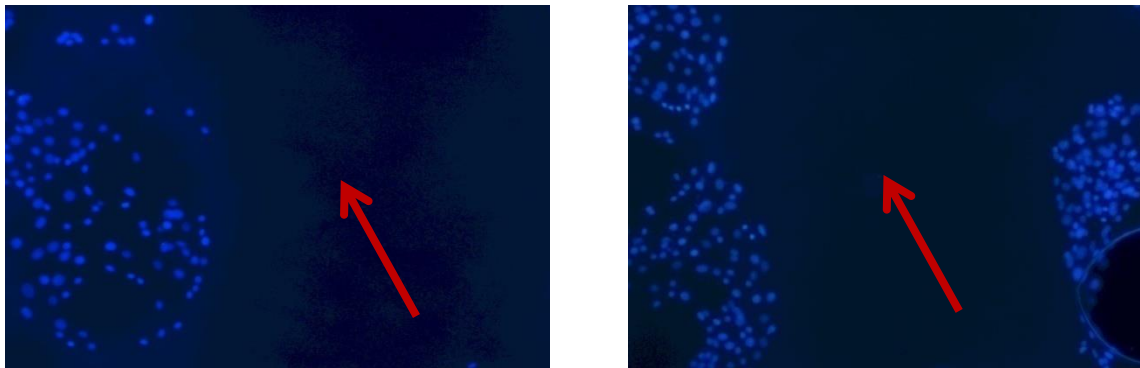


Cells attach selectively to the irradiated areas

Protein Scaffolds with Fibroblasts - UV Microscope DAPI DAY 5



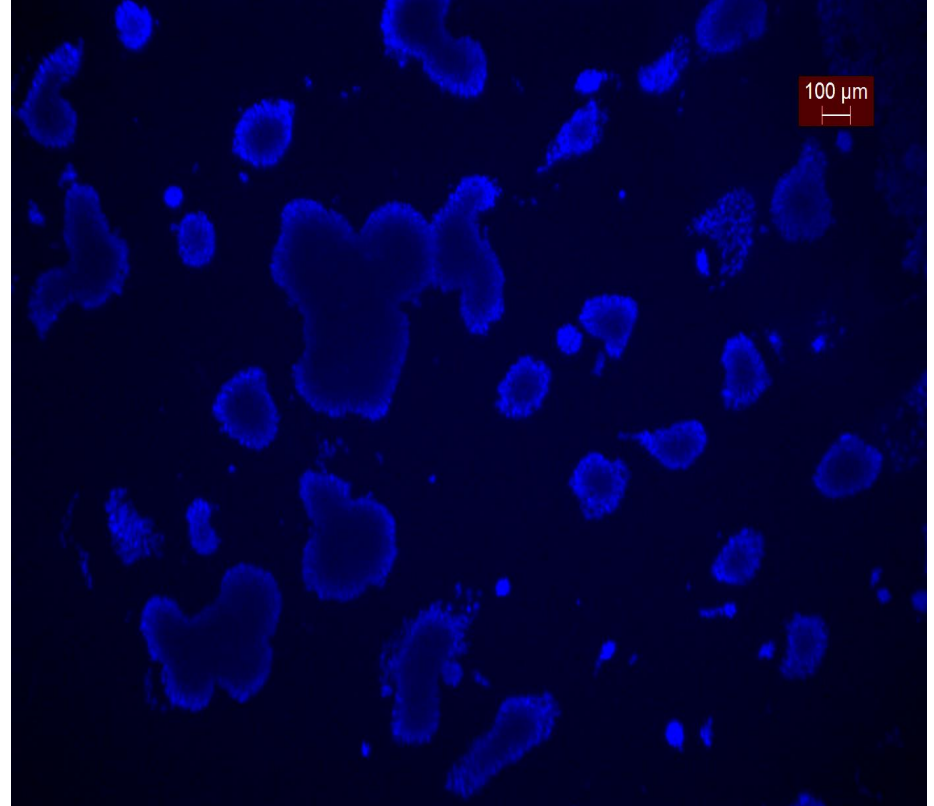
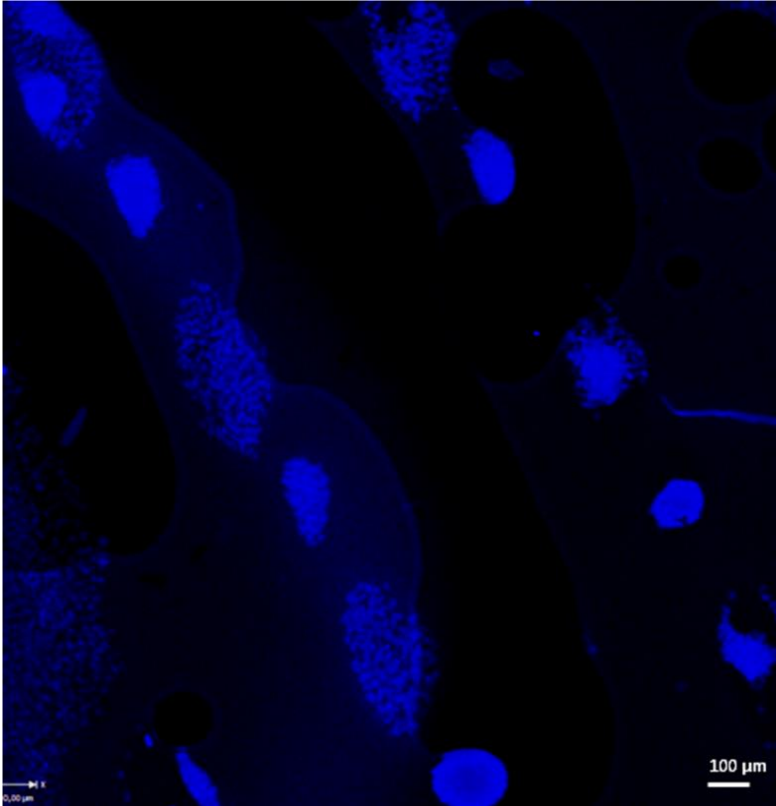
irradiated area



Non - irradiated area (red arrow)

Protein Scaffolds with Fibroblasts

Selective Cell Patterning



Fluorescence microscopy images of fibroblasts on laser patterned surfaces after 3 days of culture-DAPI staining.

Prigipaki et al., Biofabrication, 2017: 9, 045004

Summary:

***Rational design of bio-nanomaterials
from the nano- to the macroscopic scale***

***using building blocks inspired from
Natural folds and hard-won biochemical knowledge***

and

Development of inter-disciplinary approaches

***Towards combining these bottom-up design
possibilities***

With top-down biofabrication technologies

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